

HOTLINE: Effective August 19, 2019

## MEDICARE COVERAGE OF LABORATORY TESTING

Please remember when ordering laboratory tests that are billed to Medicare/Medicaid or other federally funded programs, the following requirements apply:

1. Only tests that are medically necessary for the diagnosis or treatment of the patient should be ordered. Medicare does not pay for screening tests except for certain specifically approved procedures and may not pay for non-FDA approved tests or those tests considered experimental.
2. If there is reason to believe that Medicare will not pay for a test, the patient should be informed. The patient should then sign an Advance Beneficiary Notice (ABN) to indicate that he or she is responsible for the cost of the test if Medicare denies payment.
3. The ordering physician must provide an ICD-10 diagnosis code or narrative description, if required by the fiscal intermediary or carrier.
4. Organ- or disease-related panels should be billed only when all components of the panel are medically necessary.
5. Both ARUP- and client-customized panels should be billed to Medicare only when every component of the customized panel is medically necessary.
6. Medicare National Limitation Amounts for CPT codes are available through the Centers for Medicare & Medicaid Services (CMS) or its intermediaries. Medicaid reimbursement will be equal to or less than the amount of Medicare reimbursement.

The CPT Code(s) for test(s) profiled in this bulletin are for informational purposes only. The codes reflect our interpretation of CPT coding requirements, based upon AMA guidelines published annually. CPT codes are provided only as guidance to assist you in billing. ARUP strongly recommends that clients reconfirm CPT code information with their local intermediary or carrier. CPT coding is the sole responsibility of the billing party.

The regulations described above are only guidelines. Additional procedures may be required by your fiscal intermediary or carrier.

Hotline Page #	Test Number	Summary of Changes by Test Name	Name Change	Methodology	Performed/Reported Schedule	Specimen Requirements	Reference Interval	Interpretive Data	Note	CPT Code	Component Change	Other Interface Change	New Test	Inactive
9	<a href="#">3001868</a>	Acetylcholine Receptor Binding Antibody with reflex to Muscle-Specific Kinase (MuSK) Ab, IgG											x	
9	<a href="#">2005894</a>	Allergen Panel, IgE by ImmunoCap ISAC				x				x				
10	<a href="#">0055422</a>	Allergen, Drugs, Ampicillin											x	
81	<a href="#">0098422</a>	Allergen, Occupational, Phthalic Anhydride IgE												x
11	<a href="#">3001720</a>	Allergen, Region 10 Respiratory Panel IgE											x	
12	<a href="#">3001721</a>	Allergen, Region 11 Respiratory Panel IgE											x	
13	<a href="#">3001722</a>	Allergen, Region 12 Respiratory Panel IgE											x	
14	<a href="#">3001723</a>	Allergen, Region 13 Respiratory Panel IgE											x	

HOTLINE: Effective August 19, 2019

Hotline Page #	Test Number	Summary of Changes by Test Name	Name Change	Methodology	Performed/Reported Schedule	Specimen Requirements	Reference Interval	Interpretive Data	Note	CPT Code	Component Change	Other Interface Change	New Test	Inactive
15	<a href="#">3001724</a>	Allergen, Region 14 Respiratory Panel IgE											x	
16	<a href="#">3001725</a>	Allergen, Region 15 Respiratory Panel IgE											x	
17	<a href="#">3001728</a>	Allergen, Region 18 Respiratory Panel IgE											x	
18	<a href="#">3001729</a>	Allergen, Region 19 Respiratory Panel IgE											x	
19	<a href="#">3001712</a>	Allergen, Region 2 Respiratory Panel IgE											x	
20	<a href="#">3001730</a>	Allergen, Region 20 Respiratory Panel IgE											x	
21	<a href="#">3001713</a>	Allergen, Region 3 Respiratory Panel IgE											x	
22	<a href="#">3001714</a>	Allergen, Region 4 Respiratory Panel IgE											x	
23	<a href="#">3001715</a>	Allergen, Region 5 Respiratory Panel IgE											x	
24	<a href="#">3001716</a>	Allergen, Region 6 Respiratory Panel IgE											x	
25	<a href="#">3001717</a>	Allergen, Region 7 Respiratory Panel IgE											x	
26	<a href="#">3001718</a>	Allergen, Region 8 Respiratory Panel IgE											x	
27	<a href="#">3001719</a>	Allergen, Region 9 Respiratory Panel IgE											x	
28	<a href="#">3001726</a>	Allergen, Region 16 Respiratory Panel IgE											x	
29	<a href="#">3001727</a>	Allergen, Region 17 Respiratory Panel IgE											x	
29	<a href="#">0097933</a>	Allergen, Tree, Oak Red IgE		x										
81	<a href="#">2006038</a>	Allergens, Respiratory Panel, Region 10, Southwestern Grasslands (OK, TX) IgE												x
81	<a href="#">2006039</a>	Allergens, Respiratory Panel, Region 11, Rocky Mountain (AZ, ID, NM, WY CO, MT, UT) IgE												x
81	<a href="#">2006040</a>	Allergens, Respiratory Panel, Region 12, Arid Southwest (S. AZ, S.E. CA) IgE												x
81	<a href="#">2006041</a>	Allergens, Respiratory Panel, Region 13, Southern Coastal (CA) IgE												x
81	<a href="#">2006042</a>	Allergens, Respiratory Panel, Region 14, Central California (CA) IgE												x
81	<a href="#">2006043</a>	Allergens, Respiratory Panel, Region 15, Intermountain West (NV, S. ID) IgE												x
81	<a href="#">2006044</a>	Allergens, Respiratory Panel, Region 16, Inland Northwest (OR, Central and East WA) IgE												x
81	<a href="#">2006045</a>	Allergens, Respiratory Panel, Region 17, Pacific Northwest (NW CA, W. OR, WA) IgE												x
81	<a href="#">2006046</a>	Allergens, Respiratory Panel, Region 18, Alaska IgE												x
81	<a href="#">2006047</a>	Allergens, Respiratory Panel, Region 19, Puerto Rico IgE												x
81	<a href="#">2005718</a>	Allergens, Respiratory Panel, Region 2, Mid-Atlantic (DE, MD, VA, DC, NC) IgE												x
81	<a href="#">2006048</a>	Allergens, Respiratory Panel, Region 20, Hawaii IgE												x

HOTLINE: Effective August 19, 2019

Hotline Page #	Test Number	Summary of Changes by Test Name	Name Change	Methodology	Performed/Reported Schedule	Specimen Requirements	Reference Interval	Interpretive Data	Note	CPT Code	Component Change	Other Interface Change	New Test	Inactive
81	<a href="#">2006025</a>	Allergens, Respiratory Panel, Region 3, South Atlantic (GA, SC, N. FL) IgE												x
81	<a href="#">2006026</a>	Allergens, Respiratory Panel, Region 4, Subtropic Florida (S. of Orlando) IgE												x
81	<a href="#">2006031</a>	Allergens, Respiratory Panel, Region 5, Ohio Valley (IN, OH, TN, WV, KY) IgE												x
81	<a href="#">2006032</a>	Allergens, Respiratory Panel, Region 6, South Central (AL, AR, LA, MS) IgE												x
81	<a href="#">2006033</a>	Allergens, Respiratory Panel, Region 7, Northern Midwest (MI, WI, MN) IgE												x
81	<a href="#">2006034</a>	Allergens, Respiratory Panel, Region 8, Central Midwest (IL, MO, IA) IgE												x
81	<a href="#">2006037</a>	Allergens, Respiratory Panel, Region 9, Great Plains (KS, NE, ND, SD) IgE												x
30	<a href="#">0051495</a>	Alpha Thalassemia ( <i>HBA1</i> and <i>HBA2</i> ) 7 Deletions			x	x								
81	<a href="#">0091195</a>	Amantadine Quantitative, Serum or Plasma												x
30	<a href="#">2006480</a>	ANCA-Associated Vasculitis Profile (ANCA/MPO/ <b>PR3</b> ) with Reflex to ANCA Titer	x				x	x	x			x		
30	<a href="#">0010004</a>	Antibody Detection RBC with Reflex to ID	x			x			x	x				
31	<a href="#">0013003</a>	Antibody ID Package (IRL)			x	x								
31	<a href="#">0013005</a>	Antibody ID RBC Prenatal-Reflex to Titer	x			x			x					
31	<a href="#">0013006</a>	Antibody Titer			x	x			x					
31	<a href="#">2002068</a>	Anti-Neutrophil Cytoplasmic Antibody with Reflex to Titer and MPO/ <b>PR3</b> Antibodies	x				x	x	x					
32	<a href="#">0050317</a>	Antinuclear Antibodies (ANA), IgG by ELISA with Reflex to ANA HEp-2 Substrate, IgG by IFA and ENA Confirmation					x	x	x					
33	<a href="#">0050080</a>	Antinuclear Antibodies (ANA), IgG by ELISA with Reflex to ANA, HEp-2 Substrate, IgG by IFA						x						
33	<a href="#">3000601</a>	Antinuclear Antibody (ANA) with HEp-2 Substrate, IgG by IFA with Reflex by Pattern					x		x					
34	<a href="#">0051415</a>	Ashkenazi Jewish Diseases, 16 Genes									x			
34	<a href="#">2014314</a>	Autism and Intellectual Disability Comprehensive Panel							x					
81	<a href="#">0091165</a>	Barium Quantitative, Urine												x
34	<a href="#">3001410</a>	Basement Membrane Zone Antibody Panel											x	
35	<a href="#">2006193</a>	B-Cell Clonality Screening (IgH and IgK) by PCR				x								
35	<a href="#">2005017</a>	<i>BCR-ABL1</i> , Major (p210), Quantitative				x								
35	<a href="#">2005016</a>	<i>BCR-ABL1</i> , Minor (p190), Quantitative				x								

HOTLINE: Effective August 19, 2019

Hotline Page #	Test Number	Summary of Changes by Test Name	Name Change	Methodology	Performed/Reported Schedule	Specimen Requirements	Reference Interval	Interpretive Data	Note	CPT Code	Component Change	Other Interface Change	New Test	Inactive
36	<a href="#">2005010</a>	<i>BCR-ABL1</i> , Qualitative with Reflex to <i>BCR-ABL1</i> Quantitative				x								
81	<a href="#">0051421</a>	Beta Globin ( <i>HBB</i> ) HbS, HbC, and HbE Mutations												x
81	<a href="#">0051422</a>	Beta Globin ( <i>HBB</i> ) HbS, HbC, and HbE Mutations, Fetal												x
36	<a href="#">3001798</a>	<i>Blastomyces</i> Antigen Quantitative by EIA, Urine											x	
36	<a href="#">2002926</a>	<i>Blastomyces dermatitidis</i> Antigen Quantitative by EIA				x			x					
37	<a href="#">0062224</a>	<i>Blastomyces dermatitidis</i> Identification	x	x	x					x				
37	<a href="#">0060108</a>	Body Fluid Culture and Gram Stain				x			x					
37	<a href="#">0070053</a>	Bone Specific Alkaline Phosphatase				x								
37	<a href="#">2002498</a>	<i>BRAF</i> Codon 600 Mutation Detection by Pyrosequencing				x								
38	<a href="#">0051750</a>	<i>BRAF</i> Codon 600 Mutation Detection with Reflex to <i>MLH1</i> Promoter Methylation				x		x						
38	<a href="#">2007132</a>	<i>BRAF</i> V600E Mutation Detection in Hairy Cell Leukemia by Real-Time PCR, Quantitative				x								
81	<a href="#">2014493</a>	Bupivacaine Quantitative, Serum or Plasma												x
81	<a href="#">2014027</a>	Calcium, RBC												x
38	<a href="#">2008708</a>	Calculi Risk Assessment, Urine				x								
39	<a href="#">2010673</a>	<i>CALR</i> (Calreticulin) Exon 9 Mutation Analysis by PCR				x								
39	<a href="#">2002918</a>	Carbohydrate Deficient Transferrin for Congenital Disorders of Glycosylation (CDG)			x	x								
39	<a href="#">3001697</a>	Carbonic Anhydrase IX by IHC											x	
40	<a href="#">0080407</a>	Catecholamines Fractionated by LC-MS/MS, Urine Free					x							
41	<a href="#">2011114</a>	<i>CBFB-MYH11</i> inv(16) Detection, Quantitative				x								
41	<a href="#">2004247</a>	<i>CEBPA</i> Mutation Detection				x								
41	<a href="#">0080469</a>	Chromogranin A					x							
41	<a href="#">2011075</a>	<i>Coccidioides</i> Antigen Quantitative by EIA			x	x								
41	<a href="#">0062225</a>	<i>Coccidioides immitis</i> Identification	x	x	x					x				
42	<a href="#">3000480</a>	Comprehensive Systemic Sclerosis Panel					x		x			x		
42	<a href="#">0051668</a>	Connective Tissue Diseases Profile					x					x		
43	<a href="#">2000133</a>	Cytology, SurePath Liquid-Based Pap Test and Human Papillomavirus (HPV), High Risk by PCR, SurePath (for routine co-testing in women over 30)							x	x				
43	<a href="#">3001783</a>	Dermatomyositis and Polymyositis Panel											x	

HOTLINE: Effective August 19, 2019

Hotline Page #	Test Number	Summary of Changes by Test Name	Name Change	Methodology	Performed/Reported Schedule	Specimen Requirements	Reference Interval	Interpretive Data	Note	CPT Code	Component Change	Other Interface Change	New Test	Inactive
44	<a href="#">3001782</a>	Dermatomyositis Autoantibody Panel											x	
81	<a href="#">2013991</a>	Dermatomyositis Panel												x
81	<a href="#">0091219</a>	Diphenhydramine Quantitative, Urine												x
44	<a href="#">2002440</a>	EGFR Mutation Detection by Pyrosequencing				x			x					
45	<a href="#">2007914</a>	EPOR Mutation Detection by Sequencing				x								
81	<a href="#">0049050</a>	Esterase Stain, Nonspecific												x
81	<a href="#">2014674</a>	Expanded Carrier Screen, Genotyping												x
81	<a href="#">2014671</a>	Expanded Carrier Screen, Genotyping with Fragile X												x
45	<a href="#">3001781</a>	Extended Myositis Panel											x	
46	<a href="#">0050652</a>	Extractable Nuclear Antigen Antibodies (Smith/RNP, Smith, SSA 52, SSA 60, and SSB)	x				x					x		
46	<a href="#">0094030</a>	Felbamate					x	x						
46	<a href="#">3001161</a>	FLT3 ITD and TKD Mutation Detection				x								
47	<a href="#">2009033</a>	Fragile X (FMR1) with Reflex to Methylation Analysis				x		x	x					
47	<a href="#">2009034</a>	Fragile X (FMR1) with Reflex to Methylation Analysis, Fetal				x			x					
81	<a href="#">0091263</a>	Furosemide Quantitative, Serum or Plasma												x
48	<a href="#">3001648</a>	Gaucher Disease (GBA) Sequencing											x	
48	<a href="#">3000258</a>	Genetic Carrier Screen, (CF, FXS, and SMA) with Reflex to Methylation							x					
48	<a href="#">0080135</a>	Glucose-6-Phosphate Dehydrogenase				x								
49	<a href="#">2013590</a>	Heat Shock Protein 70, IgG by Immunoblot			x									
49	<a href="#">2010476</a>	Helicobacter pylori Breath Test, Adult				x								
49	<a href="#">2010925</a>	Helicobacter pylori Breath Test, Pediatric				x								
49	<a href="#">2005792</a>	Hemoglobin Evaluation Reflexive Cascade									x	x		
50	<a href="#">0092522</a>	Histoplasma Antigen Quantitative by EIA, Serum	x	x	x	x	x	x	x			x		
50	<a href="#">0062226</a>	Histoplasma capsulatum Identification	x	x	x					x				
81	<a href="#">0051067</a>	HLA DRB 3*,4*,5*												x
51	<a href="#">3001791</a>	HNF-1B by Immunohistochemistry											x	
51	<a href="#">2006444</a>	IDH1 and IDH2 Mutation Analysis, exon 4				x								
51	<a href="#">2014188</a>	IDH1 and IDH2 Mutation Analysis, Exon 4, Formalin-Fixed, Paraffin-Embedded (FFPE) Tissue				x								
52	<a href="#">0040227</a>	IGHV Mutation Analysis by Sequencing				x								
52	<a href="#">3001409</a>	Immunobullous Disease Panel, Epithelial Antibody Screening Panel											x	

HOTLINE: Effective August 19, 2019

Hotline Page #	Test Number	Summary of Changes by Test Name	Name Change	Methodology	Performed/Reported Schedule	Specimen Requirements	Reference Interval	Interpretive Data	Note	CPT Code	Component Change	Other Interface Change	New Test	Inactive
81	<a href="#">2008788</a>	Influenza A Virus H1/H3 Subtype by PCR with Reflex to H1N1 (2009) Oseltamivir Resistance by Sequencing												x
81	<a href="#">0093148</a>	Interferon-Alpha by ELISA, Serum												x
53	<a href="#">3001784</a>	Interstitial Lung Disease Autoantibody Panel											x	
81	<a href="#">2013993</a>	Interstitial Lung Disease Panel												x
54	<a href="#">3001568</a>	<i>IRF4/DUSP22</i> (6p25) Gene Rearrangement by FISH											x	
54	<a href="#">2002357</a>	<i>JAK2</i> Exon 12 Mutation Analysis by PCR				x								
55	<a href="#">0051245</a>	<i>JAK2</i> Gene, V617F Mutation, Qualitative				x	x							
55	<a href="#">2012084</a>	<i>JAK2</i> Gene, V617F Mutation, Qualitative with Reflex to <i>CALR</i> (Calreticulin) Exon 9 Mutation Analysis by PCR with Reflex to <i>MPL</i> Mutation Detection				x								
55	<a href="#">2012085</a>	<i>JAK2</i> Gene, V617F Mutation, Qualitative with Reflex to <i>JAK2</i> Exon 12 Mutation Analysis by PCR				x								
55	<a href="#">0040168</a>	<i>JAK2</i> Gene, V617F Mutation, Quantitative				x	x							
56	<a href="#">0020843</a>	Kidney Stone Risk Panel, Urine				x								
56	<a href="#">3000440</a>	<i>KIT</i> (D816V) Mutation by PCR				x								
56	<a href="#">2002437</a>	<i>KIT</i> Mutations in AML by Fragment Analysis and Sequencing				x								
56	<a href="#">0040248</a>	<i>KRAS</i> Mutation Detection				x								
57	<a href="#">2001932</a>	<i>KRAS</i> Mutation Detection with Reflex to <i>BRAF</i> Codon 600 Mutation Detection				x								
57	<a href="#">3001866</a>	Krebs von den Lungen-6											x	
81	<a href="#">2008003</a>	Leukemia/Lymphoma Phenotyping by Flow Cytometry												x
58	<a href="#">3001780</a>	Leukemia/Lymphoma Phenotyping Evaluation by Flow Cytometry											x	
59	<a href="#">0020421</a>	Lipid Panel						x			x			
60	<a href="#">0020468</a>	Lipid Panel, Extended						x			x			
81	<a href="#">0091200</a>	LSD, Serum or Plasma - Screen with Reflex to Confirmation/Quantitation												x
60	<a href="#">0091224</a>	LSD, Urine - Screen with Reflex to Confirmation/Quantitation			x	x								
60	<a href="#">0050119</a>	Lupus Comprehensive Reflexive Panel							x					
81	<a href="#">0091272</a>	Mercury, Hair												x
61	<a href="#">2009310</a>	<i>MGMT</i> Promoter Methylation Detection				x								
61	<a href="#">0051740</a>	Microsatellite Instability (MSI), HNPCC/Lynch Syndrome, by PCR				x								

HOTLINE: Effective August 19, 2019

Hotline Page #	Test Number	Summary of Changes by Test Name	Name Change	Methodology	Performed/Reported Schedule	Specimen Requirements	Reference Interval	Interpretive Data	Note	CPT Code	Component Change	Other Interface Change	New Test	Inactive
61	<a href="#">2002499</a>	<i>MLH1</i> Promoter Methylation, Paraffin				x		x						
62	<a href="#">0050615</a>	Monoclonal Protein Detection Quantitation and Characterization, SPEP, IFE, IgA, IgG, IgM, Serum		x		x	x			x		x		
63	<a href="#">2002715</a>	Monoclonal Protein Detection, Quantitation, Characterization, SPEP, IFE, IgA, IgG, IgM, FLC		x		x	x			x		x		
64	<a href="#">2007967</a>	Motor and Sensory Neuropathy Evaluation with Immunofixation Electrophoresis and Reflex to Titer and Neuronal Immunoblot		x		x	x			x		x		
65	<a href="#">0051225</a>	Motor Neuropathy Panel		x		x	x			x		x		
66	<a href="#">2005545</a>	<i>MPL</i> Mutation Detection by Capillary Electrophoresis				x								
66	<a href="#">0050707</a>	MPO/ <b>PR3</b> (ANCA) Antibodies	x				x					x		
67	<a href="#">3000523</a>	Mumps Virus by PCR				x								
67	<a href="#">3001869</a>	Myasthenia Gravis Reflexive Panel											x	
68	<a href="#">2009318</a>	<i>MYD88</i> L265P Mutation Detection by PCR, Quantitative				x								
68	<a href="#">0050526</a>	Myeloperoxidase ( <b>MPO</b> ) Antibody	x											
68	<a href="#">0050742</a>	Myocardial Antibody, IgG with Reflex to Titer			x									
81	<a href="#">2013961</a>	Myositis Extended Panel												x
69	<a href="#">3001907</a>	Myotonic Dystrophy Type 1 ( <i>DMPK</i> ) CTG Expansion											x	
69	<a href="#">0099465</a>	Neuronal Cell Antibodies Quantitative, Serum			x									
70	<a href="#">0096657</a>	Neutrophil Oxidative Burst Assay (DHR)				x								
81	<a href="#">0092140</a>	Nitrogen, Total, Urine												x
70	<a href="#">3000066</a>	<i>NPM1</i> Mutation Detection by RT-PCR, Quantitative				x								
70	<a href="#">2003123</a>	<i>NRAS</i> Mutation Detection by Pyrosequencing				x								
71	<a href="#">3001760</a>	Pancreatitis ( <i>PRSS1</i> ) Deletion/Duplication											x	
81	<a href="#">2002016</a>	Pancreatitis ( <i>PRSS1</i> ) Sequencing												x
72	<a href="#">3001768</a>	Pancreatitis ( <i>PRSS1</i> ) Sequencing and Deletion/Duplication											x	
73	<a href="#">3001764</a>	Pancreatitis ( <i>SPINK1</i> ) Deletion/Duplication											x	
73	<a href="#">3000496</a>	PanFungal Identification by Sequencing				x								
73	<a href="#">3001170</a>	Platelet Antigen 1 Genotyping (HPA-1)			x									
74	<a href="#">3000193</a>	Platelet Antigen Genotyping Panel			x									
74	<a href="#">2002871</a>	<i>PML-RARA</i> Translocation, t(15;17) by RT-PCR, Quantitative				x			x					
74	<a href="#">2009226</a>	<i>Pneumocystis jirovecii</i> DFA with Reflex to <i>Pneumocystis jirovecii</i> by PCR			x									



HOTLINE: Effective August 19, 2019

Hotline Page #	Test Number	Summary of Changes by Test Name	Name Change	Methodology	Performed/Reported Schedule	Specimen Requirements	Reference Interval	Interpretive Data	Note	CPT Code	Component Change	Other Interface Change	New Test	Inactive
81	<a href="#">2013992</a>	Polymyositis and Dermatomyositis Panel												x
75	<a href="#">2002109</a>	Protein Electrophoresis with Reflex to Immunofixation Electrophoresis Monoclonal Protein Detection, Quantitation & Characterization, IgA, IgG, and IgM, Serum	x	x		x	x			x		x		
76	<a href="#">0050640</a>	Protein Electrophoresis, Serum		x		x	x			x		x		
76	<a href="#">3000010</a>	Relapsing Fever <i>Borrelia</i> Species by PCR				x			x					
76	<a href="#">0051368</a>	RhD Gene ( <i>RHD</i> ) Copy Number	x					x				x		
77	<a href="#">2003176</a>	Rufinamide, Serum or Plasma					x							
77	<a href="#">2010138</a>	<i>RUNX1-RUNX1T1</i> ( <i>AML1-ETO</i> ) t(8;21) Detection, Quantitative				x								
77	<a href="#">0050527</a>	Serine Proteinase 3 (PR3) Antibody	x									x		
77	<a href="#">0050085</a>	Smith (ENA) Antibody, IgG						x						
77	<a href="#">3000460</a>	Smith and Smith/RNP (ENA) Antibodies, IgG	x				x					x		
77	<a href="#">0050470</a>	Smith/RNP (ENA) Antibody, IgG	x					x	x			x		
78	<a href="#">0060137</a>	Stool Culture, <i>Yersinia</i>				x								
78	<a href="#">0060705</a>	<i>Streptococcus</i> Group B by PCR				x								
81	<a href="#">0091568</a>	Sulfhemoglobin Quantitative, Whole Blood												x
78	<a href="#">2008771</a>	Supersaturation Profile, Urine			x	x								
78	<a href="#">0093199</a>	T-Cell Clonality by Flow Cytometry Analysis of TCR V-Beta				x								
79	<a href="#">0055567</a>	T-Cell Clonality Screening by PCR				x								
79	<a href="#">2013484</a>	TP53 Somatic Mutation, Prognostic			x									
79	<a href="#">3001704</a>	Treponema Pallidum by IHC											x	
81	<a href="#">0098803</a>	Troponin T												x
80	<a href="#">3001831</a>	Troponin T (cTnT) 5 <sup>th</sup> Generation											x	
80	<a href="#">0065031</a>	<i>Ureaplasma</i> Species and <i>Mycoplasma hominis</i> Culture				x								
80	<a href="#">0020056</a>	Viscosity, Serum					x							
80	<a href="#">2013942</a>	Zika Virus IgM Antibody Capture (MAC) by ELISA						x	x			x		



HOTLINE: Effective August 19, 2019

**New Test**     [3001868](#)     **Acetylcholine Receptor Binding Antibody with reflex to Muscle-Specific Kinase (MuSK) Ab, IgG**     **ACHR BIN R**

[Click for Pricing](#)



#### Additional Technical Information

**Methodology:** Quantitative Radioimmunoassay  
**Performed:** Sun-Sat  
**Reported:** 2-8 days

**Specimen Required:** Collect: Serum Separator Tube (SST).  
Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP Standard Transport Tube. (Min: 0.3 mL)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Plasma. Contaminated, hemolyzed, or severely lipemic specimens.  
Stability (collection to initiation of testing): After separation from cells: Ambient: 8 hours; Refrigerated: 2 weeks; Frozen: 1 month (avoid repeated freeze/thaw cycles)

#### Reference Interval:

Test Number	Components	Reference Interval	
0080009	Acetylcholine Receptor Binding Antibody	Negative	0.0-0.4 nmol/L
		Positive	0.5 nmol/L or greater
3001576	Muscle-Specific Kinase (MuSK) Antibody, IgG	Negative	0.00-0.03 nmol/L
		Positive	0.04 nmol/L or greater

**Interpretive Data:** Approximately 85-90 percent of patients with myasthenia gravis (MG) express antibodies to the acetylcholine receptor (AChR), which can be divided into binding, blocking, and modulating antibodies. Binding antibody can activate complement and lead to loss of AChR. Blocking antibody may impair binding of acetylcholine to the receptor, leading to poor muscle contraction. Modulating antibody causes receptor endocytosis resulting in loss of AChR expression, which correlates most closely with clinical severity of disease. Approximately 10-15 percent of individuals with confirmed myasthenia gravis have no measurable binding, blocking, or modulating antibodies.

See Compliance Statement B: [www.aruplab.com/CS](http://www.aruplab.com/CS)

**Note:** If Acetylcholine Receptor Binding Antibody result is less than or equal to 0.4 nmol/L then Muscle-Specific Kinase (MuSK) Ab, IgG (ARUP test code 3001576) will be added. Additional charges apply.

**CPT Code(s):** 83519; if reflexed, add 83519

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

[2005894](#)     **Allergen Panel, IgE by ImmunoCap ISAC**     **ISAC MICRO**

**Specimen Required:** Collect: Serum Separator Tube (**SST**). Also acceptable: Lavender (EDTA) or Green (Sodium or Lithium Heparin).  
Specimen Preparation: Transfer 0.4 mL serum or plasma to an ARUP Standard Transport Tube. (Min: 0.25 mL)  
Storage/Transport Temperature: Refrigerated. Also acceptable: Frozen.  
Stability (collection to initiation of testing): Ambient: **24 hours**; Refrigerated: 1 week; **Frozen: 1 month**

**CPT Code(s):** **86008** x112

HOTLINE: Effective August 19, 2019

**New Test**      **0055422**      **Allergen, Drugs, Ampicillin**      **AMPICIL**  
 Available Now  
[Click for Pricing](#)

**Methodology:** Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay  
**Performed:** Sun-Sat  
**Reported:** 1-2 days

**Specimen Required:** Patient Prep: Multiple patient encounters should be avoided.  
Collect: Serum Separator Tube (SST). Multiple specimen tubes should be avoided.  
Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 0.25 mL serum **plus** 0.1 mL for each additional allergen ordered to an ARUP Standard Transport Tube. (Min: 0.25 mL **plus** 0.04 mL for each allergen ordered)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Hemolyzed, icteric, or lipemic specimens.  
Stability (collection to initiation of testing): After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

**Reference Interval:**

Reporting Range (reported in kU/L)	Probability of IgE Mediated Clinical Reaction	Class Scoring
Less than 0.10	No significant level detected	0
0.10 - 0.34	Clinical relevance undetermined	0/1
0.35 - 0.70	Low	1
0.71 - 3.50	Moderate	2
3.51 - 17.50	High	3
17.51 - 50.00	Very high	4
50.01 - 100.00	Very high	5
Greater than 100.00	Very high	6

**Interpretive Data:** Allergen results of 0.10-0.34 kU/L are intended for specialist use as the clinical relevance is undetermined. Even though increasing ranges are reflective of increasing concentrations of allergen-specific IgE, these concentrations may not correlate with the degree of clinical response or skin testing results when challenged with a specific allergen. The correlation of allergy laboratory results with clinical history and in vivo reactivity to specific allergens is essential. A negative test may not rule out clinical allergy or even anaphylaxis.

**CPT Code(s):** 86003

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

**New Test** [3001720](#)  
[Click for Pricing](#)

**Allergen, Region 10 Respiratory Panel IgE**

**REG10PANEL**

**Methodology:** Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay  
**Performed:** Sun-Sat  
**Reported:** 1-2 days

**Specimen Required:** Patient Prep: Multiple patient encounters should be avoided.

Collect: Serum Separator Tube (SST). Multiple specimen tubes should be avoided.

Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 4 mL serum to an ARUP Standard Transport Tube. (Min: 1.4 mL)

Storage/Transport Temperature: Refrigerated.

Unacceptable Conditions: Hemolyzed, icteric, or lipemic specimens.

Stability (collection to initiation of testing): After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

**Reference Interval:**

**Reference Intervals for all Components**

Reporting Range (reported in kU/L)	Probability of IgE Mediated Clinical Reaction	Class Scoring
Less than 0.10	No significant level detected	0
0.10-0.34	Clinical relevance undetermined	0/1
0.35-0.70	Low	1
0.71-3.50	Moderate	2
3.51-17.50	High	3
17.51 - 50.00	Very High	4
50.01 - 100.00	Very high	5
Greater than 100.00	Very high	6

Test Number	Components	Reference Interval	
0050345	Immunoglobulin E	Effective November 17, 2014	
		Age	Reference Interval
		0-5 months	13 kU/L or less
		6-12 months	34 kU/L or less
		1-2 years	97 kU/L or less
		3 years	199 kU/L or less
		4-6 years	307 kU/L or less
		7-8 years	403 kU/L or less
		9-12 years	696 kU/L or less
		13-15 years	629 kU/L or less
		16-17 years	537 kU/L or less
		18 years and older	214 kU/L or less

**Interpretive Data:** Allergen results of 0.10-0.34 kU/L are intended for specialist use as the clinical relevance is undetermined. Even though increasing ranges are reflective of increasing concentrations of allergen-specific IgE, these concentrations may not correlate with the degree of clinical response or skin testing results when challenged with a specific allergen. The correlation of allergy laboratory results with clinical history and in vivo reactivity to specific allergens is essential. A negative test may not rule out clinical allergy or even anaphylaxis.

**Note:** Allergens included in this panel: *Alternaria alternata* (*tenuis*), *Aspergillus fumigatus*, Bermuda Grass, Birch Tree, Box Elder/Maple Tree, Cat Dander, Cockroach (German), Common Short Ragweed, Cottonwood Tree, *D. farinae* (mites), *D. Pteronyssinus* (mites), Dog Dander, Elm Tree, *Hormodendrum* (*Cladosporium*), Marsh Elder, Mouse Epithelium, Mountain Cedar (Juniper) Tree, *Mucor racemosus*, Nettle, Oak Tree, Pecan (White Hickory) Tree, *Penicillium notatum*, Pigweed, Sheep Sorrel (Dock), Timothy Grass, White Ash Tree, White Mulberry Tree, and IgE Serum Total.

**CPT Code(s):** 86003 x27; 82785

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

**New Test** [3001721](#) **Allergen, Region 11 Respiratory Panel IgE** **REG11PANEL**  
[Click for Pricing](#)

**Methodology:** Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay  
**Performed:** Sun-Sat  
**Reported:** 1-2 days

**Specimen Required:** Patient Prep: Multiple patient encounters should be avoided.  
Collect: Serum Separator Tube (SST). Multiple specimen tubes should be avoided.  
Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 4 mL serum to an ARUP Standard Transport Tube. (Min: 1.3 mL)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Hemolyzed, icteric, or lipemic specimens.  
Stability (collection to initiation of testing): After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

**Reference Interval:**

**Reference Intervals for all Components**

Reporting Range (reported in kU/L)	Probability of IgE Mediated Clinical Reaction	Class Scoring
Less than 0.10	No significant level detected	0
0.10-0.34	Clinical relevance undetermined	0/1
0.35-0.70	Low	1
0.71-3.50	Moderate	2
3.51-17.50	High	3
17.51 - 50.00	Very High	4
50.01 - 100.00	Very High	5
Greater than 100.00	Very High	6

Test Number	Components	Reference Interval	
0050345	Immunoglobulin E	Effective November 17, 2014	
		Age	Reference Interval
		0-5 months	13 kU/L or less
		6-12 months	34 kU/L or less
		1-2 years	97 kU/L or less
		3 years	199 kU/L or less
		4-6 years	307 kU/L or less
		7-8 years	403 kU/L or less
		9-12 years	696 kU/L or less
		13-15 years	629 kU/L or less
		16-17 years	537 kU/L or less
		18 years and older	214 kU/L or less

**Interpretive Data:** Allergen results of 0.10-0.34 kU/L are intended for specialist use as the clinical relevance is undetermined. Even though increasing ranges are reflective of increasing concentrations of allergen-specific IgE, these concentrations may not correlate with the degree of clinical response or skin testing results when challenged with a specific allergen. The correlation of allergy laboratory results with clinical history and in vivo reactivity to specific allergens is essential. A negative test may not rule out clinical allergy or even anaphylaxis.

