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Additional ordering and billing information

Test Number	Mnemonic	Test Name	New Test	Test Name Change	Specimen Requirements	Methodology	Performed/Reported	Note	Interpretive Data	Reference Interval	Component Charting Name	Component Change	Reflex Pattern	Result Type	Ask at Order Prompt	Numeric Map	Unit of Measure	CPT Code	Pricing Change	Inactivation w/ Replacement	Inactivation w/o Replacement
0020009	GT	Gamma Glutamyl Transferase, Serum or Plasma			x																
0020012	ALDOLASE	Aldolase, Serum			x																
0020045	LA	Lactic Acid, Plasma (Change effective as of 07/21/25: Refer to 3019650 in the July Hotline)																		x	
0020053	CH HDL	HDL Cholesterol			х	х															
0020096	COPPER	Copper, Serum or Plasma			x				x												
0020097	ZINC	Zinc, Serum or Plasma			x				x												
0020098	LEAD-WB	Lead, Whole Blood (Venous)		x					x	x	x										
0020245	BICARB	Bicarbonate (HCO[3]), Urine (Change effective as of 07/21/25: Refer to 3019581 in the July Hotline)																		x	
0020257	LDL Direct	LDL Cholesterol, Direct			х	х			x												
0020468	CRISK E	Lipid Panel, Extended			х																
0020504	Lactic Acid, BF	Lactic Acid, Body Fluid (Inactive as of 07/21/25)																			x
0020516	LA-CF	Lactic Acid, CSF(Change effective as of 07/21/25: Refer to 3019672 in the July Hotline)																		x	



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0020544	ACS	Acid Phosphatase, Total, Serum			x																
0020584	HY MET B4	Heavy Metals Panel 4, Whole Blood		x						x	x										
0020614	ZPP IND	Zinc Protoporphyrin (ZPP), Whole Blood Industrial									x										
0020745	LEAD CAP	Lead, Whole Blood (Capillary)		x	x				x	x	x										
0020852	CITRIC U	Citric Acid, Urine			х																
0025013	CD EXP	Cadmium Exposure Panel - OSHA							x	x	x										
0025016	LEAD-IND	Lead, Industrial, Whole Blood		x					х	х	x										
0025023	SE S	Selenium, Serum or Plasma			x				x												
0025037	COBALT S	Cobalt, Serum or Plasma			х				х												
0050070	AMB	Entamoeba histolytica (amebiasis), Antibody, IgG			x	x	x														
0050165	CMV IGG	Cytomegalovirus Antibody, IgG			x					x											
0050225	EBV EAD	Epstein-Barr Virus Antibody to Early D Antigen (EA-D), IgG							x	x											
0050235	EBV G	Epstein-Barr Virus Antibody to Viral Capsid Antigen, IgG							x	x											
0050240	EBV M	Epstein-Barr Virus Antibody to Viral Capsid Antigen, IgM							x	x											

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0050245	EBV NA	Epstein-Barr Virus Antibody to Nuclear Antigen, IgG							x	x											
0050280	НАРТО	Haptoglobin			x																
0050293	HERPES	Herpes Simplex Virus Type 1 and/or 2 Antibodies, IgG			x					x											
0050379	HERPICSF	Herpes Simplex Virus Type 1 Glycoprotein G- Specific Antibody, IgG by ELISA, CSF			x																
0050392	HLAB27 PCR	Ankylosing Spondylitis (HLA-B27) Genotyping (Change effective as of 7/21/2025: Refer to 3019466 in the July Hotline)																		x	
0050394	HER1/2CSF	Herpes Simplex Virus Type 1 and/or 2 Antibodies, IgG, CSF			x																
0050521	TOXO PAN	Toxoplasma gondii Antibodies, IgG and IgM			x				x	x											
0050551	RUBEIGM	Rubella Antibody, IgM			х																
0050552	RUBE G/M	Rubella Antibodies, IgG and IgM			x				x	x											
0050553	CMV IGM	Cytomegalovirus Antibody, IgM			x																
0050557	TOXEIGM	Toxoplasma gondii Antibody, IgM			x																



