

MEDICARE COVERAGE OF LABORATORY TESTING

Please remember when ordering laboratory tests that are billed to Medicare/Medicaid or other federally funded programs, the following requirements apply:

1. Only tests that are medically necessary for the diagnosis or treatment of the patient should be ordered. Medicare does not pay for screening tests except for certain specifically approved procedures and may not pay for non-FDA approved tests or those tests considered experimental.
2. If there is reason to believe that Medicare will not pay for a test, the patient should be informed. The patient should then sign an Advance Beneficiary Notice (ABN) to indicate that he or she is responsible for the cost of the test if Medicare denies payment.
3. The ordering physician must provide an ICD-10 diagnosis code or narrative description, if required by the fiscal intermediary or carrier.
4. Organ- or disease-related panels should be billed only when all components of the panel are medically necessary.
5. Both ARUP- and client-customized panels should be billed to Medicare only when every component of the customized panel is medically necessary.
6. Medicare National Limitation Amounts for CPT codes are available through the Centers for Medicare & Medicaid Services (CMS) or its intermediaries. Medicaid reimbursement will be equal to or less than the amount of Medicare reimbursement.

The CPT Code(s) for test(s) profiled in this bulletin are for informational purposes only. The codes reflect our interpretation of CPT coding requirements, based upon AMA guidelines published annually. CPT codes are provided only as guidance to assist you in billing. ARUP strongly recommends that clients reconfirm CPT code information with their local intermediary or carrier. CPT coding is the sole responsibility of the billing party.

The regulations described above are only guidelines. Additional procedures may be required by your fiscal intermediary or carrier.

Hotline Page #	Test Number	Summary of Changes by Test Name	Name Change	Methodology	Performed/Reported Schedule	Specimen Requirements	Reference Interval	Interpretive Data	Note	CPT Code	Component Change	Other Interface Change	New Test	Inactive
2	0091586	Heroin - Screen with Reflex to Confirmation/Quantitation - Urine				x								
2	0099869	Homocysteine, Total				x	x							
3	2006385	Thrombotic Risk Reflexive Panel				x	x							
5	0030133	Thrombotic Risk, Inherited Etiologies (Most Common) with Reflex to Factor V Leiden				x	x							

HOTLINE: Effective **January 4, 2021**

0091586

Heroin - Screen with Reflex to Confirmation/Quantitation - Urine

HEROIN URN

Specimen Required: Collect: Random urine.

Specimen Preparation: Transfer 2 mL urine to an ARUP Standard Transport Tube. (Min: 1 mL)

Test is not performed at ARUP; separate specimens must be submitted when multiple tests are ordered.

Storage/Transport Temperature: Frozen

Unacceptable Conditions: Thawed specimens.

Stability (collection to initiation of testing): Ambient: **24 hours**; Refrigerated: 48 hours; Frozen: **3 months**

0099869

Homocysteine, Total

HOMOCY-QNT

Specimen Required: Collect: Green (lithium heparin), serum separator tube, or EDTA (K₂ and K₃).

Specimen Preparation: **Serum or plasma must be separated immediately after collection.** If immediate centrifugation is not possible, collected blood specimens should be kept on ice and centrifuged within one hour. Transfer 1 mL serum or plasma to an ARUP Standard Transport Tube. (Min: 0.5 mL)

Storage/Transport Temperature: Refrigerated.

Unacceptable Conditions: Sodium citrate.

Stability (collection to initiation of testing): Ambient: 4 days; Refrigerated: 1 month; Frozen: 10 months

Reference Interval:

Effective **January 4, 2021**

0-15 µmol/L, for both male and female

Specimen Required: Patient Prep: Fasting preferred. Refer to Specimen Handling at aruplab.com for hemostasis/thrombosis specimen handling guidelines.
Collect: Four Light Blue (Sodium Citrate) **AND** two Lavender (EDTA) **AND** two Serum Separator Tubes (SST). Also acceptable in place of one of the Serum Separator Tubes (SST): Green (Sodium or Lithium Heparin) **or** EDTA (K₂ or K₃).
Specimen Preparation: One Serum Separator Tube (SST), Green (Sodium or Lithium Heparin) or EDTA (K₂ or K₃) must be centrifuged and serum or plasma separated within 1 hour of collection. Transfer 1 mL centrifuged serum or plasma to ARUP Standard Transport Tube and label centrifuged tube for homocysteine testing. (Min: 0.5 mL) **AND** Transfer 2 mL serum into 2 ARUP Standard Transport Tubes, label as serum. (Min: 0.5 mL/tube) **AND** Transfer 7.5 mL platelet poor plasma prepared from the sodium citrate tubes to 5 ARUP Standard Transport Tubes, label as sodium citrate. (Min: 1 mL/tube) **AND** Transfer 3 mL lavender whole blood to 2 ARUP Standard Transport Tubes. (Min: 1 mL/tube)
Storage/Transport Temperature: **Light Blue (Sodium Citrate):** CRITICAL FROZEN. Separate specimens must be submitted when multiple tests are ordered.
Lavender Whole Blood and Serum, Green (Sodium or Lithium Heparin) or EDTA (K₂ or K₃): Refrigerated.
Unacceptable Conditions: Specimens collected in any tube type not listed above.
Stability (collection to initiation of testing): **Light Blue (Sodium Citrate):** Ambient: Unacceptable; Refrigerated: Unacceptable; Frozen: 2 weeks
Lavender Whole Blood: Ambient: 2 hours; Refrigerated: 1 week; Frozen: Unacceptable
Serum: Ambient: 2 hours; Refrigerated: 1 week; Frozen: 2 weeks
Green (Sodium or Lithium Heparin) or EDTA (K₂ or K₃): Ambient: 4 days; Refrigerated: 1 month; Frozen: 10 months