**Note:** Allergens included in this panel: Alder Tree, *Alternaria alternata (tenuis)*, *Aspergillus fumigatus*, Bermuda Grass, Box Elder/Maple Tree, Cat Dander, Cockroach (German), Common Short Ragweed, Cottonwood Tree, *D. farinae* (mites), *D. Pteronyssinus* (mites), Dog Dander, Elm Tree, *Hormodendrum (Cladosporium)*, Mouse Epithelium, Mountain Cedar (Juniper) Tree, *Mucor racemosus*, Mugwort, Oak Tree, Olive Tree, *Penicillium notatum*, Pigweed, Russian Thistle, Sheep Sorrel (Dock), Timothy Grass, White Mulberry Tree, and IgE Serum Total.

**CPT Code(s):** 86003 x26; 82785

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

**New Test** [3001722](#) **Allergen, Region 12 Respiratory Panel IgE** **REG12PANEL**  
[Click for Pricing](#)

**Methodology:** Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay  
**Performed:** Sun-Sat  
**Reported:** 1-2 days

**Specimen Required:** Patient Prep: Multiple patient encounters should be avoided.  
Collect: Serum Separator Tube (SST). Multiple specimen tubes should be avoided.  
Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 4 mL serum to an ARUP Standard Transport Tube. (Min: 1.3 mL)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Hemolyzed, icteric, or lipemic specimens.  
Stability (collection to initiation of testing): After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

**Reference Interval:**

**Reference Intervals for all Components**

Reporting Range (reported in kU/L)	Probability of IgE Mediated Clinical Reaction	Class Scoring
Less than 0.10	No significant level detected	0
0.10-0.34	Clinical relevance undetermined	0/1
0.35-0.70	Low	1
0.71-3.50	Moderate	2
3.51-17.50	High	3
17.51 - 50.00	Very High	4
50.01 - 100.00	Very high	5
Greater than 100.00	Very high	6

Test Number	Components	Reference Interval	
0050345	Immunoglobulin E	Effective November 17, 2014	
		Age	Reference Interval
		0-5 months	13 kU/L or less
		6-12 months	34 kU/L or less
		1-2 years	97 kU/L or less
		3 years	199 kU/L or less
		4-6 years	307 kU/L or less
		7-8 years	403 kU/L or less
		9-12 years	696 kU/L or less
		13-15 years	629 kU/L or less
		16-17 years	537 kU/L or less
		18 years and older	214 kU/L or less

**Interpretive Data:** Allergen results of 0.10-0.34 kU/L are intended for specialist use as the clinical relevance is undetermined. Even though increasing ranges are reflective of increasing concentrations of allergen-specific IgE, these concentrations may not correlate with the degree of clinical response or skin testing results when challenged with a specific allergen. The correlation of allergy laboratory results with clinical history and in vivo reactivity to specific allergens is essential. A negative test may not rule out clinical allergy or even anaphylaxis.

**Note:** Allergens included in this panel: Acacia Tree, *Alternaria alternata* (*tenuis*), *Aspergillus fumigatus*, Bermuda Grass, Cat Dander, Cockroach (German), Common Short Ragweed, Cottonwood Tree, *D. farinae* (mites), *D. Pteronyssinus* (mites), Dog Dander, Elm Tree, *Hormodendrum* (*Cladosporium*), Johnson Grass, Mouse Epithelium, Mountain Cedar (Juniper) Tree, *Mucor racemosus*, Mugwort, Oak Tree, Olive Tree, *Penicillium notatum*, Perennial Rye Grass, Pigweed, Russian Thistle and IgE Serum Total.

**CPT Code(s):** 86003 x24; 82785

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

**New Test** [3001723](#)  
[Click for Pricing](#)

**Allergen, Region 13 Respiratory Panel IgE**

**REG13PANEL**

**Methodology:** Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay  
**Performed:** Sun-Sat  
**Reported:** 1-2 days

**Specimen Required:** Patient Prep: Multiple patient encounters should be avoided.

Collect: Serum Separator Tube (SST). Multiple specimen tubes should be avoided.

Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 4 mL serum to an ARUP Standard Transport Tube. (Min: 1.3 mL)

Storage/Transport Temperature: Refrigerated.

Unacceptable Conditions: Hemolyzed, icteric, or lipemic specimens.

Stability (collection to initiation of testing): After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

**Reference Interval:**

**Reference Intervals for all Components**

Reporting Range (reported in kU/L)	Probability of IgE Mediated Clinical Reaction	Class Scoring
Less than 0.10	No significant level detected	0
0.10-0.34	Clinical relevance undetermined	0/1
0.35-0.70	Low	1
0.71-3.50	Moderate	2
3.51-17.50	High	3
17.51 - 50.00	Very High	4
50.01 - 100.00	Very high	5
Greater than 100.00	Very high	6

Test Number	Components	Reference Interval	
0050345	Immunoglobulin E	Effective November 17, 2014	
		Age	Reference Interval
		0-5 months	13 kU/L or less
		6-12 months	34 kU/L or less
		1-2 years	97 kU/L or less
		3 years	199 kU/L or less
		4-6 years	307 kU/L or less
		7-8 years	403 kU/L or less
		9-12 years	696 kU/L or less
		13-15 years	629 kU/L or less
		16-17 years	537 kU/L or less
		18 years and older	214 kU/L or less
		</	

**Interpretive Data:** Allergen results of 0.10-0.34 kU/L are intended for specialist use as the clinical relevance is undetermined. Even though increasing ranges are reflective of increasing concentrations of allergen-specific IgE, these concentrations may not correlate with the degree of clinical response or skin testing results when challenged with a specific allergen. The correlation of allergy laboratory results with clinical history and in vivo reactivity to specific allergens is essential. A negative test may not rule out clinical allergy or even anaphylaxis.

**Note:** Allergens included in this panel: Alder Tree, *Alternaria alternata (tenuis)*, *Aspergillus fumigatus*, Bermuda Grass, Cat Dander, Cockroach (German), Common Short Ragweed, Cottonwood Tree, *D. farinae* (mites), *D. Pteronyssinus* (mites), Dog Dander, Elm Tree, *Hormodendrum (Cladosporium)*, Johnson Grass, Mouse Epithelium, Mountain Cedar (Juniper) Tree, *Mucor racemosus*, Mugwort, Oak Tree, Olive Tree, *Penicillium notatum*, Pigweed, Russian Thistle, Timothy Grass, Walnut Tree, White Mulberry Tree and IgE Serum Total.

**CPT Code(s):** 86003 x26; 82785

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

**New Test**     [3001724](#)     **Allergen, Region 14 Respiratory Panel IgE**     **REG14PANEL**  
[Click for Pricing](#)

**Methodology:** Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay  
**Performed:** Sun-Sat  
**Reported:** 1-2 days

**Specimen Required:** Patient Prep: Multiple patient encounters should be avoided.  
Collect: Serum Separator Tube (SST). Multiple specimen tubes should be avoided.  
Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 4 mL serum to an ARUP Standard Transport Tube. (Min: 1.3 mL)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Hemolyzed, icteric, or lipemic specimens.  
Stability (collection to initiation of testing): After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

**Reference Interval:**

**Reference Intervals for all Components**

Reporting Range (reported in kU/L)	Probability of IgE Mediated Clinical Reaction	Class Scoring
Less than 0.10	No significant level detected	0
0.10-0.34	Clinical relevance undetermined	0/1
0.35-0.70	Low	1
0.71-3.50	Moderate	2
3.51-17.50	High	3
17.51 - 50.00	Very High	4
50.01 - 100.00	Very high	5
Greater than 100.00	Very high	6

Test Number	Components	Reference Interval	
0050345	Immunoglobulin E	Effective November 17, 2014	
		Age	Reference Interval
		0-5 months	13 kU/L or less
		6-12 months	34 kU/L or less
		1-2 years	97 kU/L or less
		3 years	199 kU/L or less
		4-6 years	307 kU/L or less
		7-8 years	403 kU/L or less
		9-12 years	696 kU/L or less
		13-15 years	629 kU/L or less
		16-17 years	537 kU/L or less
		18 years and older	214 kU/L or less

**Interpretive Data:** Allergen results of 0.10-0.34 kU/L are intended for specialist use as the clinical relevance is undetermined. Even though increasing ranges are reflective of increasing concentrations of allergen-specific IgE, these concentrations may not correlate with the degree of clinical response or skin testing results when challenged with a specific allergen. The correlation of allergy laboratory results with clinical history and in vivo reactivity to specific allergens is essential. A negative test may not rule out clinical allergy or even anaphylaxis.

**Note:** Allergens included in this panel: Alder Tree, *Alternaria alternata (tenuis)*, *Aspergillus fumigatus*, Bermuda Grass, Birch Tree, Cat Dander, Cockroach (German), Common Short Ragweed, *D. farinae* (mites), *D. Pteronyssinus* (mites), Dog Dander, Elm Tree, *Hormodendrum (Cladosporium)*, Mouse Epithelium, Mountain Cedar (Juniper) Tree, *Mucor racemosus*, Mugwort, Oak Tree, Olive Tree, *Penicillium notatum*, Pigweed, Russian Thistle, Sycamore Tree, Timothy Grass, White Mulberry Tree, and IgE Serum Total

**CPT Code(s):** 86003 x25; 82785

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.



HOTLINE: Effective August 19, 2019

**New Test**     [3001725](#)     **Allergen, Region 15 Respiratory Panel IgE**     **REG15PANEL**  
[Click for Pricing](#)

**Methodology:** Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay  
**Performed:** Sun-Sat  
**Reported:** 1-2 days

**Specimen Required:** Patient Prep: Multiple patient encounters should be avoided.  
Collect: Serum Separator Tube (SST). Multiple specimen tubes should be avoided.  
Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 4 mL serum to an ARUP Standard Transport Tube. (Min: 1.3 mL)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Hemolyzed, icteric, or lipemic specimens.  
Stability (collection to initiation of testing): After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

**Reference Interval:**

**Reference Intervals for all Components**

Reporting Range (reported in kU/L)	Probability of IgE Mediated Clinical Reaction	Class Scoring
Less than 0.10	No significant level detected	0
0.10-0.34	Clinical relevance undetermined	0/1
0.35-0.70	Low	1
0.71-3.50	Moderate	2
3.51-17.50	High	3
17.51 - 50.00	Very High	4
50.01 - 100.00	Very high	5
Greater than 100.00	Very high	6

Test Number	Components	Reference Interval	
0050345	Immunoglobulin E	Effective November 17, 2014	
		Age	Reference Interval
		0-5 months	13 kU/L or less
		6-12 months	34 kU/L or less
		1-2 years	97 kU/L or less
		3 years	199 kU/L or less
		4-6 years	307 kU/L or less
		7-8 years	403 kU/L or less
		9-12 years	696 kU/L or less
		13-15 years	629 kU/L or less
		16-17 years	537 kU/L or less
		18 years and older	214 kU/L or less

**Interpretive Data:** Allergen results of 0.10-0.34 kU/L are intended for specialist use as the clinical relevance is undetermined. Even though increasing ranges are reflective of increasing concentrations of allergen-specific IgE, these concentrations may not correlate with the degree of clinical response or skin testing results when challenged with a specific allergen. The correlation of allergy laboratory results with clinical history and in vivo reactivity to specific allergens is essential. A negative test may not rule out clinical allergy or even anaphylaxis.

**Note:** Allergens included in this panel: *Alternaria alternata* (*tenuis*), *Aspergillus fumigatus*, Bermuda Grass, BoxElder/Maple Tree, Cat Dander, Cockroach (German), Common Short Ragweed, Cottonwood Tree, *D. farinae* (mites), *D. Pteronyssinus* (mites), Dog Dander, Elm Tree, *Hormodendrum* (*Cladosporium*), Mouse Epithelium, Mountain Cedar (Juniper) Tree, *Mucor racemosus*, Mugwort, Oak Tree, Olive Tree, *Penicillium notatum*, Pigweed, Russian Thistle, Timothy Grass, White Mulberry Tree, and IgE Serum Total

**CPT Code(s):** 86003 x24; 82785

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

**New Test**     [3001728](#)     **Allergen, Region 18 Respiratory Panel IgE**     **REG18PANEL**  
[Click for Pricing](#)

**Methodology:** Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay  
**Performed:** Sun-Sat  
**Reported:** 1-2 days

**Specimen Required:** Patient Prep: Multiple patient encounters should be avoided.  
Collect: Serum Separator Tube (SST). Multiple specimen tubes should be avoided.  
Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 3 mL serum to an ARUP Standard Transport Tube. (Min: 1 mL)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Hemolyzed, icteric, or lipemic specimens.  
Stability (collection to initiation of testing): After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

**Reference Interval:**

**Reference Intervals for all Components**

Reporting Range (reported in kU/L)	Probability of IgE Mediated Clinical Reaction	Class Scoring
Less than 0.10	No significant level detected	0
0.10-0.34	Clinical relevance undetermined	0/1
0.35-0.70	Low	1
0.71-3.50	Moderate	2
3.51-17.50	High	3
17.51 - 50.00	Very High	4
50.01 - 100.00	Very high	5
Greater than 100.00	Very high	6

Test Number	Components	Reference Interval	
0050345	Immunoglobulin E	Effective November 17, 2014	
		Age	Reference Interval
		0-5 months	13 kU/L or less
		6-12 months	34 kU/L or less
		1-2 years	97 kU/L or less
		3 years	199 kU/L or less
		4-6 years	307 kU/L or less
		7-8 years	403 kU/L or less
		9-12 years	696 kU/L or less
		13-15 years	629 kU/L or less
		16-17 years	537 kU/L or less
		18 years and older	214 kU/L or less

**Interpretive Data:** Allergen results of 0.10-0.34 kU/L are intended for specialist use as the clinical relevance is undetermined. Even though increasing ranges are reflective of increasing concentrations of allergen-specific IgE, these concentrations may not correlate with the degree of clinical response or skin testing results when challenged with a specific allergen. The correlation of allergy laboratory results with clinical history and in vivo reactivity to specific allergens is essential. A negative test may not rule out clinical allergy or even anaphylaxis.

**Note:** Allergens included in this panel: Alder Tree, *Alternaria alternata (tenuis)*, *Aspergillus fumigatus*, Birch Tree, Cat Dander, Cockroach (German), Cottonwood Tree, *D. farinae* (mites), *D. Pteronyssinus* (mites), Dog Dander, *Hormodendrum (Cladosporium)*, Mouse Epithelium, *Mucor racemosus*, Mugwort, *Penicillium notatum*, Sheep Sorrel (Dock), Timothy Grass, and IgE Serum Total.

**CPT Code(s):** 86003 x17; 82785

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

**New Test** [3001729](#) **Allergen, Region 19 Respiratory Panel IgE** **REG19PANEL**  
[Click for Pricing](#)

**Methodology:** Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay  
**Performed:** Sun-Sat  
**Reported:** 1-2 days

**Specimen Required:** Patient Prep: Multiple patient encounters should be avoided.  
Collect: Serum Separator Tube (SST). Multiple specimen tubes should be avoided.  
Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 4 mL serum to an ARUP Standard Transport Tube. (Min: 1.2 mL)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Hemolyzed, icteric, or lipemic specimens.  
Stability (collection to initiation of testing): After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

**Reference Interval:**

**Reference Intervals for all Components**

Reporting Range (reported in kU/L)	Probability of IgE Mediated Clinical Reaction	Class Scoring
Less than 0.10	No significant level detected	0
0.10-0.34	Clinical relevance undetermined	0/1
0.35-0.70	Low	1
0.71-3.50	Moderate	2
3.51-17.50	High	3
17.51 - 50.00	Very High	4
50.01 - 100.00	Very high	5
Greater than 100.00	Very high	6

Test Number	Components	Reference Interval	
0050345	Immunoglobulin E	Effective November 17, 2014	
		Age	Reference Interval
		0-5 months	13 kU/L or less
		6-12 months	34 kU/L or less
		1-2 years	97 kU/L or less
		3 years	199 kU/L or less
		4-6 years	307 kU/L or less
		7-8 years	403 kU/L or less
		9-12 years	696 kU/L or less
		13-15 years	629 kU/L or less
		16-17 years	537 kU/L or less
		18 years and older	214 kU/L or less

**Interpretive Data:** Allergen results of 0.10-0.34 kU/L are intended for specialist use as the clinical relevance is undetermined. Even though increasing ranges are reflective of increasing concentrations of allergen-specific IgE, these concentrations may not correlate with the degree of clinical response or skin testing results when challenged with a specific allergen. The correlation of allergy laboratory results with clinical history and in vivo reactivity to specific allergens is essential. A negative test may not rule out clinical allergy or even anaphylaxis.

**Note:** Allergens included in this panel: *Alternaria alternata* (tenuis), *Aspergillus fumigatus*, *Aureobasidium*, Bent/Redtop, *B. tropicalis*, Bermuda Grass, Cat Dander, Cockroach (German), *D. pteronyssinus* (mites), *D. farinae* (mites), Dog Dander, Elm Tree, Eucalyptus Tree, *Hormodendrum* (*Cladosporium*), Johnson Grass, Mouse Epithelium, *Mucor racemosus*, Oak Tree, *Penicillium notatum*, Pigweed, Pine/Australian Pine, Sheep Sorrel (Dock), Wall Pellitory, and IgE Serum Total.

**CPT Code(s):** 86003 x23; 82785

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

**New Test**     **3001712**     **Allergen, Region 2 Respiratory Panel IgE**     **REG2PANEL**  
[Click for Pricing](#)

**Methodology:** Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay  
**Performed:** Sun-Sat  
**Reported:** 1-2 days

**Specimen Required:** Patient Prep: Multiple patient encounters should be avoided.  
Collect: Serum Separator Tube (SST). Multiple specimen tubes should be avoided.  
Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 4 mL serum to an ARUP Standard Transport Tube. (Min: 1.3 mL)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Hemolyzed, icteric, or lipemic specimens.  
Stability (collection to initiation of testing): After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

**Reference Interval:**

**Allergens, Respiratory Panel, Region 2, Mid-Atlantic (DE, MD, VA, DC, NC) IgE**  
**Reference Intervals for all Components**

Reporting Range (reported in kU/L)	Probability of IgE Mediated Clinical Reaction	Class Scoring
Less than 0.10	No significant level detected	0
0.10-0.34	Clinical relevance undetermined	0/1
0.35-0.70	Low	1
0.71-3.50	Moderate	2
3.51-17.50	High	3
17.51 - 50.00	Very High	4
50.01 - 100.00	Very high	5
Greater than 100.00	Very high	6

Test Number	Components	Reference Interval
0050345	Immunoglobulin E	Effective November 17, 2014
		<b>Age</b>
		<b>Reference Interval</b>
		0-5 months
		13 kU/L or less
		6-12 months
		34 kU/L or less
		1-2 years
		97 kU/L or less
		3 years
		199 kU/L or less
		4-6 years
		307 kU/L or less
		7-8 years
		403 kU/L or less
		9-12 years
		696 kU/L or less
		13-15 years
		629 kU/L or less
		16-17 years
		537 kU/L or less
		18 years and older
		214 kU/L or less

**Interpretive Data:** Allergen results of 0.10-0.34 kU/L are intended for specialist use as the clinical relevance is undetermined. Even though increasing ranges are reflective of increasing concentrations of allergen-specific IgE, these concentrations may not correlate with the degree of clinical response or skin testing results when challenged with a specific allergen. The correlation of allergy laboratory results with clinical history and in vivo reactivity to specific allergens is essential. A negative test may not rule out clinical allergy or even anaphylaxis.

**Note:** Allergens included in this panel: *Alternaria alternata* (*tenuis*), *Aspergillus fumigatus*, Bermuda Grass, Birch Tree, Box Elder/Maple Tree, Cat Dander, Cockroach (German), Common Short Ragweed, Cottonwood Tree, *D. pteronyssinus* (mites), *D. farinae* (mites), Dog Dander, Elm Tree, *Hormodendrum* (*Cladosporium*), Johnson Grass, Mouse Epithelium, Mountain Cedar (Juniper) Tree, *Mucor racemosus*, Oak Tree, Pecan Tree, *Penicillium notatum*, Pigweed, Sheep Sorrel (Dock), Timothy Grass, White Mulberry Tree, and IgE Serum Total.

**CPT Code(s):** 86003 x25; 82785

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

**New Test** [3001730](#) **Allergen, Region 20 Respiratory Panel IgE** **REG20PANEL**  
[Click for Pricing](#)

**Methodology:** Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay  
**Performed:** Sun-Sat  
**Reported:** 1-2 days

**Specimen Required:** Patient Prep: Multiple patient encounters should be avoided.  
Collect: Serum Separator Tube (SST). Multiple specimen tubes should be avoided.  
Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 4 mL serum to an ARUP Standard Transport Tube. (Min: 1.3 mL)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Hemolyzed, icteric, or lipemic specimens.  
Stability (collection to initiation of testing): After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

**Reference Interval:**

**Allergens, Respiratory Panel, Region 20, Hawaii IgE**  
**Reference Intervals for all Components**

Reporting Range (reported in kU/L)	Probability of IgE Mediated Clinical Reaction	Class Scoring
Less than 0.10	No significant level detected	0
0.10-0.34	Clinical relevance undetermined	0/1
0.35-0.70	Low	1
0.71-3.50	Moderate	2
3.51-17.50	High	3
17.51 - 50.00	Very High	4
50.01 -100.00	Very high	5
Greater than 100.00	Very high	6

Test Number	Components	Reference Interval
0050345	Immunoglobulin E	Effective November 17, 2014
		<b>Age</b>
		<b>Reference Interval</b>
		0-5 months
		13 kU/L or less
		6-12 months
		34 kU/L or less
		1-2 years
		97 kU/L or less
		3 years
		199 kU/L or less
		4-6 years
		307 kU/L or less
		7-8 years
		403 kU/L or less
		9-12 years
		696 kU/L or less
		13-15 years
		629 kU/L or less
		16-17 years
		537 kU/L or less
		18 years and older
		214 kU/L or less

**Interpretive Data:** Allergen results of 0.10-0.34 kU/L are intended for specialist use as the clinical relevance is undetermined. Even though increasing ranges are reflective of increasing concentrations of allergen-specific IgE, these concentrations may not correlate with the degree of clinical response or skin testing results when challenged with a specific allergen. The correlation of allergy laboratory results with clinical history and in vivo reactivity to specific allergens is essential. A negative test may not rule out clinical allergy or even anaphylaxis.

**Note:** Allergens included in this panel: *Alternaria alternata* (tenuis), *Aspergillus fumigatus*, Bahia Grass, Bayberry Tree, Bermuda Grass, Cat Dander, Cocklebur, Cockroach (German), Common Short Ragweed, *D. pteronyssinus* (mites), *D. farinae* (mites), Dog Dander, English Plantain, Eucalyptus Tree, *Hormodendrum* (*Cladosporium*), Mesquite Tree, Mouse Epithelium, *Mucor racemosus*, Palm Tree, *Penicillium notatum*, Perennial Rye Grass, Pigweed, Pine/Australian Pine, Sagebrush/Wormwood, White Mulberry Tree, and IgE Serum Total.

**CPT Code(s):** 86003 x25; 82785

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

**New Test** [3001713](#) **Allergen, Region 3 Respiratory Panel IgE** **REG3PANEL**  
[Click for Pricing](#)

**Methodology:** Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay  
**Performed:** Sun-Sat  
**Reported:** 1-2 days

**Specimen Required:** Patient Prep: Multiple patient encounters should be avoided.  
Collect: Serum Separator Tube (SST). Multiple specimen tubes should be avoided.  
Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 4 mL serum to an ARUP Standard Transport Tube. (Min: 1.3 mL)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Hemolyzed, icteric, or lipemic specimens.  
Stability (collection to initiation of testing): After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

**Reference Interval:**

**Reference Intervals for all Components**

Reporting Range (reported in kU/L)	Probability of IgE Mediated Clinical Reaction	Class Scoring
Less than 0.10	No significant level detected	0
0.10-0.34	Clinical relevance undetermined	0/1
0.35-0.70	Low	1
0.71-3.50	Moderate	2
3.51-17.50	High	3
17.51 - 50.00	Very High	4
50.01 - 100.00	Very high	5
Greater than 100.00	Very high	6

Test Number	Components	Reference Interval
0050345	Immunoglobulin E	Effective November 17, 2014
		<b>Age</b>
		<b>Reference Interval</b>
		0-5 months
		13 kU/L or less
		6-12 months
		34 kU/L or less
		1-2 years
		97 kU/L or less
		3 years
		199 kU/L or less
		4-6 years
		307 kU/L or less
		7-8 years
		403 kU/L or less
		9-12 years
		696 kU/L or less
		13-15 years
		629 kU/L or less
		16-17 years
		537 kU/L or less
		18 years and older
		214 kU/L or less

**Interpretive Data:** Allergen results of 0.10-0.34 kU/L are intended for specialist use as the clinical relevance is undetermined. Even though increasing ranges are reflective of increasing concentrations of allergen-specific IgE, these concentrations may not correlate with the degree of clinical response or skin testing results when challenged with a specific allergen. The correlation of allergy laboratory results with clinical history and in vivo reactivity to specific allergens is essential. A negative test may not rule out clinical allergy or even anaphylaxis.

**Note:** Allergens included in this panel: *Alternaria alternata* (tenuis), *Aspergillus fumigatus*, Bahia Grass, Bermuda Grass, Birch Tree, Box Elder/Maple Tree, Cat Dander, Cockroach (German), Common Short Ragweed, *D. pteronyssinus* (mites), *D. farinae* (mites), Dog Dander, Elm Tree, *Hormodendrum* (*Cladosporium*), Mouse Epithelium, Mountain Cedar (Juniper) Tree, *Mucor racemosus*, Nettle, Oak Tree, Pecan (White Hickory) Tree, *Penicillium notatum*, Pigweed, Sheep Sorrel (Dock), Timothy Grass, and IgE Serum Total.

**CPT Code(s):** 86003 x24; 82785

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

**New Test** [3001714](#) **Allergen, Region 4 Respiratory Panel IgE** **REG4PANEL**  
[Click for Pricing](#)

**Methodology:** Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay  
**Performed:** Sun-Sat  
**Reported:** 1-2 days

**Specimen Required:** Patient Prep: Multiple patient encounters should be avoided.  
Collect: Serum Separator Tube (SST). Multiple specimen tubes should be avoided.  
Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 4 mL serum to an ARUP Standard Transport Tube. (Min: 1.3 mL)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Hemolyzed, icteric, or lipemic specimens.  
Stability (collection to initiation of testing): After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

**Reference Interval:**

**Reference Intervals for all Components**

Reporting Range (reported in kU/L)	Probability of IgE Mediated Clinical Reaction	Class Scoring
Less than 0.10	No significant level detected	0
0.10-0.34	Clinical relevance undetermined	0/1
0.35-0.70	Low	1
0.71-3.50	Moderate	2
3.51-17.50	High	3
17.51 - 50.00	Very High	4
50.01 - 100.00	Very high	5
Greater than 100.00	Very high	6

Test Number	Components	Reference Interval	
0050345	Immunoglobulin E	Effective November 17, 2014	
		Age	Reference Interval
		0-5 months	13 kU/L or less
		6-12 months	34 kU/L or less
		1-2 years	97 kU/L or less
		3 years	199 kU/L or less
		4-6 years	307 kU/L or less
		7-8 years	403 kU/L or less
		9-12 years	696 kU/L or less
		13-15 years	629 kU/L or less
		16-17 years	537 kU/L or less
		18 years and older	214 kU/L or less

**Interpretive Data:** Allergen results of 0.10-0.34 kU/L are intended for specialist use as the clinical relevance is undetermined. Even though increasing ranges are reflective of increasing concentrations of allergen-specific IgE, these concentrations may not correlate with the degree of clinical response or skin testing results when challenged with a specific allergen. The correlation of allergy laboratory results with clinical history and in vivo reactivity to specific allergens is essential. A negative test may not rule out clinical allergy or even anaphylaxis.

**Note:** Allergens included in this panel: *Alternaria alternata* (*tenuis*), *Aspergillus fumigatus*, *B. tropicalis*, Bahia Grass, Bermuda Grass, Box Elder/Maple Tree, Cat Dander, Cockroach (German), Common Short Ragweed, *D. pteronyssinus* (mites), *D. farinae* (mites), Dog Dander, Elm Tree, *Hormodendrum* (*Cladosporium*), Mouse Epithelium, Mountain Cedar (Juniper) Tree, *Mucor racemosus*, Nettle, Oak Tree, *Penicillium notatum*, Pigweed, Pine/Australian Pine, Sheep Sorrel (Dock), Timothy Grass, and IgE Serum Total.

**CPT Code(s):** 86003 x24; 82785

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.



HOTLINE: Effective August 19, 2019

**New Test**     [3001715](#)     **Allergen, Region 5 Respiratory Panel IgE**     **REG5PANEL**  
[Click for Pricing](#)

**Methodology:** Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay  
**Performed:** Sun-Sat  
**Reported:** 1-2 days

**Specimen Required:** Patient Prep: Multiple patient encounters should be avoided.  
Collect: Serum Separator Tube (SST). Multiple specimen tubes should be avoided.  
Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 4 mL serum to an ARUP Standard Transport Tube. (Min: 1.4 mL)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Hemolyzed, icteric, or lipemic specimens.  
Stability (collection to initiation of testing): After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

**Reference Interval:**

**Reference Intervals for all Components**

Reporting Range (reported in kU/L)	Probability of IgE Mediated Clinical Reaction	Class Scoring
Less than 0.10	No significant level detected	0
0.10-0.34	Clinical relevance undetermined	0/1
0.35-0.70	Low	1
0.71-3.50	Moderate	2
3.51-17.50	High	3
17.51 - 50.00	Very High	4
50.01 - 100.00	Very high	5
Greater than 100.00	Very high	6

Test Number	Components	Reference Interval	
0050345	Immunoglobulin E	Effective November 17, 2014	
		Age	Reference Interval
		0-5 months	13 kU/L or less
		6-12 months	34 kU/L or less
		1-2 years	97 kU/L or less
		3 years	199 kU/L or less
		4-6 years	307 kU/L or less
		7-8 years	403 kU/L or less
		9-12 years	696 kU/L or less
		13-15 years	629 kU/L or less
		16-17 years	537 kU/L or less
		18 years and older	214 kU/L or less

**Interpretive Data:** Allergen results of 0.10-0.34 kU/L are intended for specialist use as the clinical relevance is undetermined. Even though increasing ranges are reflective of increasing concentrations of allergen-specific IgE, these concentrations may not correlate with the degree of clinical response or skin testing results when challenged with a specific allergen. The correlation of allergy laboratory results with clinical history and in vivo reactivity to specific allergens is essential. A negative test may not rule out clinical allergy or even anaphylaxis.

**Note:** Allergens included in this panel: *Alternaria alternata* (*tenuis*), *Aspergillus fumigatus*, Bermuda Grass, Birch Tree, Box Elder/Maple Tree, Cat Dander, Cockroach (German), Common Short Ragweed, Cottonwood Tree, *D. farinae* (mites), *D. Pteronyssinus* (mites), Dog Dander, Elm Tree, *Hormodendrum* (*Cladosporium*), Mouse Epithelium, Mountain Cedar (Juniper) Tree, *Mucor racemosus*, Oak Tree, Pecan (White Hickory) Tree, *Penicillium notatum*, Pigweed, Russian Thistle, Sheep Sorrel (Dock), Sycamore Tree, Timothy Grass, Walnut Tree, White Ash Tree, White Mulberry Tree, and IgE Serum Total.

**CPT Code(s):** 86003 x28; 82785

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

**New Test** [3001716](#) **Allergen, Region 6 Respiratory Panel IgE** **REG6PANEL**  
[Click for Pricing](#)

**Methodology:** Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay  
**Performed:** Sun-Sat  
**Reported:** 1-2 days

**Specimen Required:** Patient Prep: Multiple patient encounters should be avoided.  
Collect: Serum Separator Tube (SST). Multiple specimen tubes should be avoided.  
Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 4 mL serum to an ARUP Standard Transport Tube. (Min: 1.3 mL)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Hemolyzed, icteric, or lipemic specimens.  
Stability (collection to initiation of testing): After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

**Reference Interval:**

**Reference Intervals for all Components**

Reporting Range (reported in kU/L)	Probability of IgE Mediated Clinical Reaction	Class Scoring
Less than 0.10	No significant level detected	0
0.10-0.34	Clinical relevance undetermined	0/1
0.35-0.70	Low	1
0.71-3.50	Moderate	2
3.51-17.50	High	3
17.51 - 50.00	Very High	4
50.01 - 100.00	Very high	5
Greater than 100.00	Very high	6

Test Number	Components	Reference Interval	
0050345	Immunoglobulin E	Effective November 17, 2014	
		Age	Reference Interval
		0-5 months	13 kU/L or less
		6-12 months	34 kU/L or less
		1-2 years	97 kU/L or less
		3 years	199 kU/L or less
		4-6 years	307 kU/L or less
		7-8 years	403 kU/L or less
		9-12 years	696 kU/L or less
		13-15 years	629 kU/L or less
		16-17 years	537 kU/L or less
		18 years and older	214 kU/L or less

**Interpretive Data:** Allergen results of 0.10-0.34 kU/L are intended for specialist use as the clinical relevance is undetermined. Even though increasing ranges are reflective of increasing concentrations of allergen-specific IgE, these concentrations may not correlate with the degree of clinical response or skin testing results when challenged with a specific allergen. The correlation of allergy laboratory results with clinical history and in vivo reactivity to specific allergens is essential. A negative test may not rule out clinical allergy or even anaphylaxis.

