

500 Chipeta Way, Salt Lake City, Utah 84108-1221

phone: 801-583-2787, toll free: 800-522-2787

Jonathan R. Genzen, MD, PhD, Chief Medical Officer

Patient Age/Sex:

Female

Specimen Collected: 18-Nov-22 10:25**Hereditary Melanoma by NGS, DelDup** | **Received: 18-Nov-22 10:26** | **Report/Verified: 18-Nov-22 10:26**

| Procedure | Result | Units | Reference Interval |
|-----------------|---------------------------|-------|--------------------|
| MELCAN Specimen | Whole Blood | | |
| MELCAN Interp | Positive ^{f1 i1} | | |

Result Footnote

f1: MELCAN Interp
RESULT
One pathogenic variant was detected in the BAP1 gene.

PATHOGENIC VARIANT

Gene: BAP1 (NM_004656.3)
Nucleic Acid Change: c.436dupA; Heterozygous
Amino Acid Alteration: p.Arg146LysfsTer9
Inheritance: Autosomal dominant

INTERPRETATION

One pathogenic variant, c.436dupA; p.Arg146LysfsTer9, was detected in the BAP1 gene by massively parallel sequencing. Pathogenic germline variants in BAP1 are associated with autosomal dominant BAP1 tumor predisposition syndrome (MIM: 614327), which includes an increased risk of uveal and cutaneous melanoma, renal cell carcinoma, and additional cancers. Other genetic/environmental factors may influence an individual's risk of developing cancer. This individual's offspring have a 50 percent chance of inheriting the pathogenic variant.

Please refer to the background information included in this report for a list of the genes analyzed, methodology, and limitations of this test.

Evidence for variant classification:

The BAP1 c.436dupA; p.Arg146LysfsTer9 variant is reported in an individual with a personal history of mesothelioma and breast cancer, and a family history of multiple cancers (Hassan, 2019). This variant is absent from the Genome Aggregation Database, indicating it is not a common polymorphism. This variant causes a frameshift by duplicating a single nucleotide, so it is predicted to result in a truncated protein or mRNA subject to nonsense-mediated decay. Based on available information, this variant is considered to be pathogenic.

RECOMMENDATIONS

Genetic consultation is indicated, including a discussion of medical screening and management. At-risk family members should be offered testing for the identified pathogenic BAP1 variant (Familial Targeted Sequencing, ARUP test code 3005867).

COMMENTS

Likely benign and benign variants are not reported.
Variants in the following region(s) may not be detected by NGS with sufficient confidence in this sample due to technical limitations:

NONE

REFERENCES

Hassan R, et al. Inherited predisposition to malignant mesothelioma and overall survival following platinum chemotherapy. Proc Natl Acad Sci U S A. 2019;116(18):9008-9013.

Test Information

i1: MELCAN Interp
BACKGROUND INFORMATION: Hereditary Melanoma Panel,

*=Abnormal, #=Corrected, C=Critical, f=Result Footnote, H-High, i-Test Information, L-Low, t-Interpretive Text, @=Performing lab

Unless otherwise indicated, testing performed at:**ARUP Laboratories**

500 Chipeta Way, Salt Lake City, UT 84108

Laboratory Director: Jonathan R. Genzen, MD, PhD

ARUP Accession: 22-322-900074**Report Request ID:** 16445772**Printed:** 23-Dec-22 14:32

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Test Information

i1: MELCAN Interp

Sequencing and
Deletion/Duplication

CHARACTERISTICS: Pathogenic germline variants in multiple genes have been implicated in hereditary melanoma. Hereditary melanoma is usually characterized by early age of cancer onset (typically before 50 years of age), multiple primary melanomas, internal organ malignancies, and similar cancers in a closely related family member(s).

EPIDEMIOLOGY: Less than 10 percent of melanoma is associated with a hereditary cause.

CAUSE: Pathogenic germline variants in genes associated with hereditary melanoma

INHERITANCE: Autosomal dominant. Additionally, some genes are also associated with autosomal recessive childhood cancer predisposition or other syndromes.

GENES TESTED: BAP1; BRCA2; CDK4; CDKN2A*; MC1R; MITF*; POT1; PTEN*; RB1*; TERT; TP53

*One or more exons are not covered by sequencing and/or deletion/duplication analysis for the indicated gene; see limitations section below.

METHODOLOGY: Probe hybridization-based capture of all coding exons and exon-intron junctions of the targeted genes (including selected PTEN promoter variants), followed by massively parallel sequencing. Sanger sequencing was performed as necessary to fill in regions of low coverage and to confirm reported variants that do not meet acceptable quality metrics. A proprietary bioinformatic algorithm was used to detect large (single exon-level or larger) deletions or duplications in the indicated genes. Large deletions/duplications confirmed using an orthogonal exon-level microarray. Human genome build 19 (Hg 19) was used for data analysis. Testing of selected exons (and exon/intron boundaries) of PTEN was performed by bidirectional Sanger sequencing.

ANALYTICAL SENSITIVITY/SPECIFICITY: The analytical sensitivity is approximately 99 percent for single nucleotide variants (SNVs) and greater than 93 percent for insertions/duplications/deletions (indels) from 1-10 base pairs in size. Indels greater than 10 base pairs may be detected, but the analytical sensitivity may be reduced. Deletions of 2 exons or larger are detected with sensitivity greater than 97 percent; single exon deletions are detected with 62 percent sensitivity. Duplications of 3 exons or larger are detected at greater than 83 percent sensitivity. Specificity is greater than 99.9 percent for all variant classes.

LIMITATIONS: A negative result does not exclude a heritable form of melanoma or other cancer. This test only detects variants within the coding regions and intron-exon boundaries of the targeted genes. Deletions/duplications/insertions of any size may not be detected by massively parallel sequencing. Regulatory region variants and deep intronic variants will not be identified. Precise breakpoints for

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Test Information

i1: MELCAN Interp

large deletions or duplications are not determined in this assay and single exon deletions/duplications may not be detected based on the breakpoints of the rearrangement. The actual breakpoints for the deletion or duplication may extend beyond or be within the exon(s) reported. This test is not intended to detect duplications of 2 or fewer exons in size, though these may be identified. Single exon deletions are reported but called at a lower sensitivity. Diagnostic errors can occur due to rare sequence variations. In some cases, variants may not be identified due to technical limitations caused by the presence of pseudogenes, repetitive, or homologous regions. This test is not intended to detect low-level mosaic or somatic variants, gene conversion events, complex inversions, translocations, mitochondrial DNA (mtDNA) variants, or repeat expansions. Interpretation of this test result may be impacted if this patient has had an allogeneic stem cell transplantation. Noncoding transcripts were not analyzed. The following regions may have reduced sequencing sensitivity due to technical limitations of the assay:

RB1 (NM_000321) exon 22

The following regions are not sequenced due to technical limitations of the assay:

MITF (NM_001354607) exon 2

Deletions/duplications will not be called for the following exons:

CDKN2A (NM_000077, NM_001195132, NM_001363763, NM_058195) 2; MITF (NM_001354607) 2;

PTEN (NM_000314, NM_001304718) 9; PTEN (NM_001304717) 1,10; RB1 (NM_000321) 22

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the U.S. Food and Drug Administration. This test was performed in a CLIA-certified laboratory and is intended for clinical purposes.

Counseling and informed consent are recommended for genetic testing. Consent forms are available online.

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