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0050564	SC PAN	Saccharomyces cerevisiae Antibodies, IgG & IgA					x														
0050596	PCA	Gastric Parietal Cell Antibody, IgG					x														
0050600	EBV PAN	Epstein-Barr Virus Antibody Panel I								x											
0050602	EBV PAN 2	Epstein-Barr Virus Antibody Panel II								x											
0050622	CMV PAN	Cytomegalovirus Antibodies, IgG and IgM			x				x	x											
0050736	EMAR TITER	Endomysial Antibody, IgA by IFA			x																
0050770	TOXEIGG	Toxoplasma gondii Antibody, IgG			x					x											
0050771	RUBEIGG	Rubella Antibody, IgG			х					х											
0050772	TORCH IGG	TORCH Antibodies, IgG			х				x	х											
0050777	MHA	Treponema pallidum Antibody by TP-PA					x														
0050779	DTH	Diphtheria, Tetanus, and H. Influenzae b Antibodies, IgG			x																
0050791	SSA/SSB	Extractable Nuclear Antigen Antibodies (SSA 52, SSA 60, and SSB)			x																
0050905	PHOS AB	Phosphatidylserine Antibodies, IgG, IgM, and IgA					x														



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0051205	MCADPCR	Medium Chain Acyl-CoA Dehydrogenase (ACADM) 2 Mutations (Change effective as of 07/21/25: Refer to 3019336 in the July Hotline)																		x	
0051244	*ASM TITER	Smooth Muscle Antibody, IgG Titer			x																
0051270	GALTDNA FE	Galactosemia (GALT) 9 Mutations, Fetal			x				x												
0051302	PROTHROM G	Prothrombin Antibody, IgG					x														
0051368	RHD	RhD Gene (RHD) Copy Number (Change effective as of 7/21/25: Refer to 3019342 in the July Hotline)																		x	
0051415	AJP	Ashkenazi Jewish Diseases, 16 Genes			x			x	x												
0051627	EBV PAN 3	Epstein-Barr Virus Antibody to Viral Capsid Antigen, IgG and IgA								x											
0051708	HERPR PAN2	Herpes Simplex Virus Type 1 and/or 2 Antibodies, IgG with Reflex to Type 1 and 2 Glycoprotein G-Specific Ab, IgG			x				x	x											
0055405	WH F IGG	Allergen, Insects and Venom, White-Faced Hornet IgG			x																



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0055410	YEL J IGG	Allergen, Insects and Venom, Yellow Jacket IgG			x																
0055415	PAP-W IGG	Allergen, Insects and Venom, Paper Wasp IgG			x																
0055420	YE F IGG	Allergen, Insects and Venom, Yellow Hornet IgG			x																
0055656	HFE PCR	Hemochromatosis (HFE) 3 Mutations (Change effective as of 07/21/25: Refer to 3019007 in the July Hotline)																		x	
0060217	MA AFB	Antimicrobial Susceptibility, AFB/Mycobacteria			x				x	x											
0070189	BILE AC	Bile Acids, Total			х																
0080315	Phenylal Qnt P	Phenylalanine Monitoring, Plasma (Change effective as of 07/21/25: Refer to 0080336)																		x	
0080355	Tyrosine Qnt P	Tyrosine, Plasma (Change effective as of 07/21/25: Refer to 0080336)																		x	
0080525	VIT A	Vitamin A (Retinol), Serum or Plasma			x																
0081105	CYS PAN	Cystinuria Panel (Change effective as of 07/21/25: Refer to 2009419)																		x	



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0090284	Almond IgG	Allergen, Food, Almond IgG			x																
0090286	Banana IgG	Allergen, Food, Banana IgG			x																
0090287	Garlic IgG	Allergen, Food, Garlic IgG			x																
0090289	Gluten IgG	Allergen, Food, Gluten IgG			x																
0090291	Whey IgG	Allergen, Food, Whey IgG			x																
0093454	Fus M IgG	Allergen, Fungi and Molds, Fusarium proliferatum/moniliform e IgG			x																
0097299	Mushroom IgG	Allergen, Food, Mushroom IgG			x																
0097302	Coffee IgG	Allergen, Food, Coffee IgG			x																
0097304	C albicans G	Allergen, Fungi and Molds, Candida albicans IgG			x																
0097305	Aureo Pu IgG	Allergen, Fungi and Molds, Aureobasidium pullulans IgG			x																
0097306	Onion IgG	Allergen, Food, Onion IgG			x																
0097307	Rhizopus IgG	Allergen, Fungi and Molds, Rhizopus nigricans IgG			x																
0097308	Stemph B IgG	Allergen, Stemphylium herbarum/botryosum, IgG			x																