**Note:** Allergens included in this panel: *Alternaria alternata* (*tenuis*), *Aspergillus fumigatus*, Bermuda Grass, Birch Tree, Box Elder/Maple Tree, Cat Dander, Cockroach (German), Common Short Ragweed, *D. farinae* (mites), *D. Pteronyssinus* (mites), Dog Dander, Elm Tree, *Hormodendrum* (*Cladosporium*), Marsh Elder, Mouse Epithelium, Mountain Cedar (Juniper) Tree, *Mucor racemosus*, Oak Tree, Pecan (White Hickory) Tree, *Penicillium notatum*, Pigweed, Timothy Grass, Walnut Tree, White Mulberry Tree, and IgE Serum Total.

**CPT Code(s):** 86003 x24; 82785

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

**New Test** [3001717](#)  
[Click for Pricing](#)

**Allergen, Region 7 Respiratory Panel IgE**

**REG7PANEL**

**Methodology:** Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay  
**Performed:** Sun-Sat  
**Reported:** 1-2 days

**Specimen Required:** Patient Prep: Multiple patient encounters should be avoided.  
Collect: Serum Separator Tube (SST). Multiple specimen tubes should be avoided.  
Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 4 mL serum to an ARUP Standard Transport Tube. (Min: 1.3 mL)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Hemolyzed, icteric, or lipemic specimens.  
Stability (collection to initiation of testing): After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

**Reference Interval:**

**Reference Intervals for all Components**

Reporting Range (reported in kU/L)	Probability of IgE Mediated Clinical Reaction	Class Scoring
Less than 0.10	No significant level detected	0
0.10-0.34	Clinical relevance undetermined	0/1
0.35-0.70	Low	1
0.71-3.50	Moderate	2
3.51-17.50	High	3
17.51 - 50.00	Very High	4
50.01 - 100.00	Very high	5
Greater than 100.00	Very high	6

Test Number	Components	Reference Interval	
0050345	Immunoglobulin E	Effective November 17, 2014	
		Age	Reference Interval
		0-5 months	13 kU/L or less
		6-12 months	34 kU/L or less
		1-2 years	97 kU/L or less
		3 years	199 kU/L or less
		4-6 years	307 kU/L or less
		7-8 years	403 kU/L or less
		9-12 years	696 kU/L or less
		13-15 years	629 kU/L or less
		16-17 years	537 kU/L or less
		18 years and older	214 kU/L or less

**Interpretive Data:** Allergen results of 0.10-0.34 kU/L are intended for specialist use as the clinical relevance is undetermined. Even though increasing ranges are reflective of increasing concentrations of allergen-specific IgE, these concentrations may not correlate with the degree of clinical response or skin testing results when challenged with a specific allergen. The correlation of allergy laboratory results with clinical history and in vivo reactivity to specific allergens is essential. A negative test may not rule out clinical allergy or even anaphylaxis.

**Note:** Allergens included in this panel: *Alternaria alternata* (*tenuis*), *Aspergillus fumigatus*, Bermuda Grass, Birch Tree, Box Elder/Maple Tree, Cat Dander, Cockroach (German), Common Short Ragweed, Cottonwood Tree, *D. farinae* (mites), *D. Pteronyssinus* (mites), Dog Dander, Elm Tree, *Hormodendrum* (*Cladosporium*), Marsh Elder, Mouse Epithelium, Mountain Cedar (Juniper) Tree, *Mucor racemosus*, Nettle, Oak Tree, *Penicillium notatum*, Russian Thistle, Timothy Grass, White Ash Tree, White Mulberry Tree, and IgE Serum Total.

**CPT Code(s):** 86003 x25; 82785

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

**New Test**     [3001718](#)     **Allergen, Region 8 Respiratory Panel IgE**     **REG8PANEL**  
[Click for Pricing](#)

**Methodology:** Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay  
**Performed:** Sun-Sat  
**Reported:** 1-2 days

**Specimen Required:** Patient Prep: Multiple patient encounters should be avoided  
Collect: Serum Separator Tube (SST). Multiple specimen tubes should be avoided.  
Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 4 mL serum to an ARUP Standard Transport Tube. (Min: 1.4 mL)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Hemolyzed, icteric, or lipemic specimens.  
Stability (collection to initiation of testing): After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

**Reference Interval:**

**Reference Intervals for all Components**

Reporting Range (reported in kU/L)	Probability of IgE Mediated Clinical Reaction	Class Scoring
Less than 0.10	No significant level detected	0
0.10-0.34	Clinical relevance undetermined	0/1
0.35-0.70	Low	1
0.71-3.50	Moderate	2
3.51-17.50	High	3
17.51 - 50.00	Very High	4
50.01 - 100.00	Very high	5
Greater than 100.01	Very high	6

Test Number	Components	Reference Interval	
0050345	Immunoglobulin E	Effective November 17, 2014	
		Age	Reference Interval
		0-5 months	13 kU/L or less
		6-12 months	34 kU/L or less
		1-2 years	97 kU/L or less
		3 years	199 kU/L or less
		4-6 years	307 kU/L or less
		7-8 years	403 kU/L or less
		9-12 years	696 kU/L or less
		13-15 years	629 kU/L or less
		16-17 years	537 kU/L or less
		18 years and older	214 kU/L or less

**Interpretive Data:** Allergen results of 0.10-0.34 kU/L are intended for specialist use as the clinical relevance is undetermined. Even though increasing ranges are reflective of increasing concentrations of allergen-specific IgE, these concentrations may not correlate with the degree of clinical response or skin testing results when challenged with a specific allergen. The correlation of allergy laboratory results with clinical history and in vivo reactivity to specific allergens is essential. A negative test may not rule out clinical allergy or even anaphylaxis.

**Note:** Allergens included in this panel: *Alternaria alternata* (*tenuis*), *Aspergillus fumigatus*, Bermuda Grass, Box Elder/Maple Tree, Cat Dander, Cockroach (German), Common Short Ragweed, Cottonwood Tree, *D. farinae* (mites), *D. Pteronyssinus* (mites), Dog Dander, Elm Tree, *Hormodendrum* (*Cladosporium*), Marsh Elder, Mouse Epithelium, Mountain Cedar (Juniper) Tree, *Mucor racemosus*, Oak Tree, Pecan (White Hickory) Tree, *Penicillium notatum*, Pigweed, Russian Thistle, Sycamore Tree, Timothy Grass, Walnut Tree, White Ash Tree, White Mulberry Tree, and IgE Serum Total.

**CPT Code(s):** 86003 x27; 82785

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

**New Test** [3001719](#) **Allergen, Region 9 Respiratory Panel IgE** **REG9PANEL**  
[Click for Pricing](#)

**Methodology:** Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay  
**Performed:** Sun-Sat  
**Reported:** 1-2 days

**Specimen Required:** Patient Prep: Multiple patient encounters should be avoided.  
Collect: Serum Separator Tube (SST). Multiple specimen tubes should be avoided.  
Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 4 mL serum to an ARUP Standard Transport Tube. (Min: 1.3 mL)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Hemolyzed, icteric, or lipemic specimens.  
Stability (collection to initiation of testing): After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

**Reference Interval:**

**Reference Intervals for all Components**

Reporting Range (reported in kU/L)	Probability of IgE Mediated Clinical Reaction	Class Scoring
Less than 0.10	No significant level detected	0
0.10-0.34	Clinical relevance undetermined	0/1
0.35-0.70	Low	1
0.71-3.50	Moderate	2
3.51-17.50	High	3
17.51 - 50.00	Very High	4
50.01 - 100.00	Very high	5
Greater than 100.00	Very high	6

Test Number	Components	Reference Interval	
0050345	Immunoglobulin E	Effective November 17, 2014	
		Age	Reference Interval
		0-5 months	13 kU/L or less
		6-12 months	34 kU/L or less
		1-2 years	97 kU/L or less
		3 years	199 kU/L or less
		4-6 years	307 kU/L or less
		7-8 years	403 kU/L or less
		9-12 years	696 kU/L or less
		13-15 years	629 kU/L or less
		16-17 years	537 kU/L or less
		18 years and older	214 kU/L or less

**Interpretive Data:** Allergen results of 0.10-0.34 kU/L are intended for specialist use as the clinical relevance is undetermined. Even though increasing ranges are reflective of increasing concentrations of allergen-specific IgE, these concentrations may not correlate with the degree of clinical response or skin testing results when challenged with a specific allergen. The correlation of allergy laboratory results with clinical history and in vivo reactivity to specific allergens is essential. A negative test may not rule out clinical allergy or even anaphylaxis.

**Note:** Allergens included in this panel: *Alternaria alternata* (*tenuis*), *Aspergillus fumigatus*, Bermuda Grass, Box Elder/Maple Tree, Cat Dander, Cockroach (German), Common Short Ragweed, Cottonwood Tree, *D. farinae* (mites), *D. Pteronyssinus* (mites), Dog Dander, Elm Tree, *Hormodendrum* (*Cladosporium*), Mouse Epithelium, Mountain Cedar (Juniper) Tree, *Mucor racemosus*, Nettle, Oak Tree, *Penicillium notatum*, Russian Thistle, Sheep Sorrel (Dock), Timothy Grass, White Ash Tree, White Mulberry Tree, and IgE Serum Total.

**CPT Code(s):** 86003 x24; 82785

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

**New Test** [3001726](#) **Allergen, Region 16 Respiratory Panel IgE** **REG16PANEL**  
[Click for Pricing](#)

**Methodology:** Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay  
**Performed:** Sun-Sat  
**Reported:** 1-2 days

**Specimen Required:** Patient Prep: Multiple patient encounters should be avoided.  
Collect: Serum Separator Tube (SST). Multiple specimen tubes should be avoided.  
Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 4 mL serum to an ARUP Standard Transport Tube. (Min: 1.2 mL)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Hemolyzed, icteric, or lipemic specimens.  
Stability (collection to initiation of testing): After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

**Reference Interval:**

**Reference Intervals for all Components**

Reporting Range (reported in kU/L)	Probability of IgE Mediated Clinical Reaction	Class Scoring
Less than 0.10	No significant level detected	0
0.10-0.34	Clinical relevance undetermined	0/1
0.35-0.70	Low	1
0.71-3.50	Moderate	2
3.51-17.50	High	3
17.51 - 50.00	Very High	4
50.01 - 100.00	Very high	5
Greater than 100.00	Very high	6

Test Number	Components	Reference Interval	
0050345	Immunoglobulin E	Effective November 17, 2014	
		Age	Reference Interval
		0-5 months	13 kU/L or less
		6-12 months	34 kU/L or less
		1-2 years	97 kU/L or less
		3 years	199 kU/L or less
		4-6 years	307 kU/L or less
		7-8 years	403 kU/L or less
		9-12 years	696 kU/L or less
		13-15 years	629 kU/L or less
		16-17 years	537 kU/L or less
		18 years and older	214 kU/L or less

**Interpretive Data:** Allergen results of 0.10-0.34 kU/L are intended for specialist use as the clinical relevance is undetermined. Even though increasing ranges are reflective of increasing concentrations of allergen-specific IgE, these concentrations may not correlate with the degree of clinical response or skin testing results when challenged with a specific allergen. The correlation of allergy laboratory results with clinical history and in vivo reactivity to specific allergens is essential. A negative test may not rule out clinical allergy or even anaphylaxis.

**Note:** Allergens included in this panel: Alder Tree, *Alternaria alternata (tenuis)*, *Aspergillus fumigatus*, Birch Tree, BoxElder/Maple Tree, Cat Dander, Cockroach (German), Cottonwood Tree, *D. farinae* (mites), *D. Pteronyssinus* (mites), Dog Dander, Elm Tree, *Hormodendrum (Cladosporium)*, Mouse Epithelium, Mountain Cedar (Juniper) Tree, *Mucor racemosus*, Mugwort, Oak Tree, *Penicillium notatum*, Pigweed, Russian Thistle, Sheep Sorrel (Dock), Timothy Grass, and IgE Serum Total.

**CPT Code(s):** 86003 x23; 82785

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

**New Test** [3001727](#) **Allergen, Region 17 Respiratory Panel IgE** **REG17PANEL**  
[Click for Pricing](#)

**Methodology:** Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay  
**Performed:** Sun-Sat  
**Reported:** 1-2 days

**Specimen Required:** Patient Prep: Multiple patient encounters should be avoided.  
Collect: Serum Separator Tube (SST). Multiple specimen tubes should be avoided.  
Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 4 mL serum to an ARUP Standard Transport Tube. (Min: 1.3 mL)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Hemolyzed, icteric, or lipemic specimens.  
Stability (collection to initiation of testing): After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

**Reference Interval:**

**Reference Intervals for all Components**

Reporting Range (reported in kU/L)	Probability of IgE Mediated Clinical Reaction	Class Scoring
Less than 0.10	No significant level detected	0
0.10-0.34	Clinical relevance undetermined	0/1
0.35-0.70	Low	1
0.71-3.50	Moderate	2
3.51-17.50	High	3
17.51 - 50.00	Very High	4
50.01 - 100.00	Very high	5
Greater than 100.00	Very high	6

Test Number	Components	Reference Interval	
0050345	Immunoglobulin E	Effective November 17, 2014	
		Age	Reference Interval
		0-5 months	13 kU/L or less
		6-12 months	34 kU/L or less
		1-2 years	97 kU/L or less
		3 years	199 kU/L or less
		4-6 years	307 kU/L or less
		7-8 years	403 kU/L or less
		9-12 years	696 kU/L or less
		13-15 years	629 kU/L or less
		16-17 years	537 kU/L or less
		18 years and older	214 kU/L or less

**Interpretive Data:** Allergen results of 0.10-0.34 kU/L are intended for specialist use as the clinical relevance is undetermined. Even though increasing ranges are reflective of increasing concentrations of allergen-specific IgE, these concentrations may not correlate with the degree of clinical response or skin testing results when challenged with a specific allergen. The correlation of allergy laboratory results with clinical history and in vivo reactivity to specific allergens is essential. A negative test may not rule out clinical allergy or even anaphylaxis.

**Note:** Allergens included in this panel: Alder Tree, *Alternaria alternata (tenuis)*, *Aspergillus fumigatus*, Birch Tree, BoxElder/Maple Tree, Cat Dander, Cockroach (German), Common Short Ragweed, Cottonwood Tree, *D. farinae* (mites), *D. Pteronyssinus* (mites), Dog Dander, Elm Tree, *Hormodendrum (Cladosporium)*, Mouse Epithelium, Mountain Cedar (Juniper) Tree, *Mucor racemosus*, Nettle, Oak Tree, *Penicillium notatum*, Pigweed, Sheep Sorrel (Dock), Timothy Grass, Walnut Tree, White Ash Tree and IgE Serum Total.

**CPT Code(s):** 86003 x25; 82785

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

[0097933](#) **Allergen, Tree, Oak Red IgE** **OAK RED**

**Methodology:** Quantitative Enzyme Immunoassay



HOTLINE: Effective August 19, 2019

**0051495**

**Alpha Thalassemia (HBA1 and HBA2) 7 Deletions**

**ALPHA THAL**



Additional Technical Information



Out of Pocket Estimator

**Performed:** Varies  
**Reported:** 7-10 days

**Specimen Required:** Collect: Lavender (EDTA), Pink (K<sub>2</sub>EDTA), or Yellow (ACD Solution A or B).  
Specimen Preparation: Transport 3 mL whole blood. (Min: 1 mL)  
Storage/Transport Temperature: Refrigerated.  
Stability (collection to initiation of testing): Ambient: 1 week; Refrigerated: 1 month; Frozen: 6 months

**2006480**

**ANCA-Associated Vasculitis Profile (ANCA/MPO/PR3) with Reflex to ANCA Titer**

**ANCA PRO**

**Reference Interval:**

Test Number	Components	Reference Interval	
	Anti-Neutrophil Cytoplasmic Antibody, IgG	< 1:20; Not significant	
	Myeloperoxidase Antibody	Negative	19 AU/mL or less
		Equivocal	20-25 AU/mL
		Positive	26 AU/mL or greater
	Serine Proteinase 3 Antibody	Negative	19 AU/mL or less
		Equivocal	20-25 AU/mL
		Positive	26 AU/mL or greater

**Interpretive Data:** Neutrophil cytoplasmic antibodies (C-ANCA = granular cytoplasmic staining, P-ANCA = perinuclear staining) are found in the serum of over 90 percent of patients with certain necrotizing systemic vasculitides, and usually in less than 5 percent of patients with collagen vascular disease or arthritis. Approximately 90 percent of patients with a P-ANCA pattern by IFA have antibodies specific for MPO. Approximately 85 percent of patients with a C-ANCA pattern by IFA have antibodies specific for PR3.

**Note:** Specimens are screened for ANCA, MPO and PR3. If the ANCA screen detects antibodies at a 1:20 dilution or greater, then a titer to end point will be added. Additional charges apply.

ANCA IFA is simultaneously tested on ethanol- and formalin-fixed slides to allow differentiation of C- and P-ANCA patterns.

**HOTLINE NOTE:** There is a clinically significant charting name change associated with this test.  
Change the charting name for component 0050527, Serine Protease 3, IgG to Serine Proteinase 3, IgG

**0010004**

**Antibody Detection RBC with Reflex to ID**

**IRL-ABSC**

**Specimen Required:** Collect: Plain Red AND Lavender (K<sub>2</sub>EDTA) or Pink (K<sub>2</sub>EDTA).  
Specimen Preparation: Do not freeze.  
Transport 10 mL whole blood (Plain Red) AND 5 mL whole blood (EDTA). (Min: 7 mL (Plain Red) and 3 mL (EDTA))  
Pediatric: Transport 1 mL whole blood (Plain Red) AND 0.5 mL whole blood (EDTA). (Min: 1 mL (Plain Red) and 0.5 mL (EDTA))  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Separator tubes.  
Stability (collection to initiation of testing): Ambient: Unacceptable; Refrigerated: 1 week; Frozen: Unacceptable

**Note:** Panel identification will be performed on all positive specimens at an additional charge.  
Positive screens are reflexed:  
Female 15-45 years are reflexed to Antibody Identification, RBC (Prenatal Only) (ARUP test code 0013005)  
All other Positive Screens are reflexed to Antibody ID Package (IRL) (ARUP test code 0013003)

**CPT Code(s):** 86850; additional CPT codes may apply

HOTLINE: Effective August 19, 2019

**0013003**

**Antibody ID Package (IRL)**

**IRL-AB PKG**

**Performed:** Mon-Fri  
**Reported:** 2-5 days

**Specimen Required:** Collect: Lavender (K<sub>2</sub>EDTA) or Pink (K<sub>2</sub>EDTA) AND Plain Red.  
Specimen Preparation: **Do not freeze.** Transport 10 mL whole blood (plain red) AND 20 mL whole blood (EDTA). (Min: 7 mL plain red AND 10 mL EDTA)  
Storage/Transport Temperature: Refrigerated. Deliver to lab immediately.  
Unacceptable Conditions: Separator tubes.  
Stability (collection to initiation of testing): Ambient: **Unacceptable**; Refrigerated: 1 week; Frozen: Unacceptable

**0013005**

**Antibody ID RBC Prenatal-Reflex to Titer**

**IRL-ABID**

**Specimen Required:** Collect: 3 (7mL) Lavender (K<sub>2</sub>EDTA) or Pink (K<sub>2</sub>EDTA) AND 1 (7mL) Plain Red.  
Specimen Preparation: **Do not freeze.** Transport 3 (7 mL) whole blood EDTA AND 1 (7 mL) whole blood plain red. (Min: 10 mL EDTA and 3 mL Plain Red)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Separator tubes.  
Stability (collection to initiation of testing): Ambient: **Unacceptable**; Refrigerated: 1 week; Frozen: Unacceptable

**Note:** **This test is for prenatal patients only.** Includes: ABO/Rh Type, Rh Phenotype, Direct Coombs, RBC Antibody Identification by various methods. **Titers** will be performed, at an additional charge, on prenatal specimens for clinically significant antibodies.

**0013006**

**Antibody Titer**

**IRL-ABTR1**

**Performed:** Mon-Fri  
**Reported:** 2-5 days

**Specimen Required:** Collect: Lavender (K<sub>2</sub>EDTA) or Pink (K<sub>2</sub>EDTA).  
Specimen Preparation: **Do not freeze.** Transport 10 mL whole blood. (Min: 10 mL)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Separator tubes.  
Stability (collection to initiation of testing): Ambient: **Unacceptable**; Refrigerated: 1 week; Frozen: Unacceptable

**Note:** Antibody identification must be performed prior to performing this test. Additional charges apply. **Titer result is the inverse of the highest dilution that has a positive reaction.** Example: if the highest dilution is 1:16 then the titer result is 16.

**2002068**

**Anti-Neutrophil Cytoplasmic Antibody with Reflex to Titer and MPO/PR3 Antibodies**

**ANCA PN**

**Reference Interval:**

Test Number	Components	Reference Interval	
	Anti-Neutrophil Cytoplasmic Antibody, IgG	< 1:20: Not significant	
0050526	Myeloperoxidase Antibody	Negative	19 AU/mL or less
		Equivocal	20-25 AU/mL
		Positive	26 AU/mL or greater
0050527	Serine Proteinase 3 Antibody	Negative	19 AU/mL or less
		Equivocal	20-25 AU/mL
		Positive	26 AU/mL or greater

**Interpretive Data:** Neutrophil cytoplasmic antibodies (C-ANCA = granular cytoplasmic staining, P-ANCA = perinuclear staining) are found in the serum of over 90% of patients with certain necrotizing systemic vasculitides, and usually in less than 5% of patients with collagen vascular disease or arthritis. Approximately 90% of patients with a P-ANCA pattern by IFA have antibodies specific for MPO. Approximately 85% of patients with a C-ANCA pattern by IFA have antibodies specific for PR3.

**Note:** Specimens are screened by IFA on ethanol-fixed neutrophils, formalin-fixed neutrophils, and HEp-2 slides that allow differentiation of C- and P-ANCA patterns.

If screen is positive, then titer and MPO/PR3 antibodies will be added to aid in antibody determination. Additional charges apply.

HOTLINE: Effective August 19, 2019

**0050317**

**Antinuclear Antibodies (ANA), IgG by ELISA with Reflex to ANA Hep-2 Substrate, IgG by IFA and ENA Confirmation**

**ANA REF**

**Reference Interval:**

Effective May 18, 2015

Test Number	Components	Reference Interval					
	Antinuclear Antibodies (ANA), IgG by ELISA	None Detected					
3000082	Antinuclear Antibody (ANA) with HEp-2 Substrate, IgG by IFA	Less than 1:80					
	Double-Stranded DNA (dsDNA) Antibody, IgG by ELISA	None Detected					
2002693	Double-Stranded DNA (dsDNA) Antibody, IgG by IFA (using <i>Crithidia luciliae</i> )	Less than 1:10					
0050470	Smith/RNP (ENA) Antibody, IgG	29 AU/mL or less	Negative				
		30-40 AU/mL	Equivocal				
		41 AU/mL or greater	Positive				
0050085	Smith (ENA) Antibody, IgG	29 AU/mL or less	Negative				
		30-40 AU/mL	Equivocal				
		41 AU/mL or greater	Positive				
2012074	SSA 52 and 60 (Ro) (ENA) Antibodies, IgG	Test Number	Components	Reference Interval			
					SSA-52 (Ro52) (ENA) Antibody, IgG	29 AU/mL or Less	Negative
						30-40 AU/mL	Equivocal
		41 AU/mL or	Positive				
			SSA-60 (Ro60) (ENA) Antibody, IgG	29 AU/mL or Less	Negative		
				30-40 AU/mL	Equivocal		
				41 AU/mL or	Positive		
0050692	SSB (La) (ENA) Antibody, IgG	29 AU/mL or less	Negative				
		30-40 AU/mL	Equivocal				
		41 AU/mL or greater	Positive				
0099592	Jo-1 Antibody, IgG	29 AU/mL or less	Negative				
		30-40 AU/mL	Equivocal				
		41 AU/mL or greater	Positive				
0050599	Scleroderma (Scl-70) (ENA) Antibody, IgG	29 AU/mL or less	Negative				
		30-40 AU/mL	Equivocal				
		41 AU/mL or greater	Positive				

**Interpretive Data: Antinuclear Antibodies (ANA), IgG by ELISA:** ANA specimens are screened using enzyme-linked immunosorbent assay (ELISA) methodology. All ELISA results reported as detected are further tested by indirect fluorescent assay (IFA) using Hep-2 substrate with an IgG-specific conjugate. The ANA ELISA screen is designed to detect antibodies against dsDNA, histone, SS-A (Ro), SS-B (La), Smith, **Smith/RNP**, Scl-70, Jo-1, **centromeric proteins**, and **other antigens extracted from the Hep-2 cell nucleus**. ANA ELISA assays have been reported to have lower sensitivities than ANA IFA for systemic autoimmune rheumatic diseases (SARD).

Negative results do not necessarily rule out SARD.

**Note:** ANA lacks diagnostic specificity, and is associated with in variety diseases (cancers, autoimmune, infectious, and inflammatory conditions) and occurs in healthy individuals in varying prevalence. The lack of diagnostic specificity requires confirmation of positive ANA by more-specific serologic tests, which may be guided by the pattern(s) observed.

Specimens are screened for ANA using ELISA. If ANA IgG is detected by ELISA, then Antinuclear Antibody (ANA), Hep-2, IgG by IFA will be added. If ANA, IgG by IFA is confirmed positive with a titer of 1:80 or greater, then a titer and pattern will be reported. In addition, samples positive for ANA, IgG by IFA will reflex to Double-Stranded DNA (dsDNA) Antibody, IgG by ELISA; Jo-1 Antibody, IgG; **Smith/RNP (ENA)** Antibody, IgG; Scleroderma (Scl-70) (ENA) Antibody, IgG; **Smith** (ENA) Antibody, IgG; SSA 52 and 60 (Ro) (ENA) Antibodies, IgG; and SSB (La) (ENA) Antibody, IgG. If Double-Stranded DNA (dsDNA) Antibody, IgG by ELISA is detected, then Double-Stranded DNA (dsDNA) Antibody, IgG by IFA (using *Crithidia luciliae*) will be added. Additional charges apply.

ANA determined by indirect fluorescence assay (IFA) use Hep-2 substrate and IgG-specific conjugate at a screening dilution of 1:80. If positive, patterns reported include homogeneous, speckled, centromere, nucleolar, nuclear dots, or cytoplasmic. All positive results are reported with endpoint titers.

HOTLINE: Effective August 19, 2019

**0050080**

**Antinuclear Antibodies (ANA), IgG by ELISA with Reflex to ANA, HEp-2 Substrate, IgG by IFA**

**ANA**

**Interpretive Data:** Antinuclear Antibodies (ANA), IgG by ELISA: ANA specimens are screened using enzyme-linked immunosorbent assay (ELISA) methodology. All ELISA results reported as Detected are further tested by indirect fluorescent assay (IFA) using HEp-2 substrate with an IgG-specific conjugate. The ANA ELISA screen is designed to detect antibodies against dsDNA, **histones**, SS-A (Ro), SS-B (La), Smith, **Smith/RNP**, Scl-70, Jo-1, **centromeric proteins**, and **other antigens extracted from the HEp-2 cell nucleus**. ANA ELISA assays have been reported to have lower sensitivities than ANA IFA for systemic autoimmune rheumatic diseases (SARD).

Negative results do not necessarily rule out SARD.

**3000601**

**Antinuclear Antibody (ANA) with HEp-2 Substrate, IgG by IFA with Reflex by Pattern**

**ANA AB PAT**

**Reference Interval:**

Test Number	Components	Reference Interval			
3000082	Antinuclear Antibody (ANA) with HEp-2 Substrate, IgG by IFA	Less than 1:80			
2003040	PM/Scl-100 Antibody, IgG by Immunoblot	Negative			
2012173	Fibrillarin (U3 RNP) Antibody, IgG (Temporary Delay as of 3/18/2019 - no referral available)	Negative			
0050215	Double-Stranded DNA (dsDNA) Antibody, IgG by ELISA with Reflex to dsDNA Antibody, IgG by IFA	Effective August 20, 2012			
		Test Number	Components	Reference Interval	
			Double-Stranded DNA (dsDNA) Antibody, IgG by ELISA	None Detected.	
		2002693	Double-Stranded DNA (dsDNA) Antibody, IgG by IFA (using Crithidia luciliae)	Less than 1:10	
2002693	Double-Stranded DNA (dsDNA) Antibody, IgG by IFA (using Crithidia luciliae)	Less than 1:10			
2005287	Chromatin Antibody, IgG	19 Units or less	Negative		
		20-60 Units	Moderate Positive		
		61 Units or greater	Strong Positive		
2001601	RNA Polymerase III Antibody, IgG	19 Units or less	Negative		
		20-39 Units	Weak Positive		
		40-80 Units	Moderate Positive		
		81 Units or greater	Strong Positive		
0050599	Scleroderma (Scl-70) (ENA) Antibody, IgG	29 AU/mL or less	Negative		
		30-40 AU/mL	Equivocal		
		41 AU/mL or greater	Positive		
0050470	Smith/RNP (ENA) Antibody, IgG	29 AU/mL or less	Negative		
		30-40 AU/mL	Equivocal		
		41 AU/mL or greater	Positive		
0050085	Smith (ENA) Antibody, IgG	29 AU/mL or less	Negative		
		30-40 AU/mL	Equivocal		
		41 AU/mL or greater	Positive		
2012074	SSA 52 and 60 (Ro) (ENA) Antibodies, IgG	Test Number	Components	Reference Interval	
			SSA-52 (Ro52) (ENA) Antibody, IgG	29 AU/mL or Less	Negative
				30-40 AU/mL	Equivocal
				41 AU/mL or	Positive
			SSA-60 (Ro60) (ENA) Antibody, IgG	29 AU/mL or Less	Negative
				30-40 AU/mL	Equivocal
41 AU/mL or	Positive				
0050692	SSB (La) (ENA) Antibody, IgG	29 AU/mL or less	Negative		
		30-40 AU/mL	Equivocal		
		41 AU/mL or greater	Positive		

**Note:** The Antinuclear Antibody (ANA), IgG by IFA with Reflex by IFA Pattern begins with Nuclear Antibody (ANA) by IFA, IgG. Depending on findings, one or more reflexive tests may be required. Tests added may include Double-Stranded DNA (dsDNA) Antibody, IgG by ELISA; Double-Stranded DNA (dsDNA) Antibody, IgG by IFA (using *Crithidia luciliae*); Chromatin Antibody, IgG; **Smith/RNP (ENA)** Antibody, IgG; Fibrillarin (U3 RNP) Antibody, IgG; Smith (ENA) Antibody, IgG; SSA 52 (Ro) (ENA) Antibody, IgG; SSA 60 (Ro) (ENA) Antibody, IgG; SSB (La) (ENA) Antibody, IgG; Scleroderma (Scl-70) (ENA) Antibody, IgG; PM/Scl-100 Antibody, IgG, by Immunoblot; and/or RNA Polymerase III Antibody, IgG. Additional charges apply.

ANA determined by indirect fluorescence assay (IFA) use HEp-2 substrate and IgG-specific conjugate at a screening dilution of 1:80. If positive, patterns reported include homogeneous, speckled, centromere, nucleolar, nuclear dots, or cytoplasmic. All positive results are reported with endpoint titers.

HOTLINE: Effective August 19, 2019

**0051415**

**Ashkenazi Jewish Diseases, 16 Genes**

**AJP**

**HOTLINE NOTE:** There is a component change associated with this test.

Remove component 0051417, Ashkenazi Jewish Diseases, Allele 1  
 Removed component 0051418, Ashkenazi Jewish Diseases, Allele 2  
 Add component 3001464, Ashkenazi Jewish Diseases, Gene 1  
 Add component 3001465, Ashkenazi Jewish Diseases, Gene 2  
 Add component 3001466, AJP Gene 1, Allele 1  
 Add component 3001467, AJP Gene 1, Allele 2  
 Add component 3001468, AJP Gene 2, Allele 1  
 Add component 3001469, AJP Gene 2, Allele 2  
 Add component 3001470, Ashkenazi Jewish Diseases Carrier Status  
 Add component 3001754, Ashkenazi Jewish Diseases, Panel Results

**2014314**

**Autism and Intellectual Disability Comprehensive Panel**

**AID COMP**

**Note:** This panel includes Acylcarnitine Quantitative Profile, Plasma (ARUP test code 0040033); Mucopolysaccharides Screen - Electrophoresis and Quantitation, Urine (ARUP test code 0081352); Organic Acids, Urine (ARUP test code 0098389); Creatine Disorders Panel, Serum or Plasma (ARUP test code 2002328 ); Creatine Disorders Panel, Urine (ARUP test code 2002333); Amino Acids Quantitative by LC-MS/MS, Plasma (ARUP test code 2009389); Cytogenomic SNP Microarray (ARUP test code 2003414); Fragile X (FMR1) with Reflex to Methylation Analysis (ARUP test code 2009033). If Fragile X testing detects a CGG repeat of **100** or greater by PCR and capillary electrophoresis, methylation analysis will be added. Additional charges apply.

**New Test**

**3001410**

**Basement Membrane Zone Antibody Panel**

**BMZ AB PAN**

Available Now

[Click for Pricing](#)

**Methodology:** Indirect Fluorescent Antibody/Enzyme-Linked Immunosorbent Assay

**Performed:** Varies

**Reported:** 4-9 days

**Specimen Required:** Collect: Plain Red or Serum Separator Tube (SST).

Specimen Preparation: Transfer 2 mL serum to an ARUP Standard Transport Tube. (Min: 0.5 mL)

Storage/Transport Temperature: Refrigerated.

Unacceptable Conditions: Plasma. Hemolyzed or lipemic specimens.