Reference Interval:

HOTLINE: Effective **January 4, 2021**

Test Number	Components	Reference Interval																												
	Prothrombin Time	12.0-15.5 seconds																												
	Dilute Russell Viper Venom Time (dRVVT)	33-44 seconds																												
	Dilute Russell Viper Venom (dRVVT) 1:1 Mix (performed if dRVVT > 44 seconds)	33-44 seconds																												
	Dilute Russell Viper Venom Time (dRVVT) Confirmation Test (performed if dRVVT 1:1 Mix > 44 seconds)	Negative																												
	Partial Thromboplastin Time	32-48 seconds																												
	Thrombin Time	14.7-19.5 seconds																												
	Reptilase Time	Less than 22.0 seconds																												
	PTT Heparin Neutralized	32-48 seconds																												
	Partial Thromboplastin Time 1:1 Mix (performed if PTT > 48 seconds)	32-48 seconds																												
	Platelet Neutralization Procedure (performed if PTT 1:1 Mix > 48 seconds)	Negative																												
	Hexagonal Phospholipid Neutralization	Negative																												
0050901	Cardiolipin Antibody, IgG	Effective August 18, 2014 0-14 GPL Negative 15-19 GPL Indeterminate 20-80 GPL Low to Moderately Positive 81 GPL or above High Positive																												
0050902	Cardiolipin Antibody, IgM	Effective August 18, 2014 0-12 MPL Negative 13-19 MPL Indeterminate 20-80 MPL Low to Moderately Positive 81 MPL or above High Positive																												
	Beta-2 Glycoprotein 1 Antibody, IgG	Effective August 18, 2014 0-20 SGU																												
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0098894	Protein S Free, Antigen	<table border="1"> <thead> <tr> <th>Age</th> <th>Male</th> <th>Female</th> </tr> </thead> <tbody> <tr> <td>1-89 days</td> <td>15-55%</td> <td>15-55%</td> </tr> <tr> <td>90-179 days</td> <td>35-92%</td> <td>35-92%</td> </tr> <tr> <td>180-364 days</td> <td>45-115%</td> <td>45-115%</td> </tr> <tr> <td>1-5 years</td> <td>62-120%</td> <td>62-120%</td> </tr> <tr> <td>6-9 years</td> <td>62-130%</td> <td>62-130%</td> </tr> <tr> <td>10-17 years</td> <td>60-140%</td> <td>60-140%</td> </tr> <tr> <td>18 years and older</td> <td>74-147%</td> <td>55-123%</td> </tr> </tbody> </table>	Age	Male	Female	1-89 days	15-55%	15-55%	90-179 days	35-92%	35-92%	180-364 days	45-115%	45-115%	1-5 years	62-120%	62-120%	6-9 years	62-130%	62-130%	10-17 years	60-140%	60-140%	18 years and older	74-147%	55-123%				
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0030010	Antithrombin, Enzymatic (Activity)	<table border="1"> <thead> <tr> <th>Age</th> <th>Reference Interval</th> </tr> </thead> <tbody> <tr> <td>1-4 days</td> <td>39-87%</td> </tr> <tr> <td>5-29 days</td> <td>41-93%</td> </tr> <tr> <td>30-89 days</td> <td>48-108%</td> </tr> <tr> <td>90-179 days</td> <td>73-121%</td> </tr> <tr> <td>180-364 days</td> <td>84-124%</td> </tr> <tr> <td>1-5 years</td> <td>82-139%</td> </tr> <tr> <td>6 years</td> <td>90-131%</td> </tr> <tr> <td>7-9 years</td> <td>90-135%</td> </tr> <tr> <td>10-11 years</td> <td>90-134%</td> </tr> <tr> <td>12-13 years</td> <td>90-132%</td> </tr> <tr> <td>14-15 years</td> <td>90-131%</td> </tr> <tr> <td>16-17 years</td> <td>87-131%</td> </tr> <tr> <td>18 years and older</td> <td>76-128%</td> </tr> </tbody> </table>	Age	Reference Interval	1-4 days	39-87%	5-29 days	41-93%	30-89 days	48-108%	90-179 days	73-121%	180-364 days	84-124%	1-5 years	82-139%	6 years	90-131%	7-9 years	90-135%	10-11 years	90-134%	12-13 years	90-132%	14-15 years	90-131%	16-17 years	87-131%	18 years and older	76-128%
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0030113	Protein C, Functional	Effective November 17, 2014 <table border="1"> <thead> <tr> <th>Age</th> <th>Reference Interval</th> </tr> </thead> <tbody> <tr> <td>1-4 days</td> <td>17-53%</td> </tr> <tr> <td>5-29 days</td> <td>20-64%</td> </tr> <tr> <td>30-89 days</td> <td>21-65%</td> </tr> <tr> <td>90-179 days</td> <td>28-80%</td> </tr> <tr> <td>180-364 days</td> <td>37-81%</td> </tr> <tr> <td>1-6 years</td> <td>40-92%</td> </tr> </tbody> </table>	Age	Reference Interval	1-4 days	17-53%	5-29 days	20-64%	30-89 days	21-65%	90-179 days	28-80%	180-364 days	37-81%	1-6 years	40-92%														
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HOTLINE: Effective **January 4, 2021**