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0097309	Phoma b IgG	Allergen, Fungi and Molds, Phoma betae IgG			x																
0097310	Penicil n G	Allergen, Fungi and Molds, Penicillium chrysogenum/notatum IgG			x																
0097313	Helmin IgG	Allergen, Fungi and Molds, Helminthosporium halodes/Setomelanom ma rostrata IgG			x																
0097314	Cladosporlg G	Allergen, Fungi and Molds, Cladosporium IgG			x																
0097315	Egg Yolk IgG	Allergen, Food, Egg Yolk IgG			x																
0097316	Mucor IgG	Allergen, Fungi and Molds, Mucor racemosus IgG			x																
0097323	Rice IgG	Allergen, Food, Rice IgG			x																
0097636	Wheat IgG	Allergen, Food, Wheat IgG			x																
0097641	Potato IgG	Allergen, Food, Potato (White) IgG			x																
0097642	Rye lgG	Allergen, Food, Rye IgG			х																
0097643	Soybean IgG	Allergen, Food, Soybean IgG			x																
0097644	Tomato IgG	Allergen, Tomato IgG			x																
0097647	Orange IgG	Allergen, Food, Orange IgG			x																



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0097648	Peanut IgG	Allergen, Food, Peanut IgG			x																
0097649	Pork IgG	Allergen, Food, Pork IgG			х																
0097651	Lettuce IgG	Allergen, Food, Lettuce IgG			x																
0097652	Malt IgG	Allergen, Food, Malt IgG			х																
0097653	Casein IgG	Allergen, Food, Casein (Cow's Milk) IgG			x																
0097654	Oat IgG	Allergen, Food, Oat IgG			х																
0097656	Chicken IgG	Allergen, Food, Chicken IgG			x																
0097657	Choco IgG	Allergen, Food, Chocolate IgG			x																
0097658	Corn IgG	Allergen, Food, Corn IgG			х																
0097659	Egg White G	Allergen, Food, Egg White IgG			x																
0097706	Yeast IgG	Allergen, Food, Baker's Yeast IgG			x																
0097707	Barley IgG	Allergen, Food, Barley IgG			x																
0097708	Beef IgG	Allergen, Food, Beef IgG			х																
0097773	Alt Tenuis G	Allergen, Fungi and Molds, Alternaria tenuis IgG			x																
0098627	KEPPRA	Keppra (Levetiracetam)				x										х					
0098830	CR S	Chromium, Serum			x				x												
0099045	Arsenic Blood	Arsenic, Whole Blood		x					x		x										



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0099231	COBALT B	Cobalt, Whole Blood		х					x	x	х										
0099249	RIBPP	Ribosomal P Protein Antibody			x																
0099266	AL S	Aluminum, Serum			х																
0099272	MANG WB	Manganese, Whole Blood							x		x										
0099305	Mercury Blood	Mercury, Whole Blood							x		x										
0099452	NICKEL	Nickel, Serum			x				x												
0099470	HY MET B	Heavy Metals Panel 3, Whole Blood		х						x	x										
0099478	BS B	Bismuth, Whole Blood		х					x												
0099592	JO 1 IgG	Jo-1 Antibody, IgG			x																
0099610	THALB	Thallium, Whole Blood							x		х										
0099675	Cadmium Bld	Cadmium, Whole Blood		x					x		x										
2000136	TH REQUEST	Cytology, ThinPrep Pap Test and Human Papillomavirus (HPV) High Risk Screen by Transcription-Mediated Amplification (TMA), With Reflex to Genotypes 16 and 18/45 (Change effective as of 07/21/25: Refer to 3018973 in the July Hotline)																		x	