Stability (collection to initiation of testing): Ambient: 1 week; Refrigerated: 2 weeks; Frozen: Indefinitely

**Reference Interval:** By report

**Interpretive Data:** Refer to report.

**Note:** For specimens less than 0.5 mL, call the Immunodermatology Laboratory at (866) 266-5699.

**CPT Code(s):** 88346, 88350 x3, 83516 x3

New York DOH approval pending. Call for status update.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

**2006193**

**B-Cell Clonality Screening (IgH and IgK) by PCR**

**BCELL SCRNL**

**Specimen Required:** Collect: Lavender (EDTA), bone marrow (EDTA), or tissue. **Also acceptable: DNA extracted by CLIA certified lab.**  
**Specimen Preparation: Whole Blood:** Do not freeze. Transport 5 mL whole blood. (Min: 1 mL)  
**Bone Marrow:** Do not freeze. Transport 3 mL bone marrow. (Min: 1 mL)  
**Fresh Tissue:** Freeze immediately. Transport 100 mg or 0.5-2.0 cm<sup>3</sup> tissue.  
**FFPE Tumor Tissue:** Formalin fix (10 percent neutral buffered formalin) and paraffin embed tissue. Protect from excessive heat.  
**Tissue block will be returned after testing.** Transport tissue block or four 10-micron shavings in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect<sup>™</sup> or contact ARUP Client Services at (800) 522-2787.  
**Extracted DNA:** Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect<sup>™</sup> or contact ARUP Client Services at (800) 522-2787.  
**Storage/Transport Temperature: Whole Blood, Bone Marrow or Extracted DNA:** Refrigerated.  
**Fresh Tissue:** Frozen on dry ice.  
**FFPE Tumor Tissue:** Room temperature. **Also acceptable:** Refrigerated. Ship in cooled container during summer months.  
**Unacceptable Conditions:** Plasma, serum, frozen tissue, DNA extracted by a non-CLIA lab. Specimens collected in anticoagulants other than EDTA or sodium heparin. Clotted or grossly hemolyzed specimens. Specimens fixed/processed in alternative fixatives or heavy metal fixatives (B-4 or B-5) or tissue sections on slides. Decalcified specimens.  
**Stability (collection to initiation of testing): Whole Blood or Bone Marrow:** Ambient: 24 hours; Refrigerated: 5 days; Frozen: Unacceptable  
**Fresh Tissue:** Ambient: Unacceptable; Refrigerated: 2 hours; Frozen: 1 year  
**FFPE Tumor Tissue:** Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable  
**Extracted DNA:** Ambient: 1 month; Refrigerate: Indefinitely; Frozen: Indefinitely

**2005017**

**BCR-ABLI, Major (p210), Quantitative**

**BCR MAJ**

**Specimen Required:** Collect: Lavender (EDTA) or bone marrow (EDTA). **Also acceptable: RNA extracted by CLIA certified lab.**  
**Specimen Preparation: Whole Blood:** Transport 5 mL whole blood. (Min: 1 mL)  
**Bone Marrow:** Transport 3 mL bone marrow. (Min: 1 mL)  
**Specimens must be received within 48 hours of collection due to lability of RNA.**  
**Extracted RNA:** Transport 40 uL RNA with at least 40 ng/uL concentration. (Min: 40 uL) Transport RNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect<sup>™</sup> or contact ARUP Client Services at (800) 522-2787.  
**Storage/Transport Temperature: Whole Blood or Bone Marrow: CRITICAL REFRIGERATED.** Separate specimens must be submitted when multiple tests are ordered.  
**Extracted RNA: CRITICAL FROZEN.** Separate specimens must be submitted when multiple tests are ordered.  
**Unacceptable Conditions:** Serum, plasma, CSF, extracted DNA, RNA extracted by a non-CLIA lab, bone core, or FFPE tissue. Specimens collected in anticoagulants other than EDTA. Severely hemolyzed or clotted specimens.  
**Stability (collection to initiation of testing):** Ambient: 1 hour; Refrigerated: 48 hours; Frozen: Unacceptable  
**Extracted RNA:** Ambient: Unacceptable; Refrigerate: Unacceptable; Frozen: Indefinitely

**2005016**

**BCR-ABLI, Minor (p190), Quantitative**

**BCR MIN**

**Specimen Required:** Collect: Lavender (EDTA) or bone marrow (EDTA). **Also acceptable: RNA extracted by CLIA certified lab.**  
**Specimen Preparation: Whole Blood:** Transport 5 mL whole blood. (Min: 1 mL)  
**Bone Marrow:** Transport 3 mL bone marrow. (Min: 1 mL)  
**Specimens must be received within 48 hours of collection due to lability of RNA.**  
**Extracted RNA:** Transport 40 uL RNA with at least 40 ng/uL concentration. (Min: 40 uL) Transport RNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect<sup>™</sup> or contact ARUP Client Services at (800) 522-2787.  
**Storage/Transport Temperature: Whole Blood or Bone Marrow: CRITICAL REFRIGERATED.** Separate specimens must be submitted when multiple tests are ordered.  
**Extracted RNA: CRITICAL FROZEN.** Separate specimens must be submitted when multiple tests are ordered.  
**Unacceptable Conditions:** Serum, plasma, CSF, extracted DNA, RNA extracted by a non-CLIA lab, bone core, or FFPE tissue. Specimens collected in anticoagulants other than EDTA. Severely hemolyzed or clotted specimens.  
**Stability (collection to initiation of testing):** Ambient: 1 hour; Refrigerated: 48 hours; Frozen: Unacceptable  
**Extracted RNA:** Ambient: Unacceptable; Refrigerate: Unacceptable; Frozen: Indefinitely

HOTLINE: Effective August 19, 2019

**2005010**

**BCR-ABL1, Qualitative with Reflex to BCR-ABL1 Quantitative**

**BCR RFLX**

**Specimen Required:** Collect: Lavender (EDTA) or bone marrow (EDTA). **Also acceptable: RNA extracted by CLIA certified lab.**  
Specimen Preparation: **Whole Blood:** Transport 5 mL whole blood. (Min: 1 mL)  
**Bone Marrow:** Transport 3 mL bone marrow. (Min: 1 mL)  
**Specimens must be received within 48 hours of collection due to lability of RNA.**  
**Extracted RNA:** Transport 40 uL RNA with at least 40 ng/uL concentration. (Min: 40 uL) Transport RNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  
Storage/Transport Temperature: **Whole Blood or Bone Marrow:** CRITICAL REFRIGERATED. Separate specimens must be submitted when multiple tests are ordered.  
**Extracted RNA:** CRITICAL FROZEN. Separate specimens must be submitted when multiple tests are ordered.  
Unacceptable Conditions: Serum, plasma, CSF, extracted DNA, RNA extracted by a non-CLIA lab, bone core, or FFPE tissue.  
Specimens collected in anticoagulants other than EDTA. Severely hemolyzed or clotted specimens.  
Stability (collection to initiation of testing): Ambient: 1 hour; Refrigerated: 48 hours; Frozen: Unacceptable  
**Extracted RNA:** Ambient: Unacceptable; Refrigerate: Unacceptable; Frozen: Indefinitely

**New Test**

**3001798**

**Blastomyces Antigen Quantitative by EIA, Urine**

**BLASTOAG U**

[Click for Pricing](#)

**Methodology:** Quantitative Enzyme Immunoassay  
**Performed:** Tue, Thu, Sat  
**Reported:** 1-4 days

**Specimen Required:** Collect: Urine.  
Specimen Preparation: Transfer 1 mL urine to an ARUP Standard Transport Tube. (Min: 0.5 mL)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Specimens preserved in boric acid.  
Stability (collection to initiation of testing): Ambient: 2 weeks; Refrigerated: 2 weeks; Frozen: Indefinitely

**Reference Interval:**

Less than 1.25 ng/mL	Not Detected
1.25-200 ng/mL	Detected
Greater than 200 ng/mL	Detected (above the limit of quantification)

**Interpretive Data:** The quantitative range of this assay is 1.25-200 ng/mL. Antigen concentrations between 200 ng/mL fall outside the linear range of the assay and cannot be accurately quantified.

This EIA test should be used in conjunction with other diagnostic procedures, including microbiological culture, histological examination of biopsy samples, and/or radiographic evidence, to aid in the diagnosis of blastomycosis.

Cross-reactivity with other endemic mycoses (*Histoplasma* and *Coccidioides*) may occur. Positive test results should be correlated with other clinical findings and relevant exposure history.

**CPT Code(s):** 87449

New York DOH approval pending. Call for status update.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

**2002926**

**Blastomyces dermatitidis Antigen Quantitative by EIA**

**BLAST DERM**

**Specimen Required:** Collect: **Plain** Red, Serum Separator Tube (SST), Lavender (**K<sub>2</sub>** or **K<sub>3</sub>** EDTA), Green (Sodium or Lithium Heparin), Light Blue (Sodium Citrate), CSF, or BAL.  
Specimen Preparation: **Transfer** 2 mL serum or plasma to an ARUP Standard Transport Tube. (Min: 1.2 mL)  
**Transfer** 1 mL CSF to an ARUP Standard Transport Tube. (Min: 0.8 mL)  
**Transfer** 1 mL BAL to an ARUP Standard Transport Tube. (Min: 0.5 mL)  
Storage/Transport Temperature: Refrigerated. Also acceptable: Room temperature or frozen.  
Unacceptable Conditions: **Urine.**  
Stability (collection to initiation of testing): Ambient: 2 weeks; Refrigerated: 2 weeks; Frozen: Indefinitely

**Note:** For urine specimens refer to *Blastomyces* Antigen Quantitative by EIA, Urine (ARUP test code 3001798).



HOTLINE: Effective August 19, 2019

<b><u>0062224</u></b>	<b><i>Blastomyces dermatitidis</i> Identification</b>	<b>MC BP</b>
<b>Methodology:</b>	Nucleic Acid Probe/MALDI (Matrix-Assisted Laser Desorption/Ionization)	
<b>Performed:</b>	Sun-Sat	
<b>Reported:</b>	2-7 days	
<b>CPT Code(s):</b>	Varies based on identification methods.	
<b><u>0060108</u></b>	<b>Body Fluid Culture and Gram Stain</b>	<b>MC BF</b>
<b>Specimen Required:</b> <u>Collect:</u> Aspirate of amniotic, culdocentesis, pericardial, peritoneal, synovial, thoracentesis or other normally sterile body fluid. <u>Specimen Preparation:</u> Transfer 5 mL aspirate to a <b>Sterile Container</b> . (Min: 2 mL) Place each specimen in an individually sealed bag. <u>Storage/Transport Temperature:</u> Room temperature. <u>Remarks:</u> Specimen source required. <u>Unacceptable Conditions:</u> <b>CSF or blood. Syringes</b> with needle attached. <u>Stability (collection to initiation of testing):</u> Ambient: 24 hours; Refrigerated: Unacceptable; Frozen: Unacceptable		
<b>Note:</b> Gram stain, identification, and susceptibility tests are billed separately from culture. Anaerobe culture is NOT included with this order. Anaerobe culture is recommended for body fluids, tissue, and deep wound/surgical cultures. If anaerobe culture is needed, please order Anaerobe Culture (ARUP test code 0060143) and use anaerobic collection device for transportation.		
For CSF, order Cerebrospinal Fluid (CSF) Culture and Gram Stain (ARUP test code 0060106). For blood, order Blood Culture (ARUP test code 0060102) or Blood Culture, AFB and Fungal (ARUP test code 0060024).		
<b><u>0070053</u></b>	<b>Bone Specific Alkaline Phosphatase</b>	<b>BSAP</b>
<b>Specimen Required:</b> <u>Collect:</u> Serum separator tube. Also acceptable: Green (sodium or lithium heparin). <u>Specimen Preparation:</u> Transfer 0.5 mL serum or plasma to an ARUP Standard Transport Tube. (Min: 0.3 mL) <u>Storage/Transport Temperature:</u> <b>CRITICAL FROZEN. Separate specimens must be submitted when multiple tests are ordered.</b> <u>Unacceptable Conditions:</u> Urine. Grossly hemolyzed specimens. <u>Stability (collection to initiation of testing):</u> After separation from cells: Ambient: 2 hours; Refrigerated: 48 hours; Frozen: 2 months		
<b><u>2002498</u></b>	<b>BRAF Codon 600 Mutation Detection by Pyrosequencing</b>	<b>BRAF PCR</b>
<b>Specimen Required:</b> <u>Collect:</u> Tumor tissue. Also acceptable: DNA extracted by CLIA certified lab with corresponding client-circled H&E slide. <u>Specimen Preparation:</u> <b>Tumor Tissue:</b> Formalin fix (10 percent neutral buffered formalin) and paraffin embed tissue. Protect from excessive heat. <b>Tissue block will be returned after testing.</b> Transport tissue block or 5 unstained 5 micron slides. (Min: 3 slides). <b>Transport</b> block and/or slide(s) in a tissue transport kit (ARUP Supply # 47808) available online through eSupply using ARUP Connect <sup>™</sup> or contact ARUP Client Services at (800) 522-2787. <b>Extracted DNA:</b> Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect <sup>™</sup> or contact ARUP Client Services at (800) 522-2787. <u>Storage/Transport Temperature:</u> Room temperature. Also acceptable: Refrigerated. Ship in cooled container during summer months. <b>Extracted DNA: Refrigerated.</b> <u>Remarks:</u> Include surgical pathology report. <u>Unacceptable Conditions:</u> Less than 25 percent tumor. <b>DNA extracted by a non-CLIA lab. DNA extracted without a corresponding circled H&amp;E slide.</b> Specimens fixed/processed in alternative fixatives (alcohol, Prefer) or heavy metal fixatives ( <b>B-4 or B-5</b> ). Decalcified specimens. <u>Stability (collection to initiation of testing):</u> Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable <b>Extracted DNA: Ambient: 1 month; Refrigerated: Indefinitely; Frozen: Indefinitely</b>		

HOTLINE: Effective August 19, 2019

**0051750**

**BRAF Codon 600 Mutation Detection with Reflex to *MLH1* Promoter Methylation**

**BRAF RFLX**

**Specimen Required:** Collect: Tumor tissue. **Also acceptable:** DNA extracted by CLIA certified lab with corresponding client-circled H&E slide.  
Specimen Preparation: **Tumor Tissue:** Formalin fix (10 percent neutral buffered formalin) and paraffin embed tissue. Protect from excessive heat. **Tissue block will be returned after testing.** Transport tissue block or 5 unstained 5 micron slides. (Min: 3 slides). Transport block and/or slide(s) in a tissue transport kit (ARUP Supply # 47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  
**Extracted DNA:** Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  
Storage/Transport Temperature: Room temperature. **Also acceptable:** Refrigerated. Ship in cooled container during summer months.  
**Extracted DNA:** Refrigerated.  
Remarks: Include surgical pathology report.  
Unacceptable Conditions: Less than 25 percent tumor. **DNA extracted by a non-CLIA lab, DNA extracted without a corresponding circled H&E slide.** Specimens fixed/processed in alternative fixatives (alcohol, Prefer) or heavy metal fixatives (B-4 or B-5). Decalcified specimens.  
Stability (collection to initiation of testing): Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable  
**Extracted DNA:** Ambient: 1 month; Refrigerated: Indefinitely; Frozen: Indefinitely

**Interpretive Data:** Refer to report.  
 See Compliance Statement B: [www.aruplab.com/CS](http://www.aruplab.com/CS)

**2007132**

**BRAF V600E Mutation Detection in Hairy Cell Leukemia by Real-Time PCR, Quantitative**

**BRAF HCL**

**Specimen Required:** Collect: Lavender (EDTA) or bone marrow (EDTA). **Also acceptable:** DNA extracted by CLIA certified lab.  
Specimen Preparation: **Whole Blood:** Do not freeze. Transport 5 mL whole blood. (Min: 1 mL)  
**Bone Marrow:** Transport 3 mL bone marrow. (Min: 1 mL)  
**Extracted DNA:** Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Plasma, serum, FFPE tissue blocks/slides, or frozen tissue, **DNA extracted by a non-CLIA lab.** Specimens collected in anticoagulants other than EDTA or sodium heparin. Clotted or grossly hemolyzed specimens.  
Stability (collection to initiation of testing): Ambient: 24 hours; Refrigerated: 5 days; Frozen: Unacceptable  
**Extracted DNA:** Ambient: 1 month; Refrigerate: Indefinitely; Frozen: Indefinitely

**2008708**

**Calculi Risk Assessment, Urine**

**CRA**

**Specimen Required:** Collect: 24-hour urine. Refrigerate during collection.  
Specimen Preparation: Thoroughly mix entire collection (24-hour) in one container. Transport four separate 4 mL aliquots of urine using Calculi Risk/Supersaturation Urine Collection Kit (ARUP supply# 46007). Available online through eSupply using ARUP Connect™ or contact Client Services at (800) 522-2787. **Do not exceed 4 mL in tubes.**  
 Aliquot according to the following specifications:  
 1st aliquot (pH 2): Transfer 4 mL urine into a Sulfamic Acid Tube. (Min: 4 mL) Mix well. Freeze immediately.  
 2nd aliquot (pH 2): Transfer 4 mL urine into a Sulfamic Acid Tube. (Min: 4 mL) Mix well. Freeze immediately.  
 3rd aliquot (pH 9): Transfer 4 mL urine into a Sodium Carbonate Tube. (Min: 4mL) Mix well. Freeze immediately.  
 4th aliquot: Transfer 4 mL urine into an Unpreserved Tube. (Min: 4 mL) Freeze immediately.  
 If collection kit is unavailable, transport four 4 mL unadjusted aliquots of urine.  
Storage/Transport Temperature: **CRITICAL FROZEN.** Separate specimens must be submitted when multiple tests are ordered. Ship patient and control specimens together.  
Remarks: Record total volume and collection time interval on transport tube and test request form.  
Stability (collection to initiation of testing): **Ambient:** Unacceptable; Refrigerated: Unacceptable; Frozen: 2 weeks

HOTLINE: Effective August 19, 2019

**2010673**

**CALR (Calreticulin) Exon 9 Mutation Analysis by PCR**

**CALR**

**Specimen Required:** Collect: Lavender (EDTA) or bone marrow (EDTA). **Also acceptable:** DNA extracted by CLIA certified lab.

Specimen Preparation: **Whole Blood:** Do not freeze. Transport 5 mL whole blood. (Min: 1 mL)

**Bone Marrow:** Do not freeze. Transport 3 mL bone marrow. (Min: 1 mL)

**Extracted DNA:** Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.

Storage/Transport Temperature: Refrigerated.

Unacceptable Conditions: Plasma, serum, FFPE tissue blocks/slides, or frozen tissue, DNA extracted by a non-CLIA lab. Specimens collected in anticoagulants other than EDTA or sodium heparin. Clotted or grossly hemolyzed specimens.

Stability (collection to initiation of testing): Ambient: 24 hours; Refrigerated: 5 days; Frozen: Unacceptable

**Extracted DNA:** Ambient: 1 month; Refrigerate: Indefinitely; Frozen: Indefinitely

**2002918**

**Carbohydrate Deficient Transferrin for Congenital Disorders of Glycosylation (CDG)**

**CARBOH-CDG**

**Performed:** Varies

**Reported:** 5-13 days

**Specimen Required:** Collect: Plain red or serum separator tube (SST).

Specimen Preparation: Transfer 0.1 mL serum to an ARUP Standard Transport Tube. (Min: 0.05 mL)

Storage/Transport Temperature: Frozen. Also acceptable: Room temperature or refrigerated.

Remarks: Patient age is required on the test request form. Provide reason (eg, diagnosis) for referral with each specimen.

Stability (collection to initiation of testing): Ambient: 1 week; Refrigerated: 28 days; Frozen: 45 days

**New Test**

**3001697**

**Carbonic Anhydrase IX by IHC**

**CAIX IHC**

Available Now

[Click for Pricing](#)

**Methodology:** Immunohistochemistry

**Performed:** Mon-Fri

**Reported:** 1-3 days

**Specimen Required:** Collect: Tissue.

Specimen Preparation: Formalin fix (10 percent neutral buffered formalin) and paraffin embed specimen (cells must be prepared into a cellblock). Protect paraffin block and/or slides from excessive heat. Transport tissue block or 5 unstained (3- to 5-micron thick sections), positively charged slides in a tissue transport kit (recommended but not required), (ARUP supply #47808) available online through eSupply using ARUP Connect or contact ARUP Client Services at (800) 522-2787. (Min: 2 slides) If sending precut slides, do not oven bake.

Storage/Transport Temperature: Room temperature. Also acceptable: Refrigerated. Ship in cooled container during summer months.

Unacceptable Conditions: Specimens submitted with non-representative tissue type. Depleted specimens.

Stability (collection to initiation of testing): Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable

**Note:** All stains will be handled as "Stain and Return" unless a consultation is requested. To request a consultation, submit the pathology report, all associated case materials (clinical history, blocks, slides, etc.), and the Anatomic Pathology requisition form (#32960) in place of the Immunohistochemistry Stain Form.

**CPT Code(s):** 88342

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

0080407

Catecholamines Fractionated by LC-MS/MS, Urine Free

CATE UF

Reference Interval:

Reference Intervals for 24-Hour Calculations (24-Hour Urine)

Test Number	Components	Reference Interval		
	Dopamine	Effective August 17, 2015		
		Age	Dopamine	
		0-3 years	Not Established	
		4-6 years	95-221 µg/d	
		7-12 years	76-371 µg/d	
		13-17 years	137-393 µg/d	
		18-69 years	77-324 µg/d	
		70 years and older	56-272 µg/d	
	Epinephrine	Effective August 19, 2019		
		Age	Epinephrine	
		0-3 years	Not Established	
		4-10 years	1-14 µg/d	
		11-17 years	1-18 µg/d	
		18 years and older	1-14 µg/d	
	Norepinephrine	Effective August 17, 2015		
		0-3 years	Not Established	
		4-12 years	6-45 µg/d	
		13-17 years	15-57 µg/d	
		18-69 years	16-71 µg/d	
		70 years and older	11-60 µg/d	
0020473	Creatinine, Urine - per 24h	Age	Male	Female
		3-8 years	140-700 mg/d	140-700 mg/d
		9-12 years	300-1300 mg/d	300-1300 mg/d
		13-17 years	500-2300 mg/d	400-1600 mg/d
		18-50 years	1000-2500 mg/d	700-1600 mg/d
		51-80 years	800-2100 mg/d	500-1400 mg/d
		81 years and older	600-2000 mg/d	400-1300 mg/d

Reference Intervals for Ratio-to-Creatinine (CRT) Calculations (Random Urine)

Test Number	Components	Reference Interval	
	Dopamine	Age	Dopamine
		0-11 months	240-1290 µg/g crt
		1-3 years	80-1220 µg/g crt
		4-10 years	220-720 µg/g crt
		11-17 years	120-450 µg/g crt
		18 years and older	0-250 µg/g crt
	Epinephrine	Age	Epinephrine
		0-11 months	0-380 µg/g crt
		1-3 years	0-82 µg/g crt
		4-10 years	5-93 µg/g crt
		11-17 years	3-58 µg/g crt
		18 years and older	0-20 µg/g crt
	Norepinephrine	Age	Norepinephrine
		0-11 months	25-310 µg/g crt
		1-3 years	25-290 µg/g crt
		4-10 years	27-110 µg/g crt
		11-17 years	4-105 µg/g crt
		18 years and older	0-45 µg/g crt

HOTLINE: Effective August 19, 2019

<a href="#">2011114</a>	CBFB-MYH11 inv(16) Detection, Quantitative	INV 16 QNT
<b>Specimen Required:</b> <u>Collect:</u> Lavender (EDTA) or bone marrow (EDTA). Also acceptable: RNA extracted by CLIA certified lab. <u>Specimen Preparation:</u> <b>Whole Blood:</b> Transport 5 mL whole blood. (Min: 1 mL) <b>Bone Marrow:</b> Transport 3 mL bone marrow. (Min: 1 mL) <b>Specimens must be received within 48 hours of collection due to lability of RNA.</b> <b>Extracted RNA:</b> Transport 40 uL RNA with at least 40 ng/uL concentration. (Min: 40 uL) Transport RNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787. <u>Storage/Transport Temperature:</u> <b>Whole Blood or Bone Marrow:</b> CRITICAL REFRIGERATED. Separate specimens must be submitted when multiple tests are ordered. <b>Extracted RNA:</b> CRITICAL FROZEN. Separate specimens must be submitted when multiple tests are ordered. <u>Unacceptable Conditions:</u> Serum, plasma, CSF, extracted DNA, RNA extracted by a non-CLIA lab, bone core, or FFPE tissues. Specimens collected in anticoagulants other than EDTA. Severely hemolyzed or clotted specimens. <u>Stability (collection to initiation of testing):</u> Ambient: 1 hour; Refrigerated: 48 hours; Frozen: Unacceptable <b>Extracted RNA:</b> Ambient: Unacceptable; Refrigerate: Unacceptable; Frozen: Indefinitely		
<a href="#">2004247</a>	CEBPA Mutation Detection	CEBPA MUT
<b>Specimen Required:</b> <u>Collect:</u> Lavender (EDTA) or bone marrow (EDTA). Also acceptable: DNA extracted by CLIA certified lab. <u>Specimen Preparation:</u> <b>Whole Blood:</b> Do not freeze. Transport 5 mL whole blood (Min: 1 mL) <b>Bone Marrow:</b> Transport 3 mL bone marrow. (Min: 1 mL) <b>Extracted DNA:</b> Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787. <u>Storage/Transport Temperature:</u> Refrigerated. <u>Unacceptable Conditions:</u> Plasma, serum, FFPE tissue blocks/slides, or frozen tissue, DNA extracted by a non-CLIA lab. Specimens collected in anticoagulants other than EDTA or sodium heparin. Clotted or grossly hemolyzed specimens. <u>Stability (collection to initiation of testing):</u> Ambient: 24 hours; Refrigerated: 5 days; Frozen: Unacceptable <b>Extracted DNA:</b> Ambient: 1 month; Refrigerate: Indefinitely; Frozen: Indefinitely		
<a href="#">0080469</a>	Chromogranin A	CGA SERUM
<b>Reference Interval:</b> Effective August 19, 2019 0-160 ng/mL		
<a href="#">2011075</a>	Coccidioides Antigen Quantitative by EIA	COCCI AG
<b>Performed:</b>	Varies	
<b>Reported:</b>	3-4 days	
<b>Specimen Required:</b> <u>Collect:</u> Plain Red, Serum Separator Tube (SST), Lavender (K <sub>2</sub> or K <sub>3</sub> EDTA), Pink (K <sub>2</sub> EDTA), Green (Sodium or Lithium Heparin), or Light Blue (CTAD). Also acceptable: Urine, CSF, or BAL. <u>Specimen Preparation:</u> Transfer 2 mL serum or plasma to an ARUP Standard Transport Tube. (Min: 1.2 mL) Transfer 1 mL urine or BAL to an ARUP Standard Transport Tube. (Min: 0.5 mL) Transfer 1 mL CSF to an ARUP Standard Transport Tube. (Min: 0.8 mL) <u>Storage/Transport Temperature:</u> Refrigerated. Also acceptable: Room temperature or frozen. <u>Remarks:</u> Specimen source required. <u>Stability (collection to initiation of testing):</u> Ambient: 28 days; Refrigerated: 28 days; Frozen: Indefinitely		
<a href="#">0062225</a>	Coccidioides immitis Identification	MC CP
<b>Methodology:</b>	Nucleic Acid Probe/MALDI (Matrix-Assisted Laser Desorption/Ionization)	
<b>Performed:</b>	Sun-Sat	
<b>Reported:</b>	2-7 days	
<b>CPT Code(s):</b>	Varies based on identification methods.	

HOTLINE: Effective August 19, 2019

**3000480**

**Comprehensive Systemic Sclerosis Panel**

**SCL COMPRE**

**Reference Interval:**

Test Number	Components	Reference Interval	
0050599	Scleroderma (Scl-70) (ENA) Antibody, IgG	29 AU/mL or less	Negative
		30-40 AU/mL	Equivocal
		41 AU/mL or greater	Positive
0050470	Smith/RNP (ENA) Antibody, IgG	29 AU/mL or less	Negative
		30-40 AU/mL	Equivocal
		41 AU/mL or greater	Positive
3000082	Antinuclear Antibody (ANA) with HEp-2 Substrate, IgG by IFA	Less than 1:80	
2012173	Fibrillarin (U3 RNP) Antibody, IgG	Negative	
2003040	PM/Scl-100 Antibody, IgG by Immunoblot	Negative	
2001601	RNA Polymerase III Antibody, IgG	19 Units or less	Negative
		20-39 Units	Weak Positive
		40-80 Units	Moderate Positive
		81 Units or greater	Strong Positive

**Note:** Panel includes: Antinuclear Antibody (ANA) with HEp-2 Substrate, IgG by IFA, Scleroderma (Scl-70) (ENA) Antibody, IgG, RNA Polymerase III Antibody, IgG, Smith/RNP (ENA) Antibody, IgG, Fibrillarin (U3 RNP) Antibody, IgG, PM/Scl-100 Antibody, IgG by Immunoblot.

**HOTLINE NOTE:** There is a clinically significant charting name change associated with this test.

Change the charting name for component 0050470 from Ribonucleic Protein (U1) (ENA) Ab, IgG to Smith/RNP (ENA) Ab, IgG.

**0051668**

**Connective Tissue Diseases Profile**

**CONN**

**Reference Interval:**

Effective May 18, 2015

Test Number	Components	Reference Interval				
0050085	Smith (ENA) Antibody, IgG	29 AU/mL or less		Negative		
		30-40 AU/mL		Equivocal		
		41 AU/mL or greater		Positive		
0050470	Smith/RNP (ENA) Antibody, IgG	29 AU/mL or less		Negative		
		30-40 AU/mL		Equivocal		
		41 AU/mL or greater		Positive		
2012074	SSA 52 and 60 (Ro) (ENA) Antibodies, IgG	Test Number	Components		Reference Interval	
			SSA-52 (Ro52) (ENA) Antibody, IgG	29 AU/mL or Less	Negative	
				30-40 AU/mL	Equivocal	
				41 AU/mL or	Positive	
			SSA-60 (Ro60) (ENA) Antibody, IgG	29 AU/mL or Less	Negative	
				30-40 AU/mL	Equivocal	
				41 AU/mL or	Positive	
0050692	SSB (La) (ENA) Antibody, IgG	29 AU/mL or less		Negative		
		30-40 AU/mL		Equivocal		
		41 AU/mL or greater		Positive		
0099592	Jo-1 Antibody, IgG	29 AU/mL or less		Negative		
		30-40 AU/mL		Equivocal		
		41 AU/mL or greater		Positive		
0099249	Ribosomal P Protein Antibody	29 AU/mL or less		Negative		
		30-40 AU/mL		Equivocal		
		41 AU/mL or greater		Positive		
0050714	Centromere Antibody, IgG	29 AU/mL or less		Negative		
		30-40 AU/mL		Equivocal		
		41 AU/mL or greater		Positive		
0050599	Scleroderma (Scl-70) (ENA) Antibody, IgG	29 AU/mL or less		Negative		
		30-40 AU/mL		Equivocal		
		41 AU/mL or greater		Positive		

**HOTLINE NOTE:** There is a clinically significant charting name change associated with this test.

Change the charting name for component 0050470 from Ribonucleic Protein (U1) (ENA) Ab, IgG to Smith/RNP (ENA) Ab, IgG.

HOTLINE: Effective August 19, 2019

**2000133**

**Cytology, SurePath Liquid-Based Pap Test and Human Papillomavirus (HPV), High Risk by PCR, SurePath (for routine co-testing in women over 30)**

**GH REQUEST**

**Note:** In addition to the SurePath Pap Test, Human Papillomavirus (HPV) High Risk by PCR will be performed and reported under a separate accession. The Pap test is a screening test for cervical cancer and its precursors with an inherent false-negative rate.

**CPT Code(s):** 87624; 88142, if reviewed by pathologist add 88141

**New Test** **3001783**  
[Click for Pricing](#)

**Dermatomyositis and Polymyositis Panel**

**COMBI PAN**



**Additional Technical Information**

**Methodology:** Qualitative Immunoprecipitation/Semi-Quantitative Multiplex Bead Assay/Qualitative Immunoblot  
**Performed:** Sun-Sat  
**Reported:** 7-18 days

**Specimen Required:** Collect: Serum Separator Tube (SST).

Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer two 1 mL serum aliquots to ARUP

Standard Transport Tubes: (Min: 0.5 mL/aliquot)

Storage/Transport Temperature: Refrigerated.

Unacceptable Conditions: Hemolyzed, hyperlipemic, icteric, heat-treated or contaminated specimens.

Stability (collection to initiation of testing): Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

**Reference Interval:**

Test Number	Components	Reference Interval	
0099592	Jo-1 Antibody, IgG	29 AU/mL or less	Negative
		30-40 AU/mL	Equivocal
		41 AU/mL or greater	Positive
	PL-7 (threonyl-tRNA synthetase) Antibody	Negative	
	PL-12 (alanyl-tRNA synthetase) Antibody	Negative	
	EJ (glycyl-tRNA synthetase) Antibody	Negative	
	SRP (Signal Recognition Particle) Ab	Negative	
	OJ (isoleucyl-tRNA synthetase) Antibody	Negative	
	Mi-2 (nuclear helicase protein) Antibody	Negative	
	P155/140 Antibody	Negative	
	SAE1 (SUMO activating enzyme) Ab	Negative	
	MDA5 (CADM-140) Ab	Negative	
	NXP2 (Nuclear matrix protein-2) Ab	Negative	
	TIF1-gamma Ab	Negative	

**Interpretive Data:** Refer to report.

See Compliance Statement D: [www.aruplab.com/CS](http://www.aruplab.com/CS)

**Note:** Antibodies: PL-7, PL12, EJ, OJ, SRP, Jo-1, Mi-2, P155/140, SAE1, MDA5, NXP2, TIF1-gamma

**CPT Code(s):** 83516 x11; 86235

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

**New Test**     [3001782](#)  
[Click for Pricing](#)

**Dermatomyositis Autoantibody Panel**

**DERM PAN**



**Additional Technical Information**

**Methodology:** Qualitative Immunoprecipitation/Qualitative Immunoblot  
**Performed:** Sun-Sat  
**Reported:** 7-18 days

**Specimen Required:** Collect: Serum Separator Tube (SST).

Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer one 1 mL serum aliquots to ARUP Standard Transport Tubes. (Min: 0.5 mL/aliquot)

Storage/Transport Temperature: Refrigerated.

Unacceptable Conditions: Hemolyzed, hyperlipemic, icteric, heat-treated or contaminated specimens.