		7-9 years	70-142%
		10-11 years	68-143%
		12-13 years	66-162%
		14-15 years	69-170%
		16-17 years	70-171%
		18 years and older	83-168%
	APC Resistance Profile	Effective February 21, 2011 2.00 or greater	
		Test Number	Components
		0030127	APC Resistance Profile
		0097720	Factor V Leiden (F5) R506Q Mutation
			Refer to report
			Refer to report
	Factor V Leiden by PCR & Fluorescence Monitoring	Negative: The sample is negative for factor V Leiden, R506Q mutation.	
0056060	Prothrombin (F2) c.*97G>A (G20210A) Pathogenic Variant		

0030133

Thrombotic Risk, Inherited Etiologies (Most Common) with Reflex to Factor V Leiden

THROM COM

Specimen Required: Patient Prep: Fasting preferred. Refer to Specimen Handling at aruplab.com for hemostasis/thrombosis specimen handling guidelines. Collect: Lt. blue (sodium citrate) **AND** lavender (EDTA) or pink (K₂EDTA) **AND** serum separator tube, green (sodium or lithium heparin), or EDTA (K₂ or K₃).
Specimen Preparation: Serum Separator Tube, Green (sodium or lithium heparin), or EDTA (K₂ or K₃) must be centrifuged and serum or plasma separated within 1 hour of collection. Transfer 1 mL centrifuged serum or plasma to ARUP Standard Transport Tube and label centrifuged tube for homocysteine testing. (Min: 0.5 mL) **AND** Transport two 2 mL aliquots platelet-poor plasma (sodium citrate) (Min: 1 mL/aliquot) **AND** Transport 5 mL whole blood (EDTA). (Min: 1 mL)
Storage/Transport Temperature: Light blue (sodium citrate): CRITICAL FROZEN. Separate specimens must be submitted when multiple tests are ordered.
Lavender whole blood and Serum or Green (sodium or lithium heparin): Refrigerated.
Unacceptable Conditions: Specimens collected in any tube type not listed above.
Stability (collection to initiation of testing): Light blue (sodium citrate): Ambient: Unacceptable; Refrigerated: Unacceptable; Frozen: 2 weeks
Lavender whole blood: Ambient: 2 hours; Refrigerated: 1 week; Frozen: Unacceptable
Serum, Green (sodium or lithium heparin), or EDTA (K₂ or K₃): Ambient: 4 days; Refrigerated: 1 month; Frozen: 10 months

Reference Interval: Effective November 18, 2013

Test Number	Components	Reference Interval	
0030235	Partial Thromboplastin Time	24-35 seconds	
0030095	Factor VIII, Activity		
		Age	Reference Interval
		0-6 years	56-191%
		7-9 years	76-199%
		10-11 years	80-209%
		12-13 years	72-198%
		14-15 years	69-237%
		16-17 years	63-221%
		18 years and older	56-191%
0099869	Homocysteine, Total	Effective January 4, 2021 0-15 µmol/L, for both male and female	
0056060	Prothrombin (F2) c.*97G>A (G20210A) Pathogenic Variant		
0030127	APC Resistance Profile	2.0 or greater	
	Factor V Leiden by PCR & Fluorescence Monitoring	Negative: The sample is negative for factor V Leiden, R506Q mutation.	