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2000137	GT REQUEST	Cytology, ThinPrep Pap Test (Change effective as of 07/21/25: Refer to 3018968 in the July Hotline)																		x	
2000138	TR REQUEST	Cytology, ThinPrep Pap Test With Reflex to Human Papillomavirus (HPV), High Risk Screen by Transcription- Mediated Amplification (TMA), With Reflex to Genotypes 16 and 18/45 (Change effective as of 07/21/25: Refer to 3018971 in the July Hotline)																		x	
2001763	HIRSUTISM	Hirsutism Evaluation Panel			x																
2002282	CAH 11-B HYDROX	Congenital Adrenal Hyperplasia Panel, 11- Beta Hydroxylase Deficiency			x																
2003248	DEXA TMS	Dexamethasone, Serum or Plasma by LC-MS/MS			x																
2004221	NMDA IGG	N-methyl-D-Aspartate Receptor (NMDAR) Antibody, IgG by CBA- IFA, Serum With Reflex to Titer			x																
2005248	BILCSF	Bilirubin, CSF														х					
2005287	CHROMATI N	Chromatin Antibody, IgG					x														



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nactivation w/o Replacement Inactivation w/ Replacement **Component Charting Name Specimen Requirements** Performed/Reported **Test Name Change Component Change** Ask at Order Prompt **Reference Interval** Interpretive Data Unit of Measure **Pricing Change Reflex Pattern** Numeric Map Methodology **Result Type** New Test CPT Code Note Test Mnemonic Test Name Number 2005405 METREXSN Methotrexate, Sensitive (Change effective as of 07/21/25: Refer to Х 3019648 in the July Hotline) 2005779 PNEUMO 23 Streptococcus pneumoniae Antibodies, х IgG (23 Serotypes) 2007192 ADENO QNT Adenovirus, Quantitative Х Х PCR 2007211 PEANUT Allergen, Food, Peanut х COM Components IgE 2007213 G FOOD PAN Allergens, Food, Х Extended Panel IgG 2007214 IGG MEATS Allergens, Food, Meat Х Panel IgG 2007215 G FOOD Allergens, Food, Х COM Common Panel IgG 2007216 **IGG FOOD** Allergens, Food, IgG Х Panel 2007220 ECHINO IGG Echinococcus Antibody, х lqG 2007463 IODINESER Iodine, Serum Х 2007601 ANTI-C1Q Anti-C1q Antibody, IgG х 2009447 APS/PT G Phosphatidylserine and Prothrombin Antibody, х lgG 2009451 APS/PT PAN Phosphatidylserine and Prothrombin Antibodies, х IgG and IgM



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2011418	MC CRO	Carbapenem-Resistant Organism Culture			x																
2011723	AVOCADOIG G	Allergen, Food, Avocado IgG			х																
2011725	BROCC IGG	Allergen, Food, Broccoli IgG			x																
2011727	CASHEW IGG	Allergen, Food, Cashew IgG			x																
2011729	CHSMLD IGG	Allergen, Food, Cheese Mold IgG			x																
2011731	CLAM IGG	Allergen, Food, Clam IgG			x																
2011733	COCONUTIG G	Allergen, Food, Coconut IgG			x																
2011735	CRAB IGG	Allergen, Food, Crab IgG			x																
2011737	LOBSTERIG G	Allergen, Food, Lobster IgG			x																
2011739	OYSTER IGG	Allergen, Food, Oyster IgG			x																
2011741	PNAPPL IGG	Allergen, Food, Pineapple IgG			x																
2011743	SCALLOPIG G	Allergen, Food, Scallop IgG			x																
2011745	SHRIMP IGG	Allergen, Food, Shrimp IgG			x																
2011747	STRWBRYIG G	Allergen, Food, Strawberry IgG			x																
2011749	TUNA IGG	Allergen, Food, Tuna IgG			х																
2011751	TURKEY IGG	Allergen, Food, Turkey IgG			x																



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2011753	WALNUT IGG	Allergen, Food, Walnut IgG			x																
2011808	CHIKG	Chikungunya