Stability (collection to initiation of testing): Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

**Reference Interval:**

Test Number	Components	Reference Interval
	Mi-2 (nuclear helicase protein) Antibody	Negative
	P155/140 Antibody	Negative
	SAE1 (SUMO activating enzyme) Ab	Negative
	MDA5 (CADM-140) Ab	Negative
	NXP2 (Nuclear matrix protein-2) Ab	Negative
	TIF1-gamma Ab	Negative

**Interpretive Data:** Refer to report.

See Compliance Statement D: [www.aruplab.com/CS](http://www.aruplab.com/CS)

**Note:** Antibodies: Mi-2, P155/140, SAE1, MDA5, NXP2, TIF1-gamma

**CPT Code(s):** 83516 x6

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

[2002440](#)

**EGFR Mutation Detection by Pyrosequencing**

**EGFR PCR**

**Specimen Required:** Patient Prep: For a general FNA collection and smear preparation refer to ARUP's Laboratory Test Directory: Cytology, Fine Needle Aspiration Collection at <http://ltd.aruplab.com/tests/pdf/366>

Collect: Tumor tissue. **Also acceptable: DNA extracted by CLIA certified lab with corresponding client-circled H&E slide.**

Specimen Preparation: **Tumor Tissue:** Formalin fix (10 percent neutral buffered formalin) and paraffin embed tissue. Protect from excessive heat. Tissue block will be returned after testing. Transport tissue block or 5 unstained 5-micron slides. (Min: 3 slides)

**Fine Needle Aspirate (FNA):** Prepare FNA smear with Diff-Quik or equivalent stain by standard methods (air-dried slides are preferred). Number of slides needed is dependent on the tumor cellularity of the smear. (Min: 1 slide). Slide(s) will be destroyed during testing process and will not be returned to client. Transport block and/or slide(s) in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.

**Extracted DNA:** Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.

Storage/Transport Temperature: Room temperature. Also acceptable: Refrigerated. Ship in cooled container during summer months.

**Extracted DNA:** Refrigerated.

Unacceptable Conditions: Less than 25 percent tumor. **DNA extracted by a non-CLIA lab. DNA extracted without a corresponding circled H&E slide.** Specimens fixed/processed in alternative fixatives (alcohol, Prefer) or heavy metal fixatives (B-4 or B-5). Decalcified specimens. FNA smears with less than 50 tumor cells.

Stability (collection to initiation of testing): Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable

**Extracted DNA:** Ambient: 1 month; Refrigerated: Indefinitely; Frozen: Indefinitely

**Note:** This test detects mutations in *EGFR* exons 18, 19, 20 and 21 (codons 719, 745-753, 768, 790, 858, and 861).



HOTLINE: Effective August 19, 2019

**2007914**

**EPOR Mutation Detection by Sequencing**

**EPOR**

**Specimen Required:** Collect: Lavender (EDTA). Also acceptable: DNA extracted by CLIA certified lab.  
**Specimen Preparation:** Whole Blood: Do not freeze. Transport 5 mL whole blood. (Min: 1 mL)  
**Extracted DNA:** Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  
**Storage/Transport Temperature:** Refrigerated.  
**Unacceptable Conditions:** Plasma, serum, FFPE tissue blocks/slides, or frozen tissue, DNA extracted by a non-CLIA lab, bone marrow. Specimens collected in anticoagulants other than EDTA or sodium heparin. Clotted or grossly hemolyzed specimens.  
**Stability (collection to initiation of testing):** Ambient: 24 hours; Refrigerated: 5 days; Frozen: Unacceptable  
**Extracted DNA:** Ambient: 1 month; Refrigerate: Indefinitely; Frozen: Indefinitely

**New Test**

**3001781**

**Extended Myositis Panel**

**MYOS EXT**

[Click for Pricing](#)



**Additional Technical Information**

**Methodology:** Qualitative Immunoprecipitation/Semi-Quantitative Multiplex Bead Assay/Qualitative Immunoblot  
**Performed:** Sun-Sat  
**Reported:** 7-18 days

**Specimen Required:** Collect: Serum Separator Tube (SST).  
**Specimen Preparation:** Separate from cells ASAP or within 2 hours of collection. Transfer three 1 mL serum aliquots to ARUP Standard Transport Tubes. (Min: 0.5 mL/aliquot)  
**Storage/Transport Temperature:** Refrigerated.  
**Unacceptable Conditions:** Hemolyzed, hyperlipemic, icteric, heat-treated or contaminated specimens.  
**Stability (collection to initiation of testing):** Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

**Reference Interval:**

Test Number	Components	Reference Interval		
2012074	SSA 52 and 60 (Ro) (ENA) Antibodies, IgG	Test Number	Components	Reference Interval
			SSA-52 (Ro52) (ENA) Antibody, IgG	29 AU/mL or Less: Negative 30-40 AU/mL: Equivocal 41 AU/mL or greater: Positive
			SSA-60 (Ro60) (ENA) Antibody, IgG	29 AU/mL or Less: Negative 30-40 AU/mL: Equivocal 41 AU/mL or greater: Positive
0050470	SM/RNP (ENA) Antibody, IgG	29 AU/mL or less	Negative	
		30-40 AU/mL	Equivocal	
		41 AU/mL or greater	Positive	
0099592	Jo-1 Antibody, IgG	29 AU/mL or less	Negative	
		30-40 AU/mL	Equivocal	
		41 AU/mL or greater	Positive	
	Mi-2 (nuclear helicase protein) Antibody	Negative		
	PL-7 (threonyl-tRNA synthetase) Antibody	Negative		
	PL-12 (alanyl-tRNA synthetase) Antibody	Negative		
	P155/140 Antibody	Negative		
	EJ (glycyl-tRNA synthetase) Antibody	Negative		
	Ku Antibody	Negative		
	SRP (Signal Recognition Particle) Ab	Negative		
	OJ (isoleucyl-tRNA synthetase) Antibody	Negative		
	SAE1 (SUMO activating enzyme) Ab	Negative		
	MDA5 (CADM-140) Ab	Negative		
	NXP2 (Nuclear matrix protein-2) Ab	Negative		
	TIF1-gamma Ab	Negative		
2012173	Fibrillarin (U3 RNP) Antibody, IgG	Negative		
2003040	PM/Sci-100 Antibody, IgG by Immunoblot	Negative		

**Interpretive Data:** Refer to report.  
 See Compliance Statement D: [www.aruplab.com/CS](http://www.aruplab.com/CS)

**Note:** Antibodies: Mi-2, PL-7, PL12, P155/140, EJ, Ku, OJ, PM/Sci, SRP, SM/RNP, Ro52, Ro60, Jo-1, U3 Fib, SAE1, NXP2, MDA5, TIF1-gamma

HOTLINE: Effective August 19, 2019

CPT Code(s): 83516 x12; 86235 x6

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

**0050652** Extractable Nuclear Antigen Antibodies (**Smith/RNP**, Smith, SSA 52, SSA 60, and SSB) ENA ABS4

**Reference Interval:**

Effective May 18, 2015

Test Number	Components	Reference Interval				
0050470	Smith/RNP (ENA) Antibody, IgG	29 AU/mL or less	Negative			
		30-40 AU/mL	Equivocal			
		41 AU/mL or greater	Positive			
0050085	Smith (ENA) Antibody, IgG	29 AU/mL or less	Negative			
		30-40 AU/mL	Equivocal			
		41 AU/mL or greater	Positive			
2012074	SSA 52 and 60 (Ro) (ENA) Antibodies, IgG	Test Number	Components		Reference Interval	
			SSA-52 (Ro52) (ENA) Antibody, IgG	29 AU/mL or Less	Negative	
				30-40 AU/mL	Equivocal	
				41 AU/mL or	Positive	
			SSA-60 (Ro60) (ENA) Antibody, IgG	29 AU/mL or Less	Negative	
				30-40 AU/mL	Equivocal	
				41 AU/mL or	Positive	
0050692	SSB (La) (ENA) Antibody, IgG	29 AU/mL or less	Negative			
		30-40 AU/mL	Equivocal			
		41 AU/mL or greater	Positive			

**HOTLINE NOTE:** There is a clinically significant charting name change associated with this test.

Change the charting name for component 0050470 from Ribonucleic Protein (U1) (ENA) Ab, IgG to **Smith/RNP (ENA) Ab, IgG**.

**0094030** Felbamate **FELBAMA**

**Reference Interval:**

Effective August 19, 2019

Therapeutic Range	30 – 60 µg/mL
Toxic Level	Greater than 120 µg/mL

**Interpretive Data:** Patient pharmacokinetics may be variable due to age, co-medications, and/or compromised renal function. Adverse effects may include nausea, vomiting, dizziness, blurred vision and ataxia. Felbamate use may increase the incidence of liver failure and aplastic anemia.

See Compliance Statement B: [www.aruplab.com/CS](http://www.aruplab.com/CS)

**3001161** FLT3 ITD and TKD Mutation Detection **FLT3-PCR**

**Specimen Required:** Collect: Lavender (EDTA) or bone marrow (EDTA). Also acceptable: DNA extracted by CLIA certified lab.

**Specimen Preparation:** Whole Blood: Do not freeze. Transport 5 mL whole blood. (Min: 1 mL)

**Bone Marrow:** Do not freeze. Transport 3 mL bone marrow. (Min: 1 mL)

**Extracted DNA:** Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.

**Storage/Transport Temperature:** Refrigerated.

**Unacceptable Conditions:** Plasma, serum, FFPE tissue blocks/slides, or frozen tissue, DNA extracted by a non-CLIA lab. Specimens collected in anticoagulants other than EDTA or sodium heparin. Clotted or grossly hemolyzed specimens.

**Stability (collection to initiation of testing):** Ambient: 24 hours; Refrigerated: 5 days; Frozen: Unacceptable

**Extracted DNA:** Ambient: 1 month; Refrigerate: Indefinitely; Frozen: Indefinitely

HOTLINE: Effective August 19, 2019

**2009033**

**Fragile X (*FMRI*) with Reflex to Methylation Analysis**

**FRAG X PCR**

**Specimen Required:** Collect: Lavender (**K<sub>2</sub>EDTA**), Pink (K<sub>2</sub>EDTA), or Yellow (ACD Solution A or B).

Specimen Preparation: Transport 5 mL whole blood. (Min: 1.5 mL)

Storage/Transport Temperature: **Refrigerated.**

Stability (collection to initiation of testing): Ambient: **1 week**; Refrigerated: **1 month**; Frozen: **6 months**

**Interpretive Data:**

**Background Information for Fragile X (*FMRI*)**

**Characteristics:** Fragile X syndrome, the most common heritable form of mental retardation, is characterized by moderate mental retardation in males and mild mental retardation in females, hyperactivity, perseverative speech, social anxiety, poor eye contact, hand flapping or biting, autism spectrum disorders behavioral phenotype, and connective tissue anomalies. Adult males may have physical findings including: macroorchidism, a long narrow face, prominent ears and jaw, and a single palmar crease.

**Incidence:** 1 in 4,000 Caucasian males and 1 in 8,000 Caucasian females; unknown in other ethnicities.

**Inheritance:** X-linked dominant.

**Penetrance:** Reduced in females.

**Cause:** Expansion of the *FMRI* gene CGG triplet repeat.

Full mutation: >200-230 CGG repeats (methylated)

Premutation: 55-200 CGG repeats (unmethylated)

Intermediate: 45-54 CGG repeats (unmethylated)

Normal: 5-44 CGG repeats (unmethylated)

**Clinical Sensitivity:** 99 percent.

**Methodology:** Triplet repeat-primed polymerase chain reaction (PCR) followed by size analysis using capillary electrophoresis. Methylation-specific PCR analysis is performed for CGG repeat lengths of **100** or greater. Methylation analysis is used to distinguish between premutation and full mutation alleles.

**Analytic Sensitivity and Specificity:** 99 percent.

**Limitations:** Diagnostic errors can occur due to rare sequence variations.

Phenotype	Number of CGG Repeats
Unaffected	<45
Intermediate	45-54
Premutation	55-200
Affected	>200

See Compliance Statement C: [www.aruplab.com/CS](http://www.aruplab.com/CS)

**Note:** If a CGG repeat of **100** or greater is detected by PCR and Capillary Electrophoresis, methylation analysis will be added. Additional charges apply.

**2009034**

**Fragile X (*FMRI*) with Reflex to Methylation Analysis, Fetal**

**FX PCR FE**

**Specimen Required:** Collect: **Fetal Specimen:** Amniotic fluid or two T-25 flasks at 80 percent confluency of cultured amniocytes. If the client is unable to culture amniocytes, this can be arranged by contacting ARUP Client Services at (800) 522-2787.

**AND Maternal Specimen:** Lavender (EDTA), pink (K<sub>2</sub>EDTA), or yellow (ACD Solution A or B).

Specimen Preparation: **Amniotic Fluid:** Transport 10 mL unspun fluid. (Min: 5 mL)

**Cultured Amniocytes:** Fill flasks with culture media. Transport two T-25 flasks at 80 percent confluency of cultured amniocytes.

Backup cultures must be retained at the client's institution until testing is complete.

**Maternal Specimen:** Transport 3 mL whole blood. (Min: 1 mL)

Storage/Transport Temperature: **Amniotic Fluid:** Room temperature.

**Cultured Amniocytes:** CRITICAL ROOM TEMPERATURE. Must be received within 48 hours of shipment due to viability of cells.

**Maternal Specimen:** Room temperature.

Remarks: Maternal specimen is recommended for proper test interpretation. Order Maternal Cell Contamination, Maternal Specimen. This can be arranged by contacting ARUP genetic counselors at (800) 242-2787 ext. 2141. Patient History Form is available on the ARUP Web site or by contacting ARUP Client Services at (800) 522-2787.

Stability (collection to initiation of testing): **Fetal Specimen:** Ambient: 48 hours; Refrigerated: Unacceptable; Frozen: Unacceptable

**Maternal Specimen:** Ambient: **1 week**; Refrigerated: **1 month**; Frozen: **6 months**

**Note:** If a CGG repeat of **100** or greater is detected by PCR and Capillary Electrophoresis; methylation analysis will be added. Additional charges apply.

HOTLINE: Effective August 19, 2019

**New Test**     [3001648](#)  
[Click for Pricing](#)

**Gaucher Disease (GBA) Sequencing**

**GBA FGS**



Patient History for Gaucher Disease Sequencing



Additional Technical Information



Out of Pocket Estimator

**Methodology:** Polymerase Chain Reaction/Sequencing  
**Performed:** Sun-Sat  
**Reported:** 2-3 weeks

**Specimen Required:** Collect: Lavender (K<sub>2</sub> or K<sub>3</sub> EDTA) or Pink (K<sub>2</sub> EDTA). Also acceptable: Yellow (ACD Solution A or B).  
Specimen Preparation: Transport 3 mL whole blood. (Min: 1 mL)  
Storage/Transport Temperature: Refrigerated.  
Stability (collection to initiation of testing): Ambient: 1 week; Refrigerated: 1 month; Frozen: 6 months

**Reference Interval:**

**Background information for Gaucher Disease (GBA) Sequencing:**

**Characteristics:** Gaucher disease (GD) is a lysosomal storage disorder with phenotypes ranging from perinatal lethality to lack of symptoms. There are three GD subtypes. Type 1 GD manifests with bone disease, hepatosplenomegaly, anemia, thrombocytopenia, and lung disease but no central nervous system (CNS) involvement. Type 2 GD exhibits CNS symptoms before age 2 and rapidly progresses resulting in death by age 4. Type 3 GD presents as early as age 2 with CNS symptoms that slowly progress resulting in death during the third or fourth decade.

**Incidence:** 1 in 900 Ashkenazi Jewish individuals; approximately 1 in 57,000 to 1 in 75,000 in general population.

**Inheritance:** Autosomal recessive.

**Cause:** Two pathogenic *GBA* variants on opposite chromosomes.

**Clinical Sensitivity:** 99 percent.

**Methodology:** Long range PCR followed by bidirectional sequencing of all coding regions and intron-exon boundaries of the *GBA* gene.

**Analytical Sensitivity and Specificity:** approximately 99 percent.

**Limitations:** Diagnostic errors can occur due to rare sequence variations. Regulatory region variants, deep intronic variants, large deletions/duplications/insertions, gene conversion and complex gene events may not be detected.

**Interpretive Data:** See Compliance Statement C: [www.aruplab.com/CS](http://www.aruplab.com/CS)

**CPT Code(s):** 81479

New York DOH approval pending. Call for status update.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

[3000258](#)

**Genetic Carrier Screen, (CF, FXS, and SMA) with Reflex to Methylation**

**CF FX SMA**

**Note:** Cystic Fibrosis: The 165-variant test includes the 23 pathogenic CF variants recommended by the American College of Medical Genetics for population carrier screening.

Fragile X: If a CGG repeat of 100 or greater is detected by PCR and Capillary Electrophoresis; methylation analysis will be added. Additional charges apply.

[0080135](#)

**Glucose-6-Phosphate Dehydrogenase**

**G6PD**

**Specimen Required:** Collect: Yellow (ACD Solution A). **Also acceptable:** Green (Sodium or Lithium Heparin), Lavender (K<sub>2</sub> EDTA or K<sub>3</sub> EDTA), or Pink (K<sub>2</sub>EDTA). Enzyme most stable in acid citrate dextrose (ACD).

Specimen Preparation: **Do not freeze.** Transport 3 mL whole blood. (Min: 1.5 mL)

Storage/Transport Temperature: Refrigerated.

Unacceptable Conditions: Clotted or hemolyzed specimens.

Stability (collection to initiation of testing): Ambient: 8 hours; Refrigerated: 1 week; Frozen: Unacceptable

HOTLINE: Effective August 19, 2019

<a href="#"><u>2013590</u></a>	<b>Heat Shock Protein 70, IgG by Immunoblot</b>	<b>HSP</b>
<b>Performed:</b>	<b>Thu</b>	
<b>Reported:</b>	<b>1-8 days</b>	
<a href="#"><u>2010476</u></a>	<b><i>Helicobacter pylori</i> Breath Test, Adult</b>	<b>UBIT</b>
<p><b>Specimen Required:</b> <u>Patient Prep:</u> This test requires the adult patient (&gt;17 years of age) to fast for 1 hour prior to test administration. The patient should not have taken antibiotics, proton pump inhibitors (e.g., Prilosec, Prevacid, Aciphex, Nexium), or bismuth preparations (e.g., Pepto-Bismol) within the previous 14 days. <b>The effect of histamine 2-receptor antagonists (e.g., Axid, Pepcid, Tagamet, Zantac) may reduce urease activity on urea breath tests and should be discontinued for 24-48 hours before the sample is collected.</b> When used to monitor treatment, the test should be performed four weeks after cessation of definitive therapy. The patient should be informed that the Pranactin-Citric drink that will be administered contains phenylalanine. Phenylketonurics restrict dietary phenylalanine.</p> <p><u>Collect:</u> BreathTek UBT Kit (ARUP Supply #51124 ) available online through eSupply using ARUP Connect™ or contact ARUPClient Services at (800) 522-2787.</p> <p><u>Specimen Preparation:</u> 1) Label breath collection bags with patient name, MRN, date and time of collection, and indicate Pre (blue) or Post (pink).</p> <p>2) Collect the baseline (Pre) breath specimen according to the instructions in the BreathTek UBT kit.</p> <p>3) After the allotted time, collect the Post breath specimen according to the instructions in the kit.</p> <p><u>Storage/Transport Temperature:</u> <b>CRITICAL ROOM TEMPERATURE. Do not freeze.</b></p> <p><u>Remarks:</u> <b>For a valid result, the post breath specimen must be collected between 15 and 30 minutes after the patient drinks the Pranactin-Citric solution.</b></p> <p><u>Unacceptable Conditions:</u> Underinflated bags. Pediatric specimens</p> <p><u>Stability (collection to initiation of testing):</u> Ambient: 1 week; Refrigerated: Unacceptable; Frozen: Unacceptable</p>		
<a href="#"><u>2010925</u></a>	<b><i>Helicobacter pylori</i> Breath Test, Pediatric</b>	<b>UBT PED</b>
<p><b>Specimen Required:</b> <u>Patient Prep:</u> This test requires the pediatric patient (3-17 years old) to fast for 1 hour prior to test administration. The patient should not have taken antibiotics, proton pump inhibitors (e.g., Prilosec, Prevacid, Aciphex, Nexium), or bismuth preparations (e.g., Pepto-Bismol) within the previous 14 days. <b>The effect of histamine 2-receptor antagonists (e.g., Axid, Pepcid, Tagamet, Zantac) may reduce urease activity on urea breath tests and should be discontinued for 24-48 hours before the sample is collected.</b> When used to monitor treatment, the test should be performed four weeks after cessation of definitive therapy. The patient should be informed that the Pranactin-Citric drink that will be administered contains phenylalanine. Phenylketonurics restrict dietary phenylalanine.</p> <p><u>Collect:</u> BreathTek UBT Kit. (ARUP Supply #51124) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.</p> <p><u>Specimen Preparation:</u> 1) Label breath collection bags with patient name, MRN, date and time of collection, and indicate Pre (blue) or Post (pink).</p> <p>2) Collect the baseline (Pre) breath specimen according to the instructions in the BreathTek UBT kit.</p> <p>3) After the allotted time, collect the Post breath specimen according to the instructions in the kit.</p> <p><u>Storage/Transport Temperature:</u> <b>CRITICAL ROOM TEMPERATURE. Do not freeze.</b></p> <p><u>Remarks:</u> <b>Submit with Order: Weight (in pounds), height (in inches), sex, and age.</b></p> <p><u>Unacceptable Conditions:</u> Underinflated bags. Specimens from patients younger than 3 years.</p> <p><u>Stability (collection to initiation of testing):</u> Ambient: 1 week; Refrigerated: Unacceptable; Frozen: Unacceptable</p>		
<a href="#"><u>2005792</u></a>	<b>Hemoglobin Evaluation Reflexive Cascade</b>	<b>HB CASCADE</b>
<p><b>HOTLINE NOTE:</b> There is a component change associated with this test.</p> <p>Remove component 2005799, Beta Globin (<i>HBB</i>) Mutations</p> <p>Remove component 2005803 Hereditary Persistent Fetal Hemoglobin</p> <p><b>There is also a reflexive pattern change associated with this test.</b></p> <p>Remove reflex to 2005828, Beta Globin (<i>HBB</i>) Mutations Bill</p> <p>Remove reflex to 2005836, Hereditary Persistent Fetal HGB Bill</p>		

HOTLINE: Effective August 19, 2019

**0092522**

***Histoplasma* Antigen **Quantitative** by EIA, Serum**

**HISTOAG S**

**Methodology:** Quantitative Enzyme Immunoassay  
**Performed:** Mon, Wed, Fri  
**Reported:** 1-4 days

**Specimen Required:** Collect: Plain Red or Serum Separator Tube (SST).  
Specimen Preparation: Transfer 2 mL serum to a sterile ARUP Standard Transport Tube (ARUP Supply #43115). (Min: 1 mL)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Specimen types other than those listed.  
Stability (collection to initiation of testing): Ambient: Unacceptable; Refrigerated: 2 weeks; Frozen: 1 month (avoid repeated freeze/thaw cycles)

**Reference Interval:**

Effective August 19, 2019

Less than 0.19 ng/ml	Not Detected
0.19-60.0 ng/mL	Detected
Greater than 60.0 ng/mL	Detected (above the limit of quantification).

**Interpretive Data:** The quantitative range of this assay is 0.19-60.0 ng/mL. Antigen concentrations greater than 60.0 ng/mL fall outside the linear range of the assay and cannot be accurately quantified.

This EIA test should be used in conjunction with other diagnostic procedures, including microbiological culture, histological examination of biopsy samples, and/or radiographic evidence, to aid in the diagnosis of histoplasmosis.

Crossreactivity with *Blastomyces dermatitidis*, *Coccidioides immitis*, and possibly *Talaromyces marneffe* have been observed with this EIA. Other clinically and geographically relevant endemic mycoses should be considered in the case of a positive test result.

Test developed and characteristics determined by ARUP Laboratories. See Compliance Statement B: aruplab.com/CS

**Note:** For **urine**, refer to test *Histoplasma Galactomannan* Antigen Quantitative by EIA, Urine (ARUP test code 2009418).

**HOTLINE NOTE:** There is a numeric map change associated with this test. There is a unit of measure change associated with this test.

Change the numeric map for component 0060749, *Histoplasma* Antigen, Serum from XXX.X to XX.XX.

Change the unit of measure for component 0060749, *Histoplasma* Antigen, Serum from U/mL to ng/mL.

**0062226**

***Histoplasma capsulatum* **Identification****

**MC HP**

**Methodology:** Nucleic Acid Probe/MALDI (Matrix-Assisted Laser Desorption/Ionization)  
**Performed:** Sun-Sat  
**Reported:** 2-7 days

**CPT Code(s):** Varies depending on identification methods.

HOTLINE: Effective August 19, 2019

<p><b>New Test</b> Available Now <a href="#">Click for Pricing</a></p>	<p><b><u>3001791</u></b></p>	<p><b>HNF-1B by Immunohistochemistry</b></p>	<p><b>HNF1B IHC</b></p>
<p><b>Methodology:</b> Immunohistochemistry  <b>Performed:</b> Mon-Fri  <b>Reported:</b> 1-3 days</p>			
<p><b>Specimen Required:</b> <u>Collect:</u> Tissue.  <u>Specimen Preparation:</u> Formalin fix (10 percent neutral buffered formalin) and paraffin embed specimen (cells must be prepared into a cellblock). Protect paraffin block and/or slides from excessive heat. Transport tissue block or 5 unstained (3- to 5-micron thick sections), positively charged slides in a tissue transport kit (recommended but not required), (ARUP supply #47808) available online through eSupply using ARUP Connect or contact ARUP Client Services at (800) 522-2787. (Min: 2 slides) If sending precut slides, do not oven bake.  <u>Storage/Transport Temperature:</u> Room temperature. Also acceptable: Refrigerated. Ship in cooled container during summer months.  <u>Unacceptable Conditions:</u> Specimens submitted with non-representative tissue type. Depleted specimens.  <u>Stability (collection to initiation of testing):</u> Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable</p>			
<p><b>Interpretive Data:</b> See Compliance Statement B: <a href="http://www.aruplab.com/CS">www.aruplab.com/CS</a></p>			
<p><b>Note:</b> All stains will be handled as "Stain and Return" unless a consultation is requested. To request a consultation, submit the pathology report, all associated case materials (clinical history, blocks, slides, etc.), and the Anatomic Pathology requisition form (#32960) in place of the Immunohistochemistry Stain Form.</p>			
<p><b>CPT Code(s):</b> 88342</p>			
<p>New York DOH Approved.</p>			
<p><b>HOTLINE NOTE:</b> Refer to the Test Mix Addendum for interface build information.</p>			

<p><b><u>2006444</u></b></p>	<p><b>IDH1 and IDH2 Mutation Analysis, exon 4</b></p>	<p><b>IDH1-2</b></p>
<p><b>Specimen Required:</b> <u>Collect:</u> Lavender (EDTA) or bone marrow (EDTA). <b>Also acceptable:</b> DNA extracted by CLIA certified lab.  <u>Specimen Preparation:</u> <b>Whole Blood:</b> Do not freeze. Transport 5 mL whole blood. (Min: 1 mL)  <b>Bone Marrow:</b> Do not freeze. Transport 3 mL bone marrow. (Min: 1 mL)  <b>Extracted DNA:</b> Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  <u>Storage/Transport Temperature:</u> Refrigerated.  <u>Unacceptable Conditions:</u> Plasma, serum, FFPE tissue blocks/slides, or frozen tissue, DNA extracted by a non-CLIA lab. Specimens collected in anticoagulants other than EDTA or sodium heparin. Clotted or grossly hemolyzed specimens.  <u>Stability (collection to initiation of testing):</u> Ambient: 24 hours; Refrigerated: 5 days; Frozen: Unacceptable  <b>Extracted DNA:</b> Ambient: 1 month; Refrigerate: Indefinitely; Frozen: Indefinitely</p>		
<p><b><u>2014188</u></b></p>	<p><b>IDH1 and IDH2 Mutation Analysis, Exon 4, Formalin-Fixed, Paraffin-Embedded (FFPE) Tissue</b></p>	<p><b>IDH1-2FFPE</b></p>
<p><b>Specimen Required:</b> <u>Collect:</u> Tumor tissue. <b>Also acceptable:</b> DNA extracted by CLIA certified lab with corresponding client-circled H&amp;E slide.  <u>Specimen Preparation:</u> <b>Tumor Tissue:</b> Formalin fix (10 percent neutral buffered formalin) and paraffin embed tissue. Protect from excessive heat. <b>Tissue block will be returned after testing.</b> Transport tissue block or 4 unstained 5-micron slides. (Min: 3 slides) Transport block and/or slide(s) in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  <b>Extracted DNA:</b> Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  <u>Storage/Transport Temperature:</u> Room temperature. Also acceptable: Refrigerated. Ship in cooled container during summer months.  <b>Extracted DNA:</b> Refrigerated.  <b>Remarks:</b> For FFPE specimens include surgical pathology report.  <u>Unacceptable Conditions:</u> Less than 25 percent tumor. DNA extracted by a non-CLIA lab. DNA extracted without a corresponding circled H&amp;E slide. Specimens fixed/processed in alternative fixatives (alcohol, Prefer) or heavy metal fixatives (B-4 or B-5). Decalcified specimens.  <u>Stability (collection to initiation of testing):</u> Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable  <b>Extracted DNA:</b> Ambient: 1 month; Refrigerated: Indefinitely; Frozen: Indefinitely</p>		

HOTLINE: Effective August 19, 2019

[0040227](#)

**IGHV Mutation Analysis by Sequencing**

**IGHV MUT**

**Specimen Required:** Collect: Lavender (EDTA) or bone marrow (EDTA). **Also acceptable: RNA extracted by CLIA certified lab.**  
Specimen Preparation: **Whole Blood:** Transport 5 mL whole blood. (Min: 1 mL)  
**Bone Marrow:** Transport 3 mL bone marrow. (Min: 1 mL)  
**Specimens must be received within 48 hours of collection due to lability of RNA.**  
**Extracted RNA:** Transport 40uL RNA with at least 40 ng/uL concentration (Min: 40uL). Transport RNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  
Storage/Transport Temperature: **Whole Blood or Bone Marrow:** CRITICAL REFRIGERATED. Separate specimens must be submitted when multiple tests are ordered.  
**Extracted RNA:** CRITICAL FROZEN. Separate specimens must be submitted when multiple tests are ordered.  
Unacceptable Conditions: Serum, plasma, CSF, extracted DNA, RNA extracted by a non-CLIA lab, bone core, or FFPE tissue.  
Specimens collected in anticoagulants other than EDTA. Severely hemolyzed or clotted specimens.  
Stability (collection to initiation of testing): Ambient: 1 hour; Refrigerated: 48 hours; Frozen: Unacceptable  
**Extracted RNA:** Ambient: Unacceptable; Refrigerate: Unacceptable; Frozen: Indefinitely

**New Test**

[3001409](#)

**Immunobullous Disease Panel, Epithelial Antibody Screening Panel**

**IMBULDZPAN**

Available Now  
[Click for Pricing](#)

**Methodology:** Indirect Fluorescent Antibody/Enzyme-Linked Immunosorbent Assay  
**Performed:** Varies  
**Reported:** 4-9 days

**Specimen Required:** Collect: Plain Red or Serum Separator Tube (SST).  
Specimen Preparation: Transfer 2 mL serum to an ARUP Standard Transport Tube. (Min: 0.5 mL)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Plasma  
Stability (collection to initiation of testing): Ambient: 1 week; Refrigerated: 2 weeks; Frozen: Indefinitely

**Reference Interval:** By Report

**Interpretive Data:** Refer to report

**Note:** For specimens less than 0.5 mL, call the Immunodermatology Laboratory at (866) 266-5699.

**CPT Code(s):** 88346, 88350 x5, 83516 x5

New York DOH approval pending. Call for status update.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.



**New Test**     [3001784](#)  
[Click for Pricing](#)

**Interstitial Lung Disease Autoantibody Panel**

**ILD PANEL**



**Additional Technical Information**

**Methodology:** Qualitative Immunoprecipitation/Semi-Quantitative Multiplex Bead Assay/Qualitative Immunoblot/Semi-Quantitative Enzyme-Linked Immunosorbent Assay/Quantitative Immunoturbidimetry  
**Performed:** Sun-Sat  
**Reported:** 7-18 days

**Specimen Required:** Collect: Serum Separator Tube (SST).  
Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer five 1 mL serum aliquots to ARUP Standard Transport Tubes. (Min: 0.5 mL/aliquot)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Hemolyzed, hyperlipemic, icteric, heat-treated or contaminated specimens.  
Stability (collection to initiation of testing): Ambient: 24 hours; Refrigerated: 1 weeks; Frozen: 1 month

**Reference Interval:**

Test Number	Components	Reference Interval	
2012074	SSA 52 and 60 (Ro) (ENA) Antibodies, IgG	Test Number	Components
			SSA-52 (Ro52) (ENA) Antibody, IgG
			SSA-60 (Ro60) (ENA) Antibody, IgG
		Reference Interval	
		29 AU/mL or Less: Negative	
		30-40 AU/mL: Equivocal	
		41 AU/mL or greater: Positive	
0050599	Scleroderma (Scl-70) (ENA) Antibody, IgG	29 AU/mL or less	Negative
		30-40 AU/mL	Equivocal
		41 AU/mL or greater	Positive
0099592	Jo-1 Antibody, IgG	29 AU/mL or less	Negative
		30-40 AU/mL	Equivocal
		41 AU/mL or greater	Positive
	PL-7 (threonyl-tRNA synthetase) Antibody	Negative	
	PL-12 (alanyl-tRNA synthetase) Antibody	Negative	
	EJ (glycyl-tRNA synthetase) Antibody	Negative	
	Ku Antibody	Negative	
	SRP (Signal Recognition Particle) Ab	Negative	
	OJ (isoleucyl-tRNA synthetase) Antibody	Negative	
2003040	PM/Scl-100 Antibody, IgG by Immunoblot	Negative	
	MDA5 (CADM-140) Ab	Negative	
	NXP2 (Nuclear matrix protein-2) Ab	Negative	
0050465	Rheumatoid Factor	0-14 IU/mL	
0055256	Cyclic Citrullinated Peptide (CCP) Antibody, IgG	19 Units or less	Negative
		20-39 Units	Weak positive
		40-59 Units	Moderate positive
		60 Units or Greater	Strong positive
3000082	Antinuclear Antibody (ANA) with HEp-2 Substrate, IgG by IFA	Less than 1:80	
2001601	RNA Polymerase III Antibody, IgG	19 Units or less	Negative
		20-39 Units	Weak Positive
		40-80 Units	Moderate Positive
		81 Units or greater	Strong Positive

**Interpretive Data:** Refer to report.  
 See Compliance Statement D: [www.aruplab.com/CS](http://www.aruplab.com/CS)

**Note:** Antibodies: Ro52, Ro60, Jo-1, PL-7, PL12, EJ, Ku, SRP, OJ, PM/Scl-100, MDA5, CCP, Scl-70, RA, ANA, NXP-2, RNA Polymerase III

**CPT Code(s):** 83516 x9; 86235 x5; 86200; 86431; 86039

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

---

<b>New Test</b>	<a href="#"><u>3001568</u></a>	<b>IRF4/DUSP22 (6p25) Gene Rearrangement by FISH</b>	<b>IRF4 FISH</b>
Available Now			
<a href="#">Click for Pricing</a>			

---



Additional Technical Information

**Methodology:** Fluorescence in situ Hybridization  
**Performed:** Varies  
**Reported:** 3-7 days

**Specimen Required:** Collect: Tumor tissue.

Specimen Preparation: Formalin fix (10 percent neutral buffered formalin) and paraffin embed tumor tissue. Transport tissue block or 4 unstained, consecutively cut, 5-micron thick sections, mounted on positively charged glass slides. (Min: 4 slides) Protect paraffin block and/or slides from excessive heat.

Storage/Transport Temperature: Room temperature. Also acceptable: Refrigerated.

Remarks: Include surgical pathology report with reason for referral.

Unacceptable Conditions: Specimens fixed or processed in alternative fixatives (alcohol, Prefer) or heavy metal fixatives (B-4 or B-5). No tumor in tissue. Decalcified specimens.

Stability (collection to initiation of testing): Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable

**Reference Interval:** By report

**Interpretive Data:** Refer to report.

See Compliance Statement B: [www.aruplab.com/CS](http://www.aruplab.com/CS)

**Note:** The laboratory will not reject specimens that arrive without a pathology report but will hold the specimen until this information is received.

**CPT Code(s):** 88366

New York DOH approval pending. Call for status update.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

---

<a href="#"><u>2002357</u></a>	<b>JAK2 Exon 12 Mutation Analysis by PCR</b>	<b>JAK2 EX12</b>
--------------------------------	--	------------------

---

**Specimen Required:** Collect: Lavender (EDTA) **or** bone marrow (EDTA). **Also acceptable:** DNA extracted by CLIA certified lab.

Specimen Preparation: **Whole Blood:** Do not freeze. Transport 5 mL whole blood. (Min: 1 mL)

**Bone Marrow:** Do not freeze. Transport 3 mL bone marrow. (Min: 1 mL)

**Extracted DNA:** Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.

Storage/Transport Temperature: Refrigerated.

Unacceptable Conditions: Plasma, serum, FFPE tissue blocks/slides, or frozen tissue, DNA extracted by a non-CLIA lab. Specimens collected in anticoagulants other than EDTA **or** sodium heparin. Clotted or grossly hemolyzed specimens.

Stability (collection to initiation of testing): Ambient: 24 hours; Refrigerated: 5 days; Frozen: Unacceptable

**Extracted DNA:** Ambient: 1 month; Refrigerate: Indefinitely; Frozen: Indefinitely

HOTLINE: Effective August 19, 2019

**0051245**

**JAK2 Gene, V617F Mutation, Qualitative**

**JAK2**

**Specimen Required:** Collect: Lavender (EDTA) **OR** bone marrow (EDTA). **Also acceptable:** DNA extracted by CLIA certified lab.  
Specimen Preparation: **Whole Blood:** Do not freeze. Transport 5 mL whole blood. (Min: 1 mL)  
**Bone Marrow:** Do not freeze. Transport 3 mL bone marrow. (Min: 1 mL)  
**Extracted DNA:** Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Plasma, serum, FFPE tissue blocks/slides, or frozen tissue, DNA extracted by a non-CLIA lab. Specimens collected in anticoagulants other than EDTA or sodium heparin. Clotted or grossly hemolyzed specimens.  
Stability (collection to initiation of testing): Ambient: 24 hours; Refrigerated: 5 days; Frozen: Unacceptable  
**Extracted DNA:** Ambient: 1 month; Refrigerate: Indefinitely; Frozen: Indefinitely

**HOTLINE NOTE:** Remove information found in the Reference Interval field.

**2012084**

**JAK2 Gene, V617F Mutation, Qualitative with Reflex to CALR (Calreticulin)  
Exon 9 Mutation Analysis by PCR with Reflex to MPL Mutation Detection**

**ET PMF RFX**

**Specimen Required:** Collect: Lavender (EDTA) or bone marrow (EDTA). **Also acceptable:** DNA extracted by CLIA certified lab.  
Specimen Preparation: **Whole Blood:** Do not freeze. Transport 5 mL whole blood. (Min: 1 mL)  
**Bone Marrow:** Do not freeze. Transport 3 mL bone marrow. (Min: 1 mL)  
**Extracted DNA:** Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Plasma, serum, FFPE tissue blocks/slides, or frozen tissue, DNA extracted by a non-CLIA lab. Specimens collected in anticoagulants other than EDTA or sodium heparin. Clotted or grossly hemolyzed specimens.  
Stability (collection to initiation of testing): Ambient: 24 hours; Refrigerated: 5 days; Frozen: Unacceptable  
**Extracted DNA:** Ambient: 1 month; Refrigerate: Indefinitely; Frozen: Indefinitely

**2012085**

**JAK2 Gene, V617F Mutation, Qualitative with Reflex to JAK2 Exon 12 Mutation  
Analysis by PCR**

**PV RFLX**

**Specimen Required:** Collect: Lavender (EDTA) or bone marrow (EDTA). **Also acceptable:** DNA extracted by CLIA certified lab.  
Specimen Preparation: **Whole Blood:** Do not freeze. Transport 5 mL whole blood. (Min: 1 mL)  
**Bone Marrow:** Do not freeze. Transport 3 mL bone marrow. (Min: 1 mL)  
**Extracted DNA:** Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Plasma, serum, FFPE tissue blocks/slides, or frozen tissue, DNA extracted by a non-CLIA lab. Specimens collected in anticoagulants other than EDTA or sodium heparin. Clotted or grossly hemolyzed specimens.  
Stability (collection to initiation of testing): Ambient: 24 hours; Refrigerated: 5 days; Frozen: Unacceptable  
**Extracted DNA:** Ambient: 1 month; Refrigerate: Indefinitely; Frozen: Indefinitely

**0040168**

**JAK2 Gene, V617F Mutation, Quantitative**

**JAK2 QNT**

**Specimen Required:** Collect: Lavender (EDTA). **Also acceptable:** DNA extracted by CLIA certified lab.  
Specimen Preparation: **Whole Blood:** Do not freeze. Transport 5 mL whole blood. (Min: 1 mL).  
**Extracted DNA:** Transport 40uL DNA with at least 50 ng/uL concentration (Min: 40uL). Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Plasma, serum, FFPE tissue blocks/slides, or frozen tissue, bone marrow, DNA extracted by a non-CLIA lab. Specimens collected in anticoagulants other than EDTA or sodium heparin. Clotted or grossly hemolyzed specimens.  
Stability (collection to initiation of testing): Ambient: 24 hours; Refrigerated: 48 hours; Frozen: Unacceptable  
**Extracted DNA:** Ambient: 1 month; Refrigerate: Indefinitely; Frozen: Indefinitely

**HOTLINE NOTE:** Remove information found in the Reference Interval field.

HOTLINE: Effective August 19, 2019

**0020843**

**Kidney Stone Risk Panel, Urine**

**KID**

**Specimen Required:** Collect: 24-hour urine. Refrigerate during collection.

Specimen Preparation: Thoroughly mix entire collection (24-hour) in one container. Transport four separate 4 mL aliquots of urine using Calculi Risk/Supersaturation Urine Collection Kit (ARUP supply# 46007). Available online through eSupply using ARUP Connect™ or contact Client Services at (800) 522-2787. Do not exceed 4 mL in tubes.

Aliquot according to the following specifications:

1st aliquot (pH 2): Transfer 4 mL urine into a Sulfamic Acid Tube. (Min: 4 mL) Mix well. Freeze immediately.

2nd aliquot (pH 2): Transfer 4 mL urine into a Sulfamic Acid Tube. (Min: 4 mL) Mix well. Freeze immediately.

3rd aliquot (pH 9): Transfer 4 mL urine into a Sodium Carbonate Tube. (Min: 4 mL) Mix well. Freeze immediately.

4th aliquot: Transfer 4 mL urine into an Unpreserved Tube. (Min: 4 mL) Freeze immediately.

If collection kit is unavailable, transport four 4 mL unadjusted aliquots of urine.

Storage/Transport Temperature: **CRITICAL FROZEN.** Separate specimens must be submitted when multiple tests are ordered. Ship patient and control specimens together.

Remarks: Record total volume and collection time interval on transport tube and test request form.

Stability (collection to initiation of testing): Ambient: Unacceptable; Refrigerated: Unacceptable; Frozen: 2 weeks

**3000440**

**KIT (D816V) Mutation by PCR**

**KIT D816V**

**Specimen Required:** Collect: Lavender (EDTA) or bone marrow (EDTA). Also acceptable: DNA extracted by CLIA certified lab.

Specimen Preparation: **Whole Blood: Do not freeze.** Transport 5 mL whole blood. (Min: 1 mL)

**Bone Marrow: Do not freeze.** Transport 3 mL bone marrow. (Min: 1 mL)

**Extracted DNA:** Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.

Storage/Transport Temperature: Refrigerated.

Unacceptable Conditions: Plasma, serum, FFPE tissue blocks/slides, or frozen tissue, DNA extracted by a non-CLIA lab. Specimens collected in anticoagulants other than EDTA or sodium heparin. Clotted or grossly hemolyzed specimens.

Stability (collection to initiation of testing): Ambient: 24 hours; Refrigerated: 5 days; Frozen: Unacceptable

**Extracted DNA: Ambient: 1 month; Refrigerate: Indefinitely; Frozen: Indefinitely**

**2002437**

**KIT Mutations in AML by Fragment Analysis and Sequencing**

**KIT AML**

**Specimen Required:** Collect: Lavender (EDTA) or bone marrow (EDTA). Also acceptable: DNA extracted by CLIA certified lab.

Specimen Preparation: **Whole Blood: Do not freeze.** Transport 5 mL whole blood. (Min: 1 mL)

**Bone Marrow: Do not freeze.** Transport 3 mL bone marrow (Min: 1 mL).

**Extracted DNA:** Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.

Storage/Transport Temperature: Refrigerated.

Unacceptable Conditions: Plasma, serum, FFPE tissue blocks/slides, or frozen tissue, DNA extracted by a non-CLIA lab. Specimens collected in anticoagulants other than EDTA or sodium heparin. Clotted or grossly hemolyzed specimens.

Stability (collection to initiation of testing): Ambient: 24 hours; Refrigerated: 5 days; Frozen: Unacceptable

**Extracted DNA: Ambient: 1 month; Refrigerated: Indefinitely; Frozen: Indefinitely**

**0040248**

**KRAS Mutation Detection**

**KRAS**

**Specimen Required:** Collect: Tumor tissue. Also acceptable: DNA extracted by CLIA certified lab with corresponding client-circled H&E slide.

Specimen Preparation: **Tumor Tissue:** Formalin fix (10 percent neutral buffered formalin) and paraffin embed tissue. Protect from excessive heat. Transport tissue block or 5 unstained 5-micron slides. (Min: 3 slides) Transport block(s) and/or slide(s) in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.

**Extracted DNA:** Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.

Storage/Transport Temperature: Room temperature. Also acceptable: Refrigerated. Ship in cooled container during summer months.

**Extracted DNA: Refrigerated.**

Remarks: Include surgical pathology report.

Unacceptable Conditions: Less than 25 percent tumor. DNA extracted by a non-CLIA lab. DNA extracted without a corresponding circled H&E slide. Specimens fixed/processed in alternative fixatives (alcohol, Prefer) or heavy metal fixatives (B-4 or B-5). Decalcified specimens.

Stability (collection to initiation of testing): Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable

**Extracted DNA: Ambient: 1 month; Refrigerate: Indefinitely; Frozen: Indefinitely**

HOTLINE: Effective August 19, 2019

**2001932**

**KRAS Mutation Detection with Reflex to BRAF Codon 600 Mutation Detection**

**KRAS RFLX**

**Specimen Required:** Collect: Tumor tissue. **Also acceptable:** DNA extracted by CLIA certified lab with corresponding client-circled H&E slide.  
Specimen Preparation: **Tumor Tissue:** Formalin fix (10 percent neutral buffered formalin) and paraffin embed tissue. Protect paraffin block from excessive heat. **Tissue block will be returned after testing.** Transport tissue block or 5 unstained 5-micron slides. (Min: 3 slides). Transport block and/or slide(s) in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  
**Extracted DNA:** Transport 40uL DNA with at least 50 ng/uL concentration. (Min: 40uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  
Storage/Transport Temperature: Room temperature. **Also acceptable:** Refrigerated. Ship in cooled container during summer months.  
**Extracted DNA:** Refrigerated.  
Remarks: Include surgical pathology report.  
Unacceptable Conditions: Less than 25 percent tumor. **DNA extracted by a non-CLIA lab. DNA extracted without a corresponding circled H&E slide.** Specimens fixed/processed in alternative fixatives (alcohol, Prefer) or heavy metal fixatives (B-4 or B-5). Decalcified specimens.  
Stability (collection to initiation of testing): **Ambient:** Indefinitely; **Refrigerated:** Indefinitely; **Frozen:** Unacceptable  
**Extracted DNA:** **Ambient:** 1 month; **Refrigerated:** Indefinitely; **Frozen:** Indefinitely

**New Test**  
[Click for Pricing](#)

**3001866**

**Krebs von den Lungen-6**

**KL 6**

**Methodology:** Quantitative Immunoturbidimetric  
**Performed:** Tue  
**Reported:** 1-8 days

**Specimen Required:** Collect: Plain Red.  
Specimen Preparation: Allow specimen to clot completely at room temperature. Separate from cells ASAP or within 2 hours of collection. Transfer 1.5 mL serum to ARUP Standard Transport Tube. (Min: 0.5 mL)  
Storage/Transport Temperature: Refrigerated.  
Unacceptable Conditions: Hemolyzed, hyperlipemic, icteric, heat-treated or contaminated.  
Stability (collection to initiation of testing): **Ambient:** 48 hours; **Refrigerated:** 2 weeks; **Frozen:** 1 month

**Reference Interval:** 0-500 U/mL

**Interpretive Data:** Krebs von den Lungen-6 (KL-6) is an indicator of lung inflammation or damage. Elevated levels are associated with various types of interstitial lung diseases (ILD). Abnormal KL-6 results should be interpreted in the appropriate clinical context. A negative result does not rule out ILD.

See Compliance Statement D: [www.aruplab.com/CS](http://www.aruplab.com/CS)

**Note:** Samples with results greater than 10,000 U/mL will be diluted to the maximum reportable range (>40,000 U/mL).

**CPT Code(s):** 83520

New York DOH approval pending. Call for status update.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

<b>New Test</b>	<b><u>3001780</u></b>	<b>Leukemia/Lymphoma Phenotyping Evaluation by Flow Cytometry</b>	<b>LL PANEL</b>
-----------------	-----------------------	---	-----------------

[Click for Pricing](#)



Time Sensitive



Additional Technical Information

**Methodology:** Flow Cytometry  
**Performed:** Sun-Sat  
**Reported:** 1-2 days

**Specimen Required:** Collect: Bone marrow. Whole blood: Green (Sodium Heparin), Lavender (K<sub>2</sub>EDTA), or Pink (K<sub>2</sub>EDTA). Tissue or fluid.

Specimen Preparation: **Bone Marrow:** Transport 1 mL heparinized bone marrow (Min: 0.5 mL\*)

**Whole Blood:** Transport 5 mL whole blood. (Min: 1mL\*)

**Tissue:** Transport 100 mg fresh tissue suspended in tissue culture media (e.g., RPMI 1640) (Min: 100 mg\*)

**Fluid:** Transport 10-100 mL fresh fluid (Min: 3 mL\*).

\*Minimum volume is dependent on cellularity.

Storage/Transport Temperature: **Specimen should be received within 24 hours of collection for optimal cell viability.**

**Bone Marrow or Whole Blood:** Room temperature. Also acceptable: Refrigerated.

**Tissue or Fluid:** Refrigerated.

Remarks: A minimum of 10,000 viable cells is required for flow cytometry phenotyping of samples containing a very limited number of markers (may also be called antibodies or antigens). For low-count specimens, supplying clinical and diagnostic information is especially important to help ensure the most appropriate marker combinations are evaluated before the specimen is depleted of cells.

**Bone Marrow or Whole Blood:** Provide specimen source, CBC, Wright stained smear (if available), clinical history, differential diagnosis, and any relevant pathology reports.

**Tissue or Fluid:** Provide specimen source, clinical history, differential diagnosis, and any relevant pathology reports.

**Follow up:** If previous leukemia/lymphoma phenotyping was performed at another lab, the outside flow cytometry report and histograms (if possible) should accompany the specimen.

Unacceptable Conditions: Clotted or hemolyzed specimens.

Stability (collection to initiation of testing): Ambient: 48 hours; Refrigerated: 48 hours; Frozen: Unacceptable

**Reference Interval:** By Report

**Interpretive Data:** Refer to report.

See Compliance Statement A: [www.aruplab.com/CS](http://www.aruplab.com/CS)

**Note:** Flow cytometric leukemia and lymphoma analysis may aid in identifying the tumor lineage for diagnostic and prognostic purposes. After review of the clinical history and morphology, a panel of markers is selected for each case by a board-certified hematopathologist. In most cases, the lineage can be identified as T-cell, B-cell, or myeloid and a diagnosis or differential diagnosis can be made.

Available Markers\*:

**T-cell:** CD1, CD2, CD3, CD4, CD5, CD7, CD8, TCR alpha-beta, TCR gamma-delta, Cytoplasmic CD3

**B-cell:** CD10, CD19, CD20, CD22, CD23, CD103, surface Kappa, surface Lambda, FMC7, Cytoplasmic Kappa, Cytoplasmic Lambda

**Myelo/Mono:** CD11b, CD13, CD14 (Mo2), CD14 (MY4), CD15, CD33, CD64, CD117, myeloperoxidase

**Misc:** CD11c, CD16, CD25, CD30, CD34, CD38, CD41, CD42b, CD45, CD56, CD57, CD61, HLA-DR, glycophorin, TdT, bcl-2, ALK-1, CD123, CD138, CD200, CD26, CD45, CRLF-2.

**\*Not all markers will be reported in all cases.** Requests for specific markers to be run must be listed on manual requisition or by footnote for electronic orders. We do not offer individual marker identification separately outside of the markers in this panel.

The report will include a pathologist interpretation and a marker interpretation range corresponding to CPT codes of 2-8 markers, 9-15 markers, and 16+ markers interpreted. Charges apply per marker.

**CPT Code(s):** 88184, 88185 each additional marker; 88187 or 88188 or 88189.

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

**0020421**

**Lipid Panel**

**CRISK**

**Interpretive Data:** An HDL cholesterol less than 40 mg/dL is low and constitutes a coronary heart disease risk factor. An HDL cholesterol greater than 60 mg/dL is a negative risk factor for coronary heart disease.

Non-HDL cholesterol is a secondary target of therapy in persons with high serum triglycerides (greater than 199 mg/dL). The goal for non-HDL cholesterol in persons with high triglycerides is 30 mg/dL higher than their LDL cholesterol goal.

**CHD Risk Factors**

- +1 Age: Men, 45 years and older  
Women, 55 years and older or premature menopause without estrogen therapy
- +1 Family history of premature CHD
- +1 Current smoking
- +1 Hypertension
- +1 Diabetes mellitus
- +1 Low HDL cholesterol: 39 mg/dL or less
- 1 High HDL cholesterol: 60 mg/dL or greater

Adult Values	Desirable	Borderline	Higher Risk
Total Cholesterol	199 mg/dL or less	200-239 mg/dL	240 mg/dL or greater
Triglycerides	149 mg/dL or less	150-199 mg/dL	200-499 mg/dL
HDL Cholesterol	40 mg/dL or greater		39 mg/dL or less
LDL Cholesterol (calculated)	129 mg/dL or less (99 mg/dL or less if patient has CHD)	130-159 mg/dL	160 mg/dL or greater
<i>removed</i>			
VLDL Cholesterol (calculated)	30 mg/dL or less		

Children & Adolescents	Desirable	Borderline	Higher Risk
Total Cholesterol	169 mg/dL or less	170-199 mg/dL	200 mg/dL or greater
Triglycerides	149 mg/dL or less	150-199 mg/dL	200-499 mg/dL
HDL Cholesterol	40 mg/dL or greater		39 mg/dL or less
LDL-Cholesterol	109 mg/dL or less	110-129 mg/dL	130 mg/dL or greater
<i>removed</i>			
VLDL Cholesterol (calculated)	30 mg/dL or less		

**HOTLINE NOTE:** There is a component change associated with this test.

Remove component 0095038, Appearance Chemistry

HOTLINE: Effective August 19, 2019

**0020468**

**Lipid Panel, Extended**

**CRISK E**

**Interpretive Data:** An HDL cholesterol less than 40 mg/dL is low and constitutes a coronary heart disease risk factor. An HDL cholesterol greater than 60 mg/dL is a negative risk factor for coronary heart disease.

Non-HDL cholesterol is a secondary target of therapy in persons with high serum triglycerides (greater than 199 mg/dL). The goal for non-HDL cholesterol in persons with high triglycerides is 30 mg/dL higher than their LDL cholesterol goal.

**CHD Risk Factors**

- +1 Age: Men, 45 years and older  
Women, 55 years and older or premature menopause without estrogen therapy
- +1 Family history of premature CHD
- +1 Current smoking
- +1 Hypertension
- +1 Diabetes mellitus
- +1 Low HDL cholesterol: 39 mg/dL or less
- 1 High HDL cholesterol: 60 mg/dL or greater

20 years and older	Desirable	Borderline	Higher Risk
Total Cholesterol	199 mg/dL or less	200-239 mg/dL	240 mg/dL or greater
Triglycerides	149 mg/dL or less	150-199 mg/dL	200-499 mg/dL
HDL Cholesterol	40 mg/dL or greater		39 mg/dL or less
LDL Cholesterol	129 mg/dL or less (99 mg/dL or less if patient has CHD)	130-159 mg/dL	160 mg/dL or greater
<i>removed</i>			
VLDL Cholesterol (calculated)	30 mg/dL or less		

0-19 years	Desirable	Borderline	High Risk
Total Cholesterol	169 mg/dL or less	170-199 mg/dL	200 mg/dL or greater
Triglycerides	149 mg/dL or less	150-199 mg/dL	200-499 mg/dL
HDL Cholesterol	40 mg/dL or greater		39 mg/dL or less
LDL-Cholesterol (measured)	109 mg/dL or less	110-129 mg/dL	130 mg/dL or greater
<i>removed</i>			
VLDL Cholesterol (calculated)	30 mg/dL or less		

**HOTLINE NOTE:** There is a component change associated with this test.

Remove component 0095038, Appearance Chemistry

**0091224**

**LSD, Urine - Screen with Reflex to Confirmation/Quantitation**

**LSD URN**

**Performed:** Varies  
**Reported:** 8-18 days

**Specimen Required:** Collect: Random urine.

Specimen Preparation: Protect from light. Transfer 2 mL urine to an ARUP Amber Transport Tube. (Min: 0.9 mL)

Storage/Transport Temperature: Refrigerated. Also acceptable: Room temperature or frozen.

Unacceptable Conditions: Specimens not protected from light. Specimens submitted in glass containers.

Stability (collection to initiation of testing): Ambient: 1 month; Refrigerated: 1 month; Frozen: 24 months

**0050119**

**Lupus Comprehensive Reflexive Panel**

**LUPUS COMP**

**Note:** Initial testing includes RF, C3, C4, and ANA. Specimens are screened for ANA using ELISA. If antibodies are detected, then an IFA titer will be added. If confirmed by IFA, then specimen will be tested for Thyroid Peroxidase (TPO) Antibody; Anti-Scl-70 (ENA), EIA; **Smith/RNP (ENA)** Antibody, IgG; Smith (ENA) Antibody, IgG; SSA 52 and 60 (Ro) (ENA) Antibodies, IgG; SSB (La) (ENA) Antibody, IgG; and Double-Stranded DNA (dsDNA) Antibody, IgG by ELISA; if Double-Stranded DNA (dsDNA) Antibody, IgG by ELISA result is Detected, then Double-Stranded DNA (dsDNA) Antibody, IgG by IFA (using *Crithidia luciliae*) is added. Additional charges apply.



HOTLINE: Effective August 19, 2019

**2009310**

**MGMT Promoter Methylation Detection**

**MGMT**

**Specimen Required:** Collect: Tumor tissue. **Also acceptable:** DNA extracted by CLIA certified lab with corresponding client-circled H&E slide.  
Specimen Preparation: **Tumor Tissue:** Formalin fix (10 percent neutral buffered formalin) and paraffin embed tissue. Protect from excessive heat. **Tissue block will be returned after testing.** Transport tissue block or 5 unstained 5-micron slides. (Min: 3 slides) Transport block and/or slide(s) in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  
**Extracted DNA:** Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  
Storage/Transport Temperature: Room temperature. Ship in cooled container during summer months.  
**Extracted DNA:** Refrigerated.  
Remarks: Include surgical pathology report.  
Unacceptable Conditions: Specimens fixed/processed in alternative fixatives (alcohol, Prefer) or heavy metal fixatives (B-4 or B-5). Decalcified specimens. Less than 25 percent tumor.  
Stability (collection to initiation of testing): Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable  
**Extracted DNA:** Ambient: 1 month; Refrigerated: Indefinitely; Frozen: Indefinitely

**0051740**

**Microsatellite Instability (MSI), HNPCC/Lynch Syndrome, by PCR**

**MSI PCR**

**Specimen Required:** Collect: Tumor **AND** normal epithelial tissue. **Also acceptable:** DNA extracted by CLIA certified lab.  
Specimen Preparation: **Tissue:** Formalin fix (10 percent neutral buffered formalin) and paraffin embed tissue. Protect from excessive heat. **Tissue block will be returned after testing.** Transport tissue block(s) or 10 unstained 5-micron slides (5 tumor and 5 normal epithelial). (Min: 3 tumor tissue and 3 normal epithelial tissue slides) Transport block(s) and/or slide(s) in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  
**Extracted DNA:** Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  
Storage/Transport Temperature: Room temperature. Also acceptable: Refrigerated. Ship in cooled container during summer months  
**Extracted DNA:** Refrigerated.  
Remarks: Include surgical pathology report.  
Unacceptable Conditions: Less than 25 percent tumor or less than 50 percent normal epithelial tissue. **DNA extracted by a non-CLIA lab. DNA extracted without a corresponding circled H&E slide.** Specimens fixed/processed in alternative fixatives (alcohol, Prefer) or heavy metal fixatives (B-4 or B-5). Decalcified specimens.  
Stability (collection to initiation of testing): Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable  
**Extracted DNA:** Ambient: 1 month; Refrigerated: Indefinitely; Frozen: Indefinitely

**2002499**

**MLH1 Promoter Methylation, Paraffin**

**MLH1PCR**

**Specimen Required:** Collect: Tumor tissue. **Also acceptable:** DNA extracted by CLIA certified lab with corresponding client-circled H&E slide.  
Specimen Preparation: **Tumor Tissue:** Formalin fix (10 percent neutral buffered formalin) and paraffin embed tissue. Protect from excessive heat. **Tissue block will be returned after testing.** Transport tissue block or 5 unstained 5-micron slides. (Min: 3 slides) Transport block and/or slide(s) in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  
**Extracted DNA:** Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  
Storage/Transport Temperature: Room temperature. Also acceptable: Refrigerated. Ship in cooled container during summer months.  
**Extracted DNA:** Refrigerated.  
Remarks: Include surgical pathology report.  
Unacceptable Conditions: Less than 25 percent tumor. **DNA extracted by a non-CLIA lab. DNA extracted without a corresponding circled H&E slide.** Specimens fixed/processed in alternative fixatives (alcohol, Prefer) or heavy metal fixatives (B-4 or B-5). Decalcified specimens.  
Stability (collection to initiation of testing): Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable  
**Extracted DNA:** Ambient: 1 month; Refrigerated: Indefinitely; Frozen: Indefinitely

**Interpretive Data:** Refer to report.

See Compliance Statement B: [www.aruplab.com/CS](http://www.aruplab.com/CS)

HOTLINE: Effective August 19, 2019

**0050615**

**Monoclonal Protein Detection Quantitation and Characterization, SPEP, IFE, IgA, IgG, IgM, Serum**

**IFE**

**Methodology:** Qualitative Immunofixation Electrophoresis/Quantitative Capillary Electrophoresis/Quantitative Nephelometry/**Quantitative Spectrophotometry**

**Specimen Required:** Collect: Serum Separator Tube (SST).

Specimen Preparation: **Separate from** cells ASAP or within 2 hours of collection. Transfer **1.5 mL** serum to an ARUP Standard Transport Tube. (Min: **1 mL**)

Storage/Transport Temperature: Refrigerated.

Unacceptable Conditions: Plasma.

Stability (collection to initiation of testing): After separation from cells: Ambient: **4 hours**; Refrigerated: 1 week; Frozen: 1 month

**Reference Interval:**

Test Number	Components	Reference Interval
0050640	Protein Electrophoresis, Serum	Effective August 19, 2019
		<b>Test Number</b> <b>Components</b> <b>Reference Interval</b>
		Total Protein, Serum      Refer to report
		Albumin      Refer to report
		Alpha-1 Globulins      Refer to report
		Alpha-2 Globulins      Refer to report
		Beta Globulins      Refer to report
		Gamma      Refer to report
0050340	Immunoglobulin A	Effective February 16, 2016
		0-30 days: 1-7 mg/dL      9-11 months: 16-83 mg/dL 1 month: 1-53 mg/dL      1 year: 14-105 mg/dL 2 months: 3-47 mg/dL      2 years: 14-122 mg/dL 3 months: 5-46 mg/dL      3 years: 22-157 mg/dL 4 months: 4-72 mg/dL      4 years: 25-152 mg/dL 5 months: 8-83 mg/dL      5-7 years: 33-200 mg/dL 6 months: 8-67 mg/dL      8-9 years: 45-234 mg/dL 7-8 months: 11-89 mg/dL      10 years and older: 68-408 mg/dL
0050350	Immunoglobulin G	0-30 days: 611-1542 mg/dL      9-11 months: 282-1026 mg/dL 1 month: 241-870 mg/dL      1 year: 331-1164 mg/dL 2 months: 198-577 mg/dL      2 years: 407-1009 mg/dL 3 months: 169-558 mg/dL      3 years: 423-1090 mg/dL 4 months: 188-536 mg/dL      4 years: 444-1187 mg/dL 5 months: 165-781 mg/dL      5-7 years: 608-1229 mg/dL 6 months: 206-676 mg/dL      8-9 years: 584-1509 mg/dL 7-8 months: 208-868 mg/dL      10 years and older: 768-1632 mg/dL
0050355	Immunoglobulin M	Effective February 16, 2016
		0-30 days: 0-24 mg/dL      9-11 months: 39-142 mg/dL 1 month: 19-83 mg/dL      1 year: 41-164 mg/dL 2 months: 16-100 mg/dL      2 years: 46-160 mg/dL 3 months: 23-85 mg/dL      3 years: 45-190 mg/dL 4 months: 26-96 mg/dL      4 years: 41-186 mg/dL 5 months: 31-103 mg/dL      5-7 years: 46-197 mg/dL 6 months: 33-97 mg/dL      8-9 years: 49-230 mg/dL 7-8 months: 32-120 mg/dL      10 years and older: 35-263 mg/dL

**CPT Code(s):** 82784 x3; **84155**; 84165; **86334**

**HOTLINE NOTE:** There is a clinically significant charting name change associated with this test.

Change the charting name for component 0050545 from Total Protein-Electrophoresis to **Total Protein, Serum**.

HOTLINE: Effective August 19, 2019

**2002715**

**Monoclonal Protein Detection, Quantitation, Characterization, SPEP, IFE, IgA, IgG, IgM, FLC**

**IFE FLC**

**Methodology:** Qualitative Immunofixation Electrophoresis/Quantitative Capillary Electrophoresis/Quantitative Nephelometry/**Quantitative Spectrophotometry**

**Specimen Required:** Collect: Serum Separator Tube (**SST**).

Specimen Preparation: **Separate from** cells ASAP or within 2 hours of collection. Transfer 2 mL serum to an ARUP Standard Transport Tube. (Min: **1 mL**)

Storage/Transport Temperature: Refrigerated.

Unacceptable Conditions: Plasma. Room temperature specimens.

Stability (collection to initiation of testing): After separation from cells: Ambient: Unacceptable; Refrigerated: 1 week; Frozen: 2 weeks

**Reference Interval:**

Test Number	Components	Reference Interval	
0050640	Protein Electrophoresis, Serum	Effective August 19, 2019	
		Test Number	Components
			Reference Interval
			Total Protein, Serum
			Refer to report
			Albumin
			Refer to report
			Alpha-1 Globulins
0050350	Immunoglobulin G		Refer to report
			Alpha-2 Globulins
			Refer to report
			Beta Globulins
			Refer to report
			Gamma
			Refer to report
0050340	Immunoglobulin A	Effective February 16, 2016	
		0-30 days: 1-7 mg/dL	9-11 months: 16-83 mg/dL
		1 month: 1-53 mg/dL	1 year: 14-105 mg/dL
		2 months: 3-47 mg/dL	2 years: 14-122 mg/dL
		3 months: 5-46 mg/dL	3 years: 22-157 mg/dL
		4 months: 4-72 mg/dL	4 years: 25-152 mg/dL
		5 months: 8-83 mg/dL	5-7 years: 33-200 mg/dL
		6 months: 8-67 mg/dL	8-9 years: 45-234 mg/dL
0050355	Immunoglobulin M	7-8 months: 11-89 mg/dL	10 years and older: 68-408 mg/dL
		Effective February 16, 2016	
		0-30 days: 0-24 mg/dL	9-11 months: 39-142 mg/dL
		1 month: 19-83 mg/dL	1 year: 41-164 mg/dL
		2 months: 16-100 mg/dL	2 years: 46-160 mg/dL
		3 months: 23-85 mg/dL	3 years: 45-190 mg/dL
		4 months: 26-96 mg/dL	4 years: 41-186 mg/dL
		5 months: 31-103 mg/dL	5-7 years: 46-197 mg/dL
	Kappa Quantitative Free Light Chains, Serum	6 months: 33-97 mg/dL	8-9 years: 49-230 mg/dL
		7-8 months: 32-120 mg/dL	10 years and older: 35-263 mg/dL
	Lambda Quantitative Free Light Chains, Serum	0.33 - 1.94 mg/dL	
	Kappa/Lambda Free Light Chain Ratio, Serum	0.57-2.63 mg/dL	
		0.26-1.65	

**CPT Code(s):** 82784 x3; **84155**; 84165; **86334**; **83883** x2

**HOTLINE NOTE:** There is a clinically significant charting name change associated with this test.

Change the charting name for component 0050545 from Total Protein-Electrophoresis to **Total Protein, Serum**.

**2007967**

**Motor and Sensory Neuropathy Evaluation with Immunofixation Electrophoresis and Reflex to Titer and Neuronal Immunoblot**

**MSNCR**

**Methodology:** Semi-Quantitative Enzyme-Linked Immunosorbent Assay/Semi-Quantitative Indirect Fluorescent Antibody/Qualitative Immunoblot/Quantitative Nephelometry/Quantitative Capillary Electrophoresis/Qualitative Immunofixation Electrophoresis/**Quantitative Spectrophotometry**

**Specimen Required:** Collect: Serum Separator Tube (SST).

**Specimen Preparation:** **Separate from** cells ASAP or within 2 hours of collection. Transfer 4 mL serum to an ARUP Standard Transport Tube. (Min: 2 mL)

**Storage/Transport Temperature:** Refrigerated.

**Unacceptable Conditions:** Plasma, CSF, or other body fluids. Contaminated, heat-inactivated, hemolyzed, severely icteric, or lipemic specimens.

**Stability (collection to initiation of testing):** Ambient: Unacceptable; Refrigerated: **1 week**; Frozen: **1 month**

**Reference Interval:**

Test Number	Components	Reference Interval	
	Albumin	3.75-5.01 g/dL	
	Alpha-1 Globulins	0.19-0.46 g/dL	
	Alpha-2 Globulins	0.48-1.05 g/dL	
	Beta Globulins	0.48-1.10 g/dL	
	Gamma	0.62-1.51 g/dL	
0050340	Immunoglobulin A	Effective February 16, 2016	
		0-30 days: 1-7 mg/dL 1 month: 1-53 mg/dL 2 months: 3-47 mg/dL 3 months: 5-46 mg/dL 4 months: 4-72 mg/dL 5 months: 8-83 mg/dL 6 months: 8-67 mg/dL 7-8 months: 11-89 mg/dL	9-11 months: 16-83 mg/dL 1 year: 14-105 mg/dL 2 years: 14-122 mg/dL 3 years: 22-157 mg/dL 4 years: 25-152 mg/dL 5-7 years: 33-200 mg/dL 8-9 years: 45-234 mg/dL 10 years and older: 68-408 mg/dL
0050350	Immunoglobulin G	0-30 days: 611-1542 mg/dL 1 month: 241-870 mg/dL 2 months: 198-577 mg/dL 3 months: 169-558 mg/dL 4 months: 188-536 mg/dL 5 months: 165-781 mg/dL 6 months: 206-676 mg/dL 7-8 months: 208-868 mg/dL	9-11 months: 282-1026 mg/dL 1 year: 331-1164 mg/dL 2 years: 407-1009 mg/dL 3 years: 423-1090 mg/dL 4 years: 444-1187 mg/dL 5-7 years: 608-1229 mg/dL 8-9 years: 584-1509 mg/dL 10 years and older: 768-1632 mg/dL
0050355	Immunoglobulin M	Effective February 16, 2016	
		0-30 days: 0-24 mg/dL 1 month: 19-83 mg/dL 2 months: 16-100 mg/dL 3 months: 23-85 mg/dL 4 months: 26-96 mg/dL 5 months: 31-103 mg/dL 6 months: 33-97 mg/dL 7-8 months: 32-120 mg/dL	9-11 months: 39-142 mg/dL 1 year: 41-164 mg/dL 2 years: 46-160 mg/dL 3 years: 45-190 mg/dL 4 years: 41-186 mg/dL 5-7 years: 46-197 mg/dL 8-9 years: 49-230 mg/dL 10 years and older: 35-263 mg/dL
	Total Protein, Serum	Effective August 19, 2019 Refer to Report	
	Asialo-GM1 Antibodies, IgG/IgM	29 IV or less 30-50 IV 51-100 IV 101 IV or greater	Negative Equivocal Positive Strong Positive
	GM1 Antibodies, IgG/IgM	29 IV or less 30-50 IV 51-100 IV 101 IV or greater	Negative Equivocal Positive Strong Positive
	GD1a Antibodies, IgG/IgM	29 IV or less 30-50 IV 51-100 IV 101 IV or greater	Negative Equivocal Positive Strong Positive
	GD1b Antibodies, IgG/IgM	29 IV or less 30-50 IV 51-100 IV 101 IV or greater	Negative Equivocal Positive Strong Positive
	GQ1b Antibodies, IgG/IgM	29 IV or less 30-50 IV 51-100 IV 101 IV or greater	Negative Equivocal Positive Strong Positive

HOTLINE: Effective August 19, 2019

0051284	Sulfate-3-Glucuronyl Paragloboside (SGPG) Antibody, IgM	Less than 1.00 IV		
0051285	Myelin Associated Glycoprotein (MAG) Antibody, IgM	Less than 1000 TU		
2007961	Paraneoplastic Antibodies (PCCA/ANNA) by IFA with Reflex to Titer and Immunoblot	<b>Test Number</b>	<b>Components</b>	<b>Reference Interval</b>
			Purkinje Cell/Neuronal Nuclear IgG Scrn	None Detected
			Neuronal Nuclear Antibody (ANNA) IFA Titer, IgG	Less than 1:10
			Purkinje Cell Antibody, Titer	Less than 1:10
2007963	Neuronal Nuclear Antibodies (Hu, Ri, Yo) IgG by Immunoblot	2007963	Neuronal Nuclear Antibodies (Hu, Ri, Yo) IgG by Immunoblot	Refer to report
2007963	Neuronal Nuclear Antibodies (Hu, Ri, Yo) IgG by Immunoblot	None Detected		

**CPT Code(s):** 83516 x7; 82784 x3; 84155; 84165; 86334; 86255 if reflexed add 83516 and/or 86256

**HOTLINE NOTE:** There is a clinically significant charting name change associated with this test.  
Change the charting name for component 0050545 from Total Protein-Electrophoresis to **Total Protein, Serum**.

**0051225**

**Motor Neuropathy Panel**

**MSN PAN**

**Methodology:** Semi-Quantitative Enzyme-Linked Immunosorbent Assay/Quantitative Nephelometry/Quantitative Capillary Electrophoresis/Qualitative Immunofixation Electrophoresis/**Quantitative Spectrophotometry**

**Specimen Required:** Collect: Serum Separator Tube (SST).

Specimen Preparation: **Separate from** cells ASAP or within 2 hours of collection. Transfer 4 mL serum to an ARUP Standard Transport Tube. (Min: 2 mL)

Storage/Transport Temperature: Refrigerated.

Unacceptable Conditions: Plasma, CSF, or other body fluids. Room temperature specimens. Contaminated, heat-inactivated, hemolyzed, severely icteric, or lipemic specimens.

Stability (collection to initiation of testing): Ambient: Unacceptable; Refrigerated: **1 week**; Frozen: **1 month**

**Reference Interval:**

Test Number	Components	Reference Interval	
	Asialo-GM1 Antibodies, IgG/IgM	29 IV or less	Negative
		30-50 IV	Equivocal
		51-100 IV	Positive
		101 IV or greater	Strong Positive
	GM1 Antibodies, IgG/IgM	29 IV or less	Negative
		30-50 IV	Equivocal
		51-100 IV	Positive
		101 IV or greater	Strong Positive
	GD1a Antibodies, IgG/IgM	29 IV or less	Negative
		30-50 IV	Equivocal
		51-100 IV	Positive
		101 IV or greater	Strong Positive
	GD1b Antibodies, IgG/IgM	29 IV or less	Negative
		30-50 IV	Equivocal
		51-100 IV	Positive
		101 IV or greater	Strong Positive
	GQ1b Antibodies, IgG/IgM	29 IV or less	Negative
		30-50 IV	Equivocal
		51-100 IV	Positive
		101 IV or greater	Strong Positive
	<b>Total Protein, Serum</b>	<b>Effective August 19, 2019</b> <b>Refer to Report</b>	
	Albumin	3.75-5.01 g/dL	
	Alpha-1 Globulins	0.19-0.46 g/dL	
	Alpha-2 Globulins	0.48-1.05 g/dL	
	Beta Globulins	0.48-1.10 g/dL	
	Gamma	0.62-1.51 g/dL	

HOTLINE: Effective August 19, 2019

0050340	Immunoglobulin A	Effective February 16, 2016	
		0-30 days: 1-7 mg/dL 1 month: 1-53 mg/dL 2 months: 3-47 mg/dL 3 months: 5-46 mg/dL 4 months: 4-72 mg/dL 5 months: 8-83 mg/dL 6 months: 8-67 mg/dL 7-8 months: 11-89 mg/dL	9-11 months: 16-83 mg/dL 1 year: 14-105 mg/dL 2 years: 14-122 mg/dL 3 years: 22-157 mg/dL 4 years: 25-152 mg/dL 5-7 years: 33-200 mg/dL 8-9 years: 45-234 mg/dL 10 years and older: 68-408 mg/dL
0050350	Immunoglobulin G	Effective February 16, 2016	
		0-30 days: 611-1542 mg/dL 1 month: 241-870 mg/dL 2 months: 198-577 mg/dL 3 months: 169-558 mg/dL 4 months: 188-536 mg/dL 5 months: 165-781 mg/dL 6 months: 206-676 mg/dL 7-8 months: 208-868 mg/dL	9-11 months: 282-1026 mg/dL 1 year: 331-1164 mg/dL 2 years: 407-1009 mg/dL 3 years: 423-1090 mg/dL 4 years: 444-1187 mg/dL 5-7 years: 608-1229 mg/dL 8-9 years: 584-1509 mg/dL 10 years and older: 768-1632 mg/dL
0050355	Immunoglobulin M	Effective February 16, 2016	
		0-30 days: 0-24 mg/dL 1 month: 19-83 mg/dL 2 months: 16-100 mg/dL 3 months: 23-85 mg/dL 4 months: 26-96 mg/dL 5 months: 31-103 mg/dL 6 months: 33-97 mg/dL 7-8 months: 32-120 mg/dL	9-11 months: 39-142 mg/dL 1 year: 41-164 mg/dL 2 years: 46-160 mg/dL 3 years: 45-190 mg/dL 4 years: 41-186 mg/dL 5-7 years: 46-197 mg/dL 8-9 years: 49-230 mg/dL 10 years and older: 35-263 mg/dL
0051285	Myelin Associated Glycoprotein (MAG) Antibody, IgM	Less than 1000 TU	
0051284	Sulfate-3-Glucuronyl Paragloboside (SGPG) Antibody, IgM	Less than 1.00 IV	

**CPT Code(s):** 83516 x7; 82784 x3; 84155; 84165; 86334

**HOTLINE NOTE:** There is a clinically significant charting name change associated with this test.  
Change the charting name for component 0050545 from Total Protein-Electrophoresis to **Total Protein, Serum**.

**2005545**

**MPL Mutation Detection by Capillary Electrophoresis**

**MPL**

**Specimen Required:** Collect: Lavender (EDTA) or bone marrow (EDTA). Also acceptable: DNA extracted by CLIA certified lab.  
**Specimen Preparation:** **Whole Blood:** Do not freeze. Transport 5 mL whole blood. (Min: 1 mL)  
**Bone Marrow:** Transport 3 mL bone marrow. (Min: 1 mL)  
**Extracted DNA:** Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  
**Storage/Transport Temperature:** Refrigerated.  
**Unacceptable Conditions:** Plasma, serum, FFPE tissue blocks/slides, or frozen tissue, DNA extracted by a non-CLIA lab. Specimens collected in anticoagulants other than EDTA or sodium heparin. Clotted or grossly hemolyzed specimens.  
**Stability (collection to initiation of testing):** Ambient: 24 hours; Refrigerated: 5 days; Frozen: Unacceptable  
**Extracted DNA:** Ambient: 1 month; Refrigerate: Indefinitely; Frozen: Indefinitely

**0050707**

**MPO/PR3 (ANCA) Antibodies**

**MPO/PR3**

**Reference Interval:**

Test Number	Components	Reference Interval	
0050526	Myeloperoxidase Antibody	Negative	19 AU/mL or less
		Equivocal	20-25 AU/mL
		Positive	26 AU/mL or greater
0050527	Serine Proteinase 3 Antibody	Negative	19 AU/mL or less
		Equivocal	20-25 AU/mL
		Positive	26 AU/mL or greater

**HOTLINE NOTE:** There is a clinically significant charting name change associated with this test.  
Change the charting name for component 0050527, Serine Protease 3, IgG to **Serine Proteinase 3, IgG**

HOTLINE: Effective August 19, 2019

**3000523**

**Mumps Virus by PCR**

**MPSPCR**

**Specimen Required:** Patient Prep: Patient should not eat, drink, smoke or chew gum for 30 minutes before collecting oral sample.

Collect: Buccal swab.

Specimen Preparation: Transfer buccal swab to viral transport media (ARUP supply #12884) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787. (Min: 0.5 mL)

Storage/Transport Temperature: Frozen.

Remarks: Specimen source required.

Unacceptable Conditions: **Urine. Nasopharyngeal swab.**

Stability (collection to initiation of testing): Ambient: 48 hours; Refrigerated: 1 week; Frozen: 1 week

**New Test**

**3001869**

**Myasthenia Gravis Reflexive Panel**

**MG R PAN**

[Click for Pricing](#)



#### Additional Technical Information

**Methodology:** Quantitative Radioimmunoassay/Semi-Quantitative Flow Cytometry

**Performed:** Sun-Sat

**Reported:** 3-8 days

**Specimen Required:** Collect: Serum Separator Tube (SST).

Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 1 mL serum to an ARUP Standard Transport Tube. (Min: 0.5 mL)

Storage/Transport Temperature: Refrigerated.

Unacceptable Conditions: Contaminated, hemolyzed, or severely lipemic specimens.

Stability (collection to initiation of testing): After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 month (avoid repeated freeze/thaw cycles)

#### Reference Interval:

Test Number	Components	Reference Interval	
0080009	Acetylcholine Receptor Binding Antibody	Negative	0.0-0.4 nmol/L
		Positive	0.5 nmol/L or greater
		Effective November 18, 2013	
0099580	Acetylcholine Receptor Blocking Antibody	Negative:	0-26% blocking
		Indeterminate:	27-41% blocking
		Positive:	42% or greater blocking
		Effective August 20, 2012	
0099521	Acetylcholine Receptor Modulating Antibody	Negative	0-45% modulating
		Positive	46% or greater modulating
		Effective August 20, 2012	
3001576	Muscle-Specific Kinase (MuSK) Antibody, IgG	Negative	0.00-0.03 nmol/L
		Positive	0.04 nmol/L or greater

**Interpretive Data:** Approximately 85-90 percent of patients with myasthenia gravis (MG) express antibodies to the acetylcholine receptor (AChR), which can be divided into binding, blocking, and modulating antibodies. Binding antibody can activate complement and lead to loss of AChR. Blocking antibody may impair binding of acetylcholine to the receptor, leading to poor muscle contraction. Modulating antibody causes receptor endocytosis resulting in loss of AChR expression, which correlates most closely with clinical severity of disease. Approximately 10-15 percent of individuals with confirmed myasthenia gravis have no measurable binding, blocking, or modulating antibodies.

See Compliance Statement B: [www.aruplab.com/CS](http://www.aruplab.com/CS)

**Note:** If Acetylcholine Receptor Binding Antibody result is greater than 0.4 nmol/L or Acetylcholine Receptor Blocking Antibody result is greater than 26 percent, then Acetylcholine Receptor Modulating Antibody (ARUP test code 0099521) will be added. If Acetylcholine Receptor Binding Antibody result is less than or equal to 0.4 nmol/L, then Muscle-Specific Kinase (MuSK) Ab, IgG (ARUP test code 3001576) will be added. Additional charges apply.

**CPT Code(s):** 83519; 83516; if reflexed, add 83516, 83519

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

---

<u><b>0009318</b></u>	<b>MYD88 L265P Mutation Detection by PCR, Quantitative</b>	<b>MYD88</b>
-----------------------	--	--------------

---

**Specimen Required:** Collect: Lavender (EDTA), bone marrow (EDTA), or tissue. **Also acceptable: DNA extracted by CLIA certified lab.**

**Specimen Preparation:** **Whole Blood:** **Do not freeze.** Transport 5 mL whole blood. (Min: 1 mL)

**Bone Marrow:** **Do not freeze.** Transport 3 mL bone marrow. (Min: 1 mL)

**FFPE Tumor Tissue:** Formalin fix (10 percent neutral buffered formalin) and paraffin embed tissue. Protect from excessive heat. Transport tissue in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.

**Extracted DNA:** Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.

**Storage/Transport Temperature:** **Whole Blood, Bone Marrow or Extracted DNA:** Refrigerated.

**FFPE Tumor Tissue:** Room temperature. Also acceptable: Refrigerated. Ship in cooled container during summer months.

**Unacceptable Conditions:** Plasma, serum, **FFPE tissue slides, frozen tissue, DNA extracted by a non-CLIA lab.** Specimens collected in anticoagulants other than EDTA **or sodium heparin.** Clotted or grossly hemolyzed specimens.

**FFPE Tumor Tissue:** Specimens fixed/processed in alternative fixatives or heavy metal fixatives (B-4 or B-5) or tissue sections on slides. Decalcified specimens.

**Stability (collection to initiation of testing):** **Whole Blood or Bone Marrow:** Ambient: 24 hours; Refrigerated: 5 days; Frozen: Unacceptable

**FFPE Tumor Tissue:** Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable

**Extracted DNA:** Ambient: 1 month; Refrigerated: Indefinitely; Frozen: Indefinitely

---

<u><b>0050526</b></u>	<b>Myeloperoxidase (MPO) Antibody</b>	<b>MPO-AB</b>
-----------------------	---------------------------------------	---------------

---

**HOTLINE NOTE:** Name change only.

---

<u><b>0050742</b></u>	<b>Myocardial Antibody, IgG with Reflex to Titer</b>	<b>MYO R</b>
-----------------------	--	--------------

---

**Performed:** Thu

**Reported:** 1-8 days



HOTLINE: Effective August 19, 2019

---

<b>New Test</b> Available Now <a href="#">Click for Pricing</a>	<a href="#"><b>3001907</b></a>	<b>Myotonic Dystrophy Type 1 (DMPK) CTG Expansion</b>	<b>DM1 PCR</b>
---	--------------------------------	---	----------------

---

**Methodology:** Polymerase Chain Reaction/Capillary Electrophoresis  
**Performed:** Varies  
**Reported:** 7-10 days

**Specimen Required:** Collect: Lavender (K<sub>2</sub>EDTA), Pink (K<sub>2</sub>EDTA), or Yellow (ACD Solution A or B).  
Specimen Preparation: Transport 5 mL whole blood. (Min: 3 mL)  
Storage/Transport Temperature: Refrigerated.  
Stability (collection to initiation of testing): Ambient: 1 week; Refrigerated: 1 month; Frozen: 6 months

**Reference Interval:** By report

**Interpretive Data:**

**Background Information for Myotonic Dystrophy Type 1 (DMPK):**

**Characteristics:** Myotonic dystrophy type 1 (DM1) is a multisystem disorder characterized by myotonic myopathy with involvement of the eye, heart, endocrine system and central nervous system. Clinical findings span a continuum from mild to severe, with overlap in the three recognized clinical subtypes of DM1: mild, classic and congenital. Mild DM1 is adult-onset and features include mild myotonia and premature cataracts or baldness. Onset of classic DM1 is typically between 10-30 years of age and findings include distal muscle weakness, myotonia, cataracts, GI disturbances, and cardiac conduction abnormalities. Congenital DM1 may present prenatally with polyhydramnios and reduced fetal movement, and postnatal features commonly include infantile hypotonia, respiratory insufficiency, facial diplegia, and intellectual disability.

**Prevalence:** 1:20,000.

**Inheritance:** Autosomal dominant.

**Penetrance:** Age-related, approaches 100 percent by age 50.

**Cause:** Expanded number of CTG repeats in the *DMPK* gene.

Normal: 5-34 CTG repeats, stably transmitted, not associated with DM1 manifestations.

Premutation: 35-49 CTG repeats, may be unstably transmitted, not associated with DM1 manifestations.

Full-penetrance disease allele: 50 or more CTG repeats, unstably transmitted, associated with DM1 manifestations.

**Clinical Sensitivity:** >99 percent for DM1.

**Methodology:** Triplet repeat-primed polymerase chain reaction (PCR) followed by size analysis using capillary electrophoresis to assess the CTG repeat in the *DMPK* 3' untranslated region. Specific allele sizing estimates cannot be determined for CTG repeats of >150. Repeat sizing precision is approximately +/- 2 repeats for alleles with 5-24 repeats and +/- 4 repeats for alleles with 77 to 150 repeats.

**Analytical Sensitivity and Specificity:** 99 percent.

**Limitations:** Diagnostic errors can occur due to rare sequence variations. This assay will not detect myotonic dystrophy type 2.

Phenotype	Number of CTG Repeats
Normal allele	Less than or equal to 34
Premutation	35 - 49
Mild	50 – approx. 150
Classic	approx.100 - approx 1000
Congenital	>1000

See Compliance Statement C: [www.aruplab.com/CS](http://www.aruplab.com/CS)

**CPT Code(s):** 81234

New York DOH approval pending. Call for status update.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

---

<a href="#"><b>0099465</b></a>	<b>Neuronal Cell Antibodies Quantitative, Serum</b>	<b>NEURON-SER</b>
--------------------------------	---	-------------------

---

**Performed:** Varies  
**Reported:** 3-8 days

HOTLINE: Effective August 19, 2019

0096657

**Neutrophil Oxidative Burst Assay (DHR)**

**DHR**

**Specimen Required:** Patient Prep: Collect control specimen from a healthy individual unrelated to patient at approximately the same time as and under similar conditions to the patient.  
Collect: Green (Sodium or Lithium Heparin) (patient) **AND** Green (Sodium or Lithium Heparin) (control). **Patient and control specimens must be collected within 48 hours of test performance.**  
Specimen Preparation: Transport 3 mL whole blood (patient) **AND** 3 mL whole blood (control) in original collection tubes. (Min: 1 mL (patient) **AND** 1 mL (control)) **Do not refrigerate or freeze. LIVE NEUTROPHILS REQUIRED.**  
Storage/Transport Temperature: **CRITICAL ROOM TEMPERATURE.**  
Stability (collection to initiation of testing): Ambient: 48 hours; Refrigerated: Unacceptable; Frozen: Unacceptable  
**New York State Clients:** Ambient 24 hours; Refrigerated: Unacceptable; Frozen: Unacceptable

**HOTLINE NOTE:** Remove information found in the Unacceptable Conditions field.

3000066

**NPM1 Mutation Detection by RT-PCR, Quantitative**

**NPM1 QNT**

**Specimen Required:** Collect: Lavender (EDTA) or bone marrow (EDTA). **Also acceptable: RNA extracted by CLIA certified lab.**  
Specimen Preparation: **Whole Blood:** Transport 5 mL whole blood. (Min: 1 mL)  
**Bone Marrow:** Transport 3 mL bone marrow. (Min: 1 mL)  
**Specimens must be received within 48 hours of collection due to lability of RNA.**  
**Extracted RNA:** Transport 40 uL RNA with at least 40 ng/uL concentration. (Min: 40 uL) Transport RNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  
Storage/Transport Temperature: **Whole Blood or Bone Marrow:** CRITICAL REFRIGERATED. Separate specimens must be submitted when multiple tests are ordered.  
**Extracted RNA:** CRITICAL FROZEN. Separate specimens must be submitted when multiple tests are ordered.  
Unacceptable Conditions: Serum, plasma, CSF, extracted DNA, RNA extracted by a non-CLIA lab, bone core, or FFPE tissue.  
Specimens collected in anticoagulants other than EDTA. Severely hemolyzed or clotted specimens.  
Stability (collection to initiation of testing): Ambient: 1 hour; Refrigerated: 48 hours; Frozen: Unacceptable  
**Extracted RNA:** Ambient: Unacceptable; Refrigerate: Unacceptable; Frozen: Indefinitely

2003123

**NRAS Mutation Detection by Pyrosequencing**

**NRAS**

**Specimen Required:** Collect: Tumor tissue. **Also acceptable: DNA extracted by CLIA certified lab with corresponding client-circled H&E slide.**  
Specimen Preparation: **Tumor Tissue:** Formalin fix (10 percent neutral buffered formalin) and paraffin embed tissue. Protect from excessive heat. **Tissue block will be returned after testing.** Transport tissue block or 5 unstained 5 micron slides. (Min: 3 slides). Transport block and/or slide(s) in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  
**Extracted DNA:** Transport 40 uL DNA with at least 50 ng/uL concentration. (Min: 40 uL) Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  
Storage/Transport Temperature: Room temperature. Ship in cooled container during summer months.  
**Extracted DNA:** Refrigerated.  
Remarks: Include surgical pathology report.  
Unacceptable Conditions: Less than 25 percent tumor. DNA extracted by a non-CLIA lab. DNA extracted without a corresponding circled H&E slide. Specimens fixed/processed in alternative fixatives (alcohol, Prefer) or heavy metal fixatives (B-4 or B-5). Decalcified specimens.  
Stability (collection to initiation of testing): Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable  
**Extracted DNA:** Ambient: 1 month; Refrigerated: Indefinitely; Frozen: Indefinitely

HOTLINE: Effective August 19, 2019

---

<b>New Test</b>	<a href="#"><u>3001760</u></a>	<b>Pancreatitis (<i>PRSSI</i>) Deletion/Duplication</b>	<b>PRSS1 DD</b>
<a href="#">Click for Pricing</a>			

---



Patient History for Pancreatitis (*PRSSI*)  
Del/Dup



Additional Technical Information

**Methodology:** Multiplex Ligation-dependent Probe Amplification  
**Performed:** Varies  
**Reported:** 12-14 days

**Specimen Required:** Collect: Lavender (K<sub>2</sub>EDTA), Pink (K<sub>2</sub>EDTA), or Yellow (ACD Solution A or B).  
Specimen Preparation: Transport 3 mL whole blood. (Min: 2 mL)  
Storage/Transport Temperature: Refrigerated.  
Stability (collection to initiation of testing): Ambient: 1 week; Refrigerated: 1 month; Frozen: 6 months

**Reference Interval:** By Report

**Interpretive Data:**

**Background Information for Pancreatitis (*PRSSI*) Deletion/Duplication:**

**Characteristics:** Hereditary pancreatitis typically presents in late childhood with recurrent episodes of pancreatic inflammation, abdominal pain, nausea, and vomiting. Ultimately, this evolves into chronic pancreatitis resulting in permanent pancreatic damage.

**Epidemiology:** Incidence of chronic pancreatitis is 5-12 in 100,000 per year and prevalence is approximately 50 in 100,000.

**Inheritance of SPINK1-related pancreatitis:** Autosomal recessive and possibly digenic.

**Penetrance:** Variable.

**Cause:** Pathogenic variants in *PRSSI*, *SPINK1*, *CFTR*, *CASR*, *CTRC*, *CPA1* and *CLDN2* genes are associated with pancreatitis.

**Clinical Sensitivity:** Unknown.

**Methodology:** Multiplex ligation-dependent probe amplification (MLPA) of the *PRSSI* gene.

**Analytical Sensitivity and Specificity:** 99 percent.

**Limitations:** Diagnostic errors can occur due to rare sequence variations. Single base pair substitutions, small deletions/duplications, regulatory region and deep intronic variants will not be detected. Deletion/duplication breakpoints will not be determined. Variants in genes other than *PRSSI* will not be detected.

Counseling and informed consent are recommended for genetic testing. Consent forms are available online at [www.aruplab.com](http://www.aruplab.com)  
 See Compliance Statement C: [www.aruplab.com/CS](http://www.aruplab.com/CS)

New York DOH approval pending. Call for status update.

**CPT Code(s):** 81479

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

---

<b>New Test</b>	<a href="#"><u>3001768</u></a>	<b>Pancreatitis (<i>PRSSI</i>) Sequencing and Deletion/Duplication</b>	<b>PRSSI FGA</b>
<a href="#">Click for Pricing</a>			

---



Patient History for Pancreatitis Testing



Additional Technical Information



Out of Pocket Estimator

**Methodology:** Polymerase Chain Reaction/Sequencing and Multiplex Ligation Dependent Probe Amplification  
**Performed:** Varies  
**Reported:** 2-3 weeks

**Specimen Required:** Collect: Lavender (K<sub>2</sub>EDTA), Pink (K<sub>2</sub>EDTA), or Yellow (ACD Solution A or B).  
Specimen Preparation: Transport 3 mL whole blood. (Min: 2 mL)  
Storage/Transport Temperature: Refrigerated.  
Stability (collection to initiation of testing): Ambient: 1 week; Refrigerated: 1 month; Frozen: 6 months

**Interpretive Data:**

**Background Information for Pancreatitis (*PRSSI*) Sequencing and Deletion/Duplication**

**Characteristics:** Characteristics of *PRSSI*-related hereditary pancreatitis: Recurrent episodes of pancreatic inflammation that typically begin to present in late childhood, often with signs and symptoms including abdominal pain, nausea, and vomiting. Ultimately, these recurrent episodes of acute pancreatitis progress to permanent damage of the pancreas.

**Epidemiology:** Incidence of chronic pancreatitis: 5-12 in 100,000 per year  
 Prevalence of chronic pancreatitis: approximately 50 in 100,000

**Inheritance:** Autosomal dominant.

**Penetrance:** Varies geographically; estimated at 80 percent in the US.

**Cause:** Pathogenic variants in the cationic trypsinogen (*PRSSI*) gene.

**Clinical Sensitivity:** 15 percent of hereditary pancreatitis is caused by pathogenic *PRSSI* copy number variants or sequence variants.

**Methodology:** Bidirectional sequencing of *PRSSI* coding regions and intron/exon boundaries and multiplex ligation-dependent probe amplification (MLPA) of the *PRSSI* gene.

**Analytical Sensitivity and Specificity:** 99 percent.

**Limitations:** Diagnostic errors can occur due to rare sequence variations. Regulatory region variants and deep intronic variants will not be detected. The breakpoints of large deletions/duplications will not be determined. Variants in genes other than *PRSSI* are not evaluated.

Counseling and informed consent are recommended for genetic testing. Consent forms are available online at [www.aruplab.com](http://www.aruplab.com)  
 See Compliance Statement C: [www.aruplab.com/CS](http://www.aruplab.com/CS)

New York DOH approval pending. Call for status update.

**CPT Code(s):** 81404, 81479

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

---

<b>New Test</b> <a href="#">Click for Pricing</a>	<a href="#"><b>3001764</b></a>	<b>Pancreatitis (<i>SPINK1</i>) Deletion/Duplication</b>	<b>SPINK1 DD</b>
--	--------------------------------	--	------------------

---



Patient History for Pancreatitis (*SPINK1*)  
Del/Dup



Additional Technical Information

**Methodology:** Multiplex Ligation-dependent Probe Amplification  
**Performed:** Varies  
**Reported:** 12-14 days

**Specimen Required:** Collect: Lavender (K<sub>2</sub>EDTA), Pink (K<sub>2</sub>EDTA), or Yellow (ACD Solution A or B).  
Specimen Preparation: Transport 3 mL whole blood. (Min: 2 mL)  
Storage/Transport Temperature: Refrigerated.  
Stability (collection to initiation of testing): Ambient: 1 week; Refrigerated: 1 month; Frozen: 6 months

**Reference Interval:** By Report

**Interpretive Data:**

**Background Information for Pancreatitis (*SPINK1*) Deletion/Duplication:**

**Characteristics:** Hereditary pancreatitis typically presents with recurrent episodes of pancreatic inflammation, abdominal pain, nausea, and vomiting. Ultimately, this evolves into chronic pancreatitis resulting in permanent pancreatic damage.

**Epidemiology:** Incidence of chronic pancreatitis is 5-12 in 100,000 per year and prevalence is approximately 50 in 100,000.

**Inheritance of *SPINK1*-related pancreatitis:** Autosomal dominant.

**Penetrance:** Variable.

**Cause:** Pathogenic variants in *SPINK1*, *PRSS1*, *CFTR*, *CASR*, *CTRC*, *CPA1* and *CLDN2* genes are associated with pancreatitis.

**Clinical Sensitivity:** 6 percent of hereditary pancreatitis is caused by pathogenic *SPINK1* copy number variants.

**Methodology:** Multiplex ligation-dependent probe amplification (MLPA)

**Analytical Sensitivity and Specificity:** 99 percent.

**Limitations:** Diagnostic errors can occur due to rare sequence variations. Single base pair substitutions, small deletions/duplications, regulatory region and deep intronic variants will not be detected. Deletion/duplication breakpoints will not be determined. Variants in genes other than *SPINK1* will not be detected.

Counseling and informed consent are recommended for genetic testing. Consent forms are available online at [www.aruplab.com](http://www.aruplab.com).  
 See Compliance Statement C: [www.aruplab.com/CS](http://www.aruplab.com/CS)

**CPT Code(s):** 81479

New York DOH approval pending. Call for status update.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

---

<a href="#"><b>3000496</b></a>	<b>PanFungal Identification by Sequencing</b>	<b>PANFUNGSEQ</b>
--------------------------------	---	-------------------

---

**Specimen Required:** Collect: Tissue.  
Specimen Preparation: Transfer fresh tissue to a sterile container and freeze immediately. (Min: 25 mg) Also acceptable: Formalin-fixed paraffin-embedded (FFPE) tissue. (Min: 3 10-micron thick sections)  
Storage/Transport Temperature: **Fresh Tissue:** Frozen.  
**FFPE:** Room temperature.  
 Unacceptable Conditions: **Finger nails, toe nails, or bone.** Formalin-fixed paraffin-embedded tissue on slides.  
Stability (collection to initiation of testing): **Fresh Tissue:** Ambient: 5 days; Refrigerated: 5 days; Frozen: 5 days  
**FFPE:** Ambient: 1 month; Refrigerated: 1 month; Frozen: Unacceptable

---

<a href="#"><b>3001170</b></a>	<b>Platelet Antigen 1 Genotyping (HPA-1)</b>	<b>HPA-1 GENO</b>
--------------------------------	--	-------------------

---

**Performed:** Mon, Thu  
**Reported:** 7-14 days

HOTLINE: Effective August 19, 2019

<u><a href="#">3000193</a></u>	<b>Platelet Antigen Genotyping Panel</b>	<b>HPA GENO</b>
<b>Performed:</b>	Varies	
<b>Reported:</b>	7-14 days	
<u><a href="#">2002871</a></u>	<b><i>PML-RARA</i> Translocation, t(15;17) by RT-PCR, Quantitative</b>	<b>PML QNT</b>
<p><b>Specimen Required:</b> <u>Collect:</u> Lavender (EDTA) or bone marrow (EDTA). <b>Also acceptable:</b> RNA extracted by CLIA certified lab.</p> <p><u>Specimen Preparation:</u> <b>Whole Blood:</b> Transport 5 mL whole blood. (Min: 1 mL)</p> <p><b>Bone Marrow:</b> Transport 3 mL bone marrow. (Min: 1 mL)</p> <p><b>Specimens must be received within 48 hours of collection due to lability of RNA.</b></p> <p><b>Extracted RNA:</b> Transport 40uL RNA with at least 40 ng/uL concentration (Min: 40uL). Transport RNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.</p> <p><u>Storage/Transport Temperature:</u> <b>Whole Blood and Bone Marrow:</b> CRITICAL REFRIGERATED. Separate specimens must be submitted when multiple tests are ordered.</p> <p><b>Extracted RNA:</b> CRITICAL FROZEN. Separate specimens must be submitted when multiple tests are ordered.</p> <p><u>Unacceptable Conditions:</u> Serum, plasma, CSF, extracted DNA, RNA extracted by a non-CLIA lab, bone core, or FFPE tissue.</p> <p>Specimens collected in anticoagulants other than EDTA. Severely hemolyzed or clotted specimens.</p> <p><u>Stability (collection to initiation of testing):</u> Ambient: 1 hour; Refrigerated: 48 hours; Frozen: Unacceptable</p> <p><b>Extracted RNA:</b> Ambient: Unacceptable; Refrigerate: Unacceptable; Frozen: Indefinitely</p>		
<b>HOTLINE NOTE:</b> Remove information found in the Note field.		
<u><a href="#">2009226</a></u>	<b><i>Pneumocystis jirovecii</i> DFA with Reflex to <i>Pneumocystis jirovecii</i> by PCR</b>	<b>PNEUMST R</b>
<b>Performed:</b>	Sun-Sat	
<b>Reported:</b>	1-6 days	

HOTLINE: Effective August 19, 2019

**2002109**

**Protein Electrophoresis with Reflex to Immunofixation Electrophoresis  
Monoclonal Protein Detection, Quantitation & Characterization, IgA, IgG, and  
IgM, Serum**

**SPEP REFLEX**

**Methodology:** Quantitative Capillary Electrophoresis/Qualitative Immunofixation Electrophoresis/Quantitative Nephelometry/**Quantitative Spectrophotometry**

**Specimen Required:** Collect: Serum Separator Tube (SST).

**Specimen Preparation:** Separate from cells ASAP or within 2 hours of collection. Transfer 1.5 mL serum to an ARUP Standard Transport Tube. (Min: 1 mL)

**Storage/Transport Temperature:** Refrigerated.

**Unacceptable Conditions:** Plasma.

**Stability (collection to initiation of testing):** After separation from cells: Ambient: 4 hours; Refrigerated: 1 week; Frozen: 1 month

**Reference Interval:**

Test Number	Components	Reference Interval	
0050640	Protein Electrophoresis, Serum	Effective August 19, 2019	
		Test Number	Components
		Reference Interval	
		Total Protein, Serum	Refer to report
		Albumin	Refer to report
		Alpha-1 Globulins	Refer to report
		Alpha-2 Globulins	Refer to report
0050350	Immunoglobulin G	Beta Globulins	Refer to report
		Gamma	Refer to report
		0-30 days: 611-1542 mg/dL	9-11 months: 282-1026 mg/dL
		1 month: 241-870 mg/dL	1 year: 331-1164 mg/dL
		2 months: 198-577 mg/dL	2 years: 407-1009 mg/dL
		3 months: 169-558 mg/dL	3 years: 423-1090 mg/dL
		4 months: 188-536 mg/dL	4 years: 444-1187 mg/dL
0050340	Immunoglobulin A	5 months: 165-781 mg/dL	5-7 years: 608-1229 mg/dL
		6 months: 206-676 mg/dL	8-9 years: 584-1509 mg/dL
		7-8 months: 208-868 mg/dL	10 years and older: 768-1632 mg/dL
		Effective February 16, 2016	
		0-30 days: 1-7 mg/dL	9-11 months: 16-83 mg/dL
		1 month: 1-53 mg/dL	1 year: 14-105 mg/dL
		2 months: 3-47 mg/dL	2 years: 14-122 mg/dL
0050355	Immunoglobulin M	3 months: 5-46 mg/dL	3 years: 22-157 mg/dL
		4 months: 4-72 mg/dL	4 years: 25-152 mg/dL
		5 months: 8-83 mg/dL	5-7 years: 33-200 mg/dL
		6 months: 8-67 mg/dL	8-9 years: 45-234 mg/dL
		7-8 months: 11-89 mg/dL	10 years and older: 68-408 mg/dL
		Effective February 16, 2016	
		0-30 days: 0-24 mg/dL	9-11 months: 39-142 mg/dL
		1 month: 19-83 mg/dL	1 year: 41-164 mg/dL
		2 months: 16-100 mg/dL	2 years: 46-160 mg/dL
		3 months: 23-85 mg/dL	3 years: 45-190 mg/dL
		4 months: 26-96 mg/dL	4 years: 41-186 mg/dL
		5 months: 31-103 mg/dL	5-7 years: 46-197 mg/dL
		6 months: 33-97 mg/dL	8-9 years: 49-230 mg/dL
		7-8 months: 32-120 mg/dL	10 years and older: 35-263 mg/dL

**CPT Code(s):** 84155; 84165; if reflexed, add 82784 x3; 86334

**HOTLINE NOTE:** There is a clinically significant charting name change associated with this test.

Change the charting name for component 0050545 from Total Protein-Electrophoresis to **Total Protein, Serum**.

HOTLINE: Effective August 19, 2019

**0050640**

**Protein Electrophoresis, Serum**

**SPEP**

**Methodology:** Quantitative Capillary Electrophoresis/Quantitative Spectrophotometry

**Specimen Required:** Collect: Serum Separator Tube (SST).

Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 1.5 mL serum to an ARUP Standard Transport Tube. (Min: 1 mL)

Storage/Transport Temperature: Refrigerated.

Unacceptable Conditions: Plasma.

Stability (collection to initiation of testing): After separation from cells: Ambient: 4 hours; Refrigerated: 1 week; Frozen: 1 month

**Reference Interval:**

Effective August 19, 2019

Test Number	Components	Reference Interval
	Total Protein, Serum	Refer to Report
	Albumin	3.75-5.01 g/dL
	Alpha-1 Globulins	0.19-0.46 g/dL
	Alpha-2 Globulins	0.48-1.05 g/dL
	Beta Globulins	0.48-1.10 g/dL
	Gamma	0.62-1.51 g/dL

**CPT Code(s):** 84155; 84165

**HOTLINE NOTE:** There is a clinically significant charting name change associated with this test.

Change the charting name for component 0050545 from Total Protein-Electrophoresis to **Total Protein, Serum**.

**3000010**

**Relapsing Fever *Borrelia* Species by PCR**

**RFBPCR**

**Specimen Required:** Collect: Lavender (K<sub>2</sub>EDTA) or Pink (K<sub>2</sub>EDTA).

Specimen Preparation: Transport 1 mL whole blood. (Min: 0.5 mL)

Storage/Transport Temperature: Refrigerated.

Remarks: Specimen source required.

Unacceptable Conditions: Serum or plasma. Heparinized specimens.

Stability (collection to initiation of testing): Ambient: 24 hours; Refrigerated: 5 days; Frozen: 1 week

**Note:** This test is designed to detect but not differentiate the nucleic acid from *B. hermsii*, *B. miyamotoi*, *B. parkeri*, and *B. turicatae*. Additional less-frequently encountered relapsing fever *Borrelia* species may also be detected, including *B. recurrentis*, *B. coriaceae*, *B. theileri*, *B. lonestari*, and *B. anserina*. A result of "Detected" indicates the presence of nucleic acid from any one of these species in the specimen.

**0051368**

**RhD Gene (RHD) Copy Number**

**RHD**

**Interpretive Data:**

**Background Information for RhD Gene (RHD) Copy Number:**

**Characteristics:** Fetal or neonatal erythroblastosis and hydrops.

**Incidence of RHD-negative genotype:** 15 percent Caucasians, 5 percent African Americans, less than 1 percent Asians

**Inheritance:** Autosomal recessive

**Cause:** Maternal-fetal Rh D antigen incompatibility

**Methods:** Determine the presence of the *RHD* exons 5, 7, and a 37 base pair insertion in the intron 3/exon 4 boundary by PCR and fluorescence monitoring. Allelic height ratios are used to determine the number of copies of *RHD* as compared to *RHCE*.

**Limitations:** Bloody amniotic fluid specimens may give false-negative results because of maternal cell contamination; specificity may be compromised by mutations in primer sites or those outside the *RHD* exons examined; fetuses predicted to be unaffected should continue to be monitored by noninvasive means. Diagnostic errors can occur due to rare sequence variations.

**Analytic Sensitivity and Specificity:** Greater than 99 percent

**Clinical Sensitivity:** Greater than 98 percent

For quality assurance purposes, ARUP Laboratories will confirm the above result at no charge following delivery. Order Confirmation of Fetal Testing and include a copy of the original fetal report (or the mother's name and date of birth) with the test submission. Please contact an ARUP genetic counselor at (800) 242-2787 extension 2141 prior to specimen submission.

See Compliance Statement C: [www.aruplab.com/CS](http://www.aruplab.com/CS)

**HOTLINE NOTE:** There is a clinically significant charting name change associated with this test.

Change the charting name for component 0051369, RhD Antigen (RhD) Specimen to **RhD Gene (RHD) Copy Number Specimen**.

Change the charting name for component 0050422 RhD, Antigen (RhD) Genotyping to **RhD Gene (RHD) Copy Number**.



HOTLINE: Effective August 19, 2019

**2003176**

**Rufinamide, Serum or Plasma**

**RUFIN SP**

**Reference Interval:**

Effective August 19, 2019

Therapeutic Range	5-30 µg/mL
Dose-related range (values at dosages of 800-7200 mg/day)	3-30 µg/mL
Toxic	Not well established

**2010138**

***RUNX1-RUNX1T1 (AML1-ETO) t(8;21) Detection, Quantitative***

**AML1-ETO Q**

**Specimen Required:** Collect: Lavender (EDTA) or bone marrow (EDTA). Also acceptable: RNA extracted by CLIA certified lab.

Specimen Preparation: **Whole Blood:** Transport 5 mL whole blood. (Min: 1 mL)

**Bone Marrow:** Transport 3 mL bone marrow. (Min: 1 mL)

**Specimens must be received within 48 hours of collection due to lability of RNA.**

**Extracted RNA:** Transport 40 µL RNA with at least 40 ng/µL concentration. (Min: 40 µL) Transport RNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.

**Storage/Transport Temperature:** **Whole Blood or Bone Marrow:** CRITICAL REFRIGERATED. Separate specimens must be submitted when multiple tests are ordered.

**Extracted RNA:** CRITICAL FROZEN. Separate specimens must be submitted when multiple tests are ordered.

**Unacceptable Conditions** Serum, plasma, CSF, extracted DNA, RNA extracted by a non-CLIA lab, bone core, or FFPE tissue.

Specimens collected in anticoagulants other than EDTA. Severely hemolyzed or clotted specimens.

**Stability (collection to initiation of testing):** Ambient: 1 hour; Refrigerated: 48 hours; Frozen: Unacceptable

**Extracted RNA:** Ambient: Unacceptable; Refrigerate: Unacceptable; Frozen: Indefinitely

**0050527**

**Serine Proteinase 3 (PR3) Antibody**

**PR3**

**HOTLINE NOTE:** There is a clinically significant charting name change associated with this test.

Change the charting name for component 0050527, Serine Protease 3, IgG to Serine Proteinase 3, IgG.

**0050085**

**Smith (ENA) Antibody, IgG**

**SMITH**

**Interpretive Data:** Smith antibody is highly specific (greater than 90 percent) for systemic lupus erythematosus (SLE) but only occurs in 30-35 percent of SLE cases. The presence of antibodies to Smith has variable associations with SLE clinical manifestations.

**3000460**

**Smith and Smith/RNP (ENA) Antibodies, IgG**

**SMITH\_RNP**

**Reference Interval:**

Test Number	Components	Reference Interval	
0050470	Smith/RNP (ENA) Antibody, IgG	29 AU/mL or less	Negative
		30-40 AU/mL	Equivocal
		41 AU/mL or greater	Positive
0050085	Smith (ENA) Antibody, IgG	29 AU/mL or less	Negative
		30-40 AU/mL	Equivocal
		41 AU/mL or greater	Positive

**HOTLINE NOTE:** There is a clinically significant charting name change associated with this test.

Change the charting name for component 0050470 from Ribonucleic Protein (U1) (ENA) Ab, IgG to Smith/RNP (ENA) Ab, IgG.

**0050470**

**Smith/RNP (ENA) Antibody, IgG**

**RNP**

**Interpretive Data:** Smith/RNP antibodies are frequently seen in patients with mixed connective tissue disease (MCTD) and are also associated with other systemic autoimmune rheumatic diseases (SARDs) such as systemic lupus erythematosus (SLE), systemic sclerosis, and myositis. Antibodies targeting the Smith/RNP antigenic complex also recognize Smith antigens, therefore, the Smith antibody response must be considered when interpreting these results.

**Note:** Purified Smith/RNP antigen is used in this assay. U1RNP IgG antibodies detected using immunoassays that employ recombinant U1-70K, A, and C proteins may yield discrepant results.

**HOTLINE NOTE:** There is a clinically significant charting name change associated with this test.

Change the charting name for component 0050470 from Ribonucleic Protein (U1) (ENA) Ab, IgG to Smith/RNP (ENA) Ab, IgG.

HOTLINE: Effective August 19, 2019

**0060137**

**Stool Culture, *Yersinia***

**MC YERS**

**Specimen Required:** Collect: Stool.

Specimen Preparation: Preserve 5 mL stool in enteric transport media (Cary-Blair) immediately (ARUP supply #29799) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787. (Min: 1 mL)  
Storage/Transport Temperature: Refrigerated.

Unacceptable Conditions: Multiple specimens (more than one in 24 hours). Delayed transport without use of appropriate preservative.  
Stability (collection to initiation of testing): **Unpreserved:** Ambient: 2 hours; Refrigerated: Unacceptable; Frozen: Unacceptable  
**Preserved:** Ambient: 4 hours; Refrigerated: 72 hours; Frozen: Unacceptable

**0060705**

***Streptococcus* Group B by PCR**

**GBS PCR**

**Specimen Required:** Collect: Vaginal then rectal specimen with the same swab.

Specimen Preparation: Transfer swab to Amies media or Liquid Stuart media (ARUP supply #43875) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.  
Storage/Transport Temperature: Refrigerated.

Remarks: Specimen source required.

Unacceptable Conditions: Specimens in any transport media other than indicated above.

Stability (collection to initiation of testing): Ambient: 24 hours; Refrigerated: 6 days; Frozen: Unacceptable

**2008771**

**Supersaturation Profile, Urine**

**SUPERSAT**

**Performed:** Mon, Wed, Fri

**Reported:** 1-8 days

**Specimen Required:** Collect: 24-hour urine. Refrigerate during collection.

Specimen Preparation: Thoroughly mix entire collection (24-hour) in one container. Transport four separate 4 mL aliquots of urine using Calculi Risk/Supersaturation Urine Collection Kit (ARUP supply# 46007). Available online through eSupply using ARUP Connect™ or contact Client Services at (800) 522-2787. **Do not exceed 4 mL in tubes.**

Aliquot according to the following specifications:

1st aliquot (pH 2): Transfer 4 mL urine into a Sulfamic Acid Tube. (Min: 4 mL) Mix well. Freeze immediately.

2nd aliquot (pH 2): Transfer 4 mL urine into a Sulfamic Acid Tube. (Min: 4 mL) Mix well. Freeze immediately.

3rd aliquot (pH 9): Transfer 4 mL urine into a Sodium Carbonate Tube. (Min: 4 mL) Mix well. Freeze immediately.

4th aliquot: Transfer 4 mL urine into an Unpreserved Tube. (Min: 4 mL) Freeze immediately.

If collection kit is unavailable, transport four 4 mL unadjusted aliquots of urine.

Storage/Transport Temperature: **CRITICAL FROZEN. Separate specimens must be submitted when multiple tests are ordered. Ship patient and control specimens together.**

Remarks: Record total volume and collection time interval on tube and test request form.

Stability (collection to initiation of testing): Ambient: Unacceptable; Refrigerated: Unacceptable; Frozen: 2 weeks

**0093199**

**T-Cell Clonality by Flow Cytometry Analysis of TCR V-Beta**

**TCR V-BETA**

**Specimen Required:** Collect: Green (sodium heparin).

Specimen Preparation: Transport 5 mL whole blood.

Storage/Transport Temperature: Room temperature. Specimen should be received within 24 hours of collection for optimal viability.

Remarks: Previous leukemia/lymphoma phenotyping is required. If prior studies were not performed at ARUP, the outside flow cytometry report and histograms must accompany the specimen. If outside reports and histograms are not provided, a

**Leukemia/Lymphoma Phenotyping Evaluation by Flow Cytometry (ARUP test code 3001780)** will be added, at an additional cost. In addition, please provide CBC, Wright's stained smear (if available), clinical history, differential diagnosis, and any relevant pathology reports.

Stability (collection to initiation of testing): Ambient: 48 hours; Refrigerated: 48 hours; Frozen: Unacceptable

HOTLINE: Effective August 19, 2019

0055567

**T-Cell Clonality Screening by PCR**

**T CELL-F**

**Specimen Required:** Collect: Lavender (EDTA), bone marrow (EDTA), or tissue. **Also acceptable:** DNA extracted by CLIA certified lab.

Specimen Preparation: **Whole Blood:** Do not freeze. Transport 5 mL. (Min: 1 mL)

**Bone marrow:** Do not freeze. Transport 3 mL. (Min: 1 mL)

**Fresh Tissue:** Freeze immediately. Transport 100 mg or 0.5-2.0 cm<sup>3</sup> tissue

**FFPE Tumor Tissue:** Formalin fix (10 percent neutral buffered formalin) and paraffin embed tissue. Protect from excessive heat.

Tissue block will be returned after testing. Transport tissue block or four 10-micron shavings in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.

**Extracted DNA:** Transport 40uL DNA with at least 50 ng/uL concentration. Transport DNA in a tissue transport kit (ARUP Supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787.

Storage/Transport Temperature: **Whole Blood, Bone Marrow or Extracted DNA:** Refrigerated.

**Fresh Tissue:** Frozen on dry ice.

**FFPE Tumor Tissue:** Room temperature. **Also acceptable:** Refrigerated. Ship in cooled container during summer months.

Unacceptable Conditions: Plasma, serum, frozen tissue, DNA extracted by a non-CLIA lab. Specimens collected in anticoagulants other than EDTA or sodium heparin. Clotted or grossly hemolyzed specimens. Specimens fixed/processed in alternative fixatives or heavy metal fixatives (B-4 or B-5) or tissue sections on slides. Decalcified specimens.

Stability (collection to initiation of testing): **Whole Blood or Bone Marrow:** Ambient: 24 hours; Refrigerated: 5 days; Frozen: Unacceptable

**Fresh Tissue:** Ambient: Unacceptable; Refrigerated: 2 hours; Frozen: 1 year

**FFPE Tumor Tissue:** Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable

**Extracted DNA:** Ambient: 1 month; Refrigerate: Indefinitely; Frozen: Indefinitely

2013484

**TP53 Somatic Mutation, Prognostic**

**P53 MUTAT**

**Performed:** Varies

**Reported:** 3-11 days

**New Test**

3001704

**Treponema Pallidum by IHC**

**TPALL IHC**

Available Now

[Click for Pricing](#)

**Methodology:** Immunohistochemistry

**Performed:** Mon-Fri

**Reported:** 1-3 days

**Specimen Required:** Collect: Tissue.

Specimen Preparation: Formalin fix (10 percent neutral buffered formalin) and paraffin embed specimen (cells must be prepared into a cellblock). Protect paraffin block and/or slides from excessive heat. Transport tissue block or 5 unstained (3- to 5-micron thick sections), positively charged slides in a tissue transport kit (recommended but not required), (ARUP supply #47808) available online through eSupply using ARUP Connect or contact ARUP Client Services at (800) 522-2787. (Min: 2 slides) If sending precut slides, do not oven bake.

Storage/Transport Temperature: Room temperature. Also acceptable: Refrigerated. Ship in cooled container during summer months.

Unacceptable Conditions: Specimens submitted with non-representative tissue type. Depleted specimens.

Stability (collection to initiation of testing): Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable

**Interpretive Data:** See Compliance Statement A: [www.aruplab.com/CS](http://www.aruplab.com/CS)

**Note:** All stains will be handled as "Stain and Return" unless a consultation is requested. To request a consultation, submit the pathology report, all associated case materials (clinical history, blocks, slides, etc.), and the Anatomic Pathology requisition form (#32960) in place of the Immunohistochemistry Stain Form.

**CPT Code(s):** 88342

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

HOTLINE: Effective August 19, 2019

---

<b>New Test</b>	<b><u>3001831</u></b>	<b>Troponin T (cTnT) 5<sup>th</sup> Generation</b>	<b>CTNT</b>
-----------------	-----------------------	--	-------------

---

[Click for Pricing](#)

**Methodology:** Quantitative Electrochemiluminescent Immunoassay (ECLIA)  
**Performed:** Mon, Wed, Fri  
**Reported:** 1-4 days

**Specimen Required:** Collect: Plasma Separator Tube (PST), or Green (Lithium Heparin).  
Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 1 mL plasma to an ARUP Standard Transport Tube. (Min: 0.5 mL)  
Storage/Transport Temperature: **CRITICAL FROZEN. Separate specimens must be submitted when multiple tests are ordered.**  
Unacceptable Conditions: Specimens collected in potassium oxalate, sodium fluoride, or sodium citrate. Grossly hemolyzed specimens.  
Stability (collection to initiation of testing): Ambient: Unacceptable; Refrigerated: 24 hours; Frozen: 1 year

**Reference Interval:**

Female	Less than or equal to 10 ng/L
Male	Less than or equal to 15 ng/L

**CPT Code(s):** 84484

New York DOH Approved.

**HOTLINE NOTE:** Refer to the Test Mix Addendum for interface build information.

---

<b><u>0065031</u></b>	<b><i>Ureaplasma</i> Species and <i>Mycoplasma hominis</i> Culture</b>	<b>V UREA</b>
-----------------------	--	---------------

---

**Specimen Required:** Collect: Body fluid, **CSF, semen**, cervical or urethral swab, tissue, **or urine**. Also acceptable: Respiratory specimens from patients younger than 1 year of age.  
Specimen Preparation: Place swab or 0.5 mL of fluid (Min: 0.3 mL). in *Mycoplasma/Ureaplasma* transport media (UTM) (ARUP supply #12884) immediately. Available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787. Also acceptable: Any transport media validated for *Mycoplasma/Ureaplasma* transport such as M4 (DO NOT USE M4 RT).  
Storage/Transport Temperature: Frozen. Transport specimen on dry ice.  
Remarks: Specimen source preferred.  
Unacceptable Conditions: **Non-patient specimens**. Specimens not in *Mycoplasma/Ureaplasma* transport media. M4 RT or bacterial transport media. Dry swabs.  
Stability (collection to initiation of testing): Ambient: 8 hours; Refrigerated: 48 hours; **Frozen at -20°C: Unacceptable**; Frozen at -70°C: 1 month

---

<b><u>0020056</u></b>	<b>Viscosity, Serum</b>	<b>VIS-S</b>
-----------------------	-------------------------	--------------

---

**Reference Interval:**  
 Effective August 19, 2019  
 1.10-1.80 cP

---

<b><u>2013942</u></b>	<b>Zika Virus IgM Antibody Capture (MAC) by ELISA</b>	<b>ZIKA M</b>
-----------------------	---	---------------

---

**Interpretive Data:** This assay is intended for in vitro diagnostic use under FDA Emergency Use Authorization (EUA) and has not been FDA cleared or approved. In compliance with this authorization, please visit <https://aruplab.com/zika> for more information and to access the applicable information sheets.

The possibility of false-positive or false-negative results must be considered. RT-PCR testing on both a serum and urine specimen is recommended by the Centers for Disease Control and Prevention (CDC) to rule out false-negative IgM results in patients experiencing symptoms for less than 2 weeks. Specimens collected for IgM testing greater than or equal to 2 weeks after symptom onset do not require any additional testing. For more information, please review the current clinical guidelines for Zika virus testing at: [www.cdc.gov/zika/](http://www.cdc.gov/zika/).

**Note:** If the result is "Presumptive Zika," then Zika IgM Ab Capture (MAC) Confirmation (ARUP test code 3001904) will be added at no additional charge.

**HOTLINE NOTE:** There is a reflexive pattern change associated with this test.  
 Add reflex to 3001904, Zika IgM Ab Capture (MAC) Confirmation

HOTLINE: Effective August 19, 2019

The following will be discontinued from ARUP's test menu on August 19, 2019.  
Replacement test options are supplied if applicable.

Test Number	Test Name	Refer To Replacement
<a href="#">0098422</a>	Allergen, Occupational, Phthalic Anhydride IgE	
<a href="#">2006038</a>	Allergens, Respiratory Panel, Region 10, Southwestern Grasslands (OK, TX) IgE	Allergen, Region 10 Respiratory Panel IgE ( <a href="#">3001720</a> )
<a href="#">2006039</a>	Allergens, Respiratory Panel, Region 11, Rocky Mountain (AZ, ID, NM, WY CO, MT, UT) IgE	Allergen, Region 11 Respiratory Panel IgE ( <a href="#">3001721</a> )
<a href="#">2006040</a>	Allergens, Respiratory Panel, Region 12, Arid Southwest (S. AZ, S.E. CA) IgE	Allergen, Region 12 Respiratory Panel IgE ( <a href="#">3001722</a> )
<a href="#">2006041</a>	Allergens, Respiratory Panel, Region 13, Southern Coastal (CA) IgE	Allergen, Region 13 Respiratory Panel IgE ( <a href="#">3001723</a> )
<a href="#">2006042</a>	Allergens, Respiratory Panel, Region 14, Central California (CA) IgE	Allergen, Region 14 Respiratory Panel IgE ( <a href="#">3001724</a> )
<a href="#">2006043</a>	Allergens, Respiratory Panel, Region 15, Intermountain West (NV, S. ID) IgE	Allergen, Region 15 Respiratory Panel IgE ( <a href="#">3001725</a> )
<a href="#">2006044</a>	Allergens, Respiratory Panel, Region 16, Inland Northwest (OR, Central and East WA) IgE	Allergen, Region16 Respiratory Panel IgE ( <a href="#">3001726</a> )
<a href="#">2006045</a>	Allergens, Respiratory Panel, Region 17, Pacific Northwest (NW CA, W. OR, WA) IgE	Allergen, Region17 Respiratory Panel IgE ( <a href="#">3001727</a> )
<a href="#">2006046</a>	Allergens, Respiratory Panel, Region 18, Alaska IgE	Allergen, Region 18 Respiratory Panel IgE ( <a href="#">3001728</a> )
<a href="#">2006047</a>	Allergens, Respiratory Panel, Region 19, Puerto Rico IgE	Allergen, Region 19 Respiratory Panel IgE ( <a href="#">3001729</a> )
<a href="#">2005718</a>	Allergens, Respiratory Panel, Region 2, Mid-Atlantic (DE, MD, VA, DC, NC) IgE	Allergen, Region 2 Respiratory Panel IgE ( <a href="#">3001712</a> )
<a href="#">2006048</a>	Allergens, Respiratory Panel, Region 20, Hawaii IgE	Allergen, Region 20 Respiratory Panel IgE ( <a href="#">3001730</a> )
<a href="#">2006025</a>	Allergens, Respiratory Panel, Region 3, South Atlantic (GA, SC, N. FL) IgE	Allergen, Region 3 Respiratory Panel IgE ( <a href="#">3001713</a> )
<a href="#">2006026</a>	Allergens, Respiratory Panel, Region 4, Subtropic Florida (S. of Orlando) IgE	Allergen, Region 4 Respiratory Panel IgE ( <a href="#">3001714</a> )
<a href="#">2006031</a>	Allergens, Respiratory Panel, Region 5, Ohio Valley (IN, OH, TN, WV, KY) IgE	Allergen, Region 5 Respiratory Panel IgE ( <a href="#">3001715</a> )
<a href="#">2006032</a>	Allergens, Respiratory Panel, Region 6, South Central (AL, AR, LA, MS) IgE	Allergen, Region 6 Respiratory Panel IgE ( <a href="#">3001716</a> )
<a href="#">2006033</a>	Allergens, Respiratory Panel, Region 7, Northern Midwest (MI, WI, MN) IgE	Allergen, Region 7 Respiratory Panel IgE ( <a href="#">3001717</a> )
<a href="#">2006034</a>	Allergens, Respiratory Panel, Region 8, Central Midwest (IL, MO, IA) IgE	Allergen, Region 8 Respiratory Panel IgE ( <a href="#">3001718</a> )
<a href="#">2006037</a>	Allergens, Respiratory Panel, Region 9, Great Plains (KS, NE, ND, SD) IgE	Allergen, Region 9 Respiratory Panel IgE ( <a href="#">3001719</a> )
<a href="#">0091195</a>	Amantadine Quantitative, Serum or Plasma	
<a href="#">0091165</a>	Barium Quantitative, Urine	
<a href="#">0051421</a>	Beta Globin ( <i>HBB</i> ) HbS, HbC, and HbE Mutations	Beta Globin ( <i>HBB</i> ) Gene Sequencing ( <a href="#">0050578</a> )
<a href="#">0051422</a>	Beta Globin ( <i>HBB</i> ) HbS, HbC, and HbE Mutations, Fetal	Beta Globin ( <i>HBB</i> ) Sequencing, Fetal ( <a href="#">0050388</a> )
<a href="#">2014493</a>	Bupivacaine Quantitative, Serum or Plasma	
<a href="#">2014027</a>	Calcium, RBC	
<a href="#">2013991</a>	Dermatomyositis Panel	Dermatomyositis Autoantibody Panel ( <a href="#">3001782</a> )
<a href="#">0091219</a>	Diphenhydramine Quantitative, Urine	
<a href="#">0049050</a>	Esterase Stain, Nonspecific	
<a href="#">2014674</a>	Expanded Carrier Screen, Genotyping	Expanded Carrier Screen by Next Generation Sequencing ( <a href="#">2014680</a> )
<a href="#">2014671</a>	Expanded Carrier Screen, Genotyping with Fragile X	Expanded Carrier Screen by Next Generation Sequencing with Fragile X ( <a href="#">2014677</a> )
<a href="#">0091263</a>	Furosemide Quantitative, Serum or Plasma	
<a href="#">0051067</a>	HLA DRB 3*,4*,5*	
<a href="#">2008788</a>	Influenza A Virus H1/H3 Subtype by PCR with Reflex to H1N1 (2009) Oseltamivir Resistance by Sequencing	
<a href="#">0093148</a>	Interferon-Alpha by ELISA, Serum	
<a href="#">2013993</a>	Interstitial Lung Disease Panel	Interstitial Lung Disease Autoantibody Panel ( <a href="#">3001784</a> )
<a href="#">2008003</a>	Leukemia/Lymphoma Phenotyping by Flow Cytometry	Leukemia/Lymphoma Phenotyping Evaluation by Flow Cytometry ( <a href="#">3001780</a> )
<a href="#">0091200</a>	LSD, Serum or Plasma - Screen with Reflex to Confirmation/Quantitation	
<a href="#">0091272</a>	Mercury, Hair	
<a href="#">2013961</a>	Myositis Extended Panel	Extended Myositis Panel ( <a href="#">3001781</a> )
<a href="#">0092140</a>	Nitrogen, Total, Urine	
<a href="#">2002016</a>	Pancreatitis ( <i>PRSSI</i> ) Sequencing	Pancreatitis ( <i>PRSSI</i> ) Deletion/Duplication ( <a href="#">3001760</a> ), Pancreatitis ( <i>SPINK1</i> ) Deletion/Duplication ( <a href="#">3001764</a> ), and Pancreatitis ( <i>PRSSI</i> ) Sequencing and Deletion/Duplication ( <a href="#">3001768</a> )
<a href="#">2013992</a>	Polymyositis and Dermatomyositis Panel	Dermatomyositis and Polymyositis Panel ( <a href="#">3001783</a> )
<a href="#">0091568</a>	Sulfhemoglobin Quantitative, Whole Blood	
<a href="#">0098803</a>	Troponin T	Troponin T (cTnT) 5th Generation ( <a href="#">3001831</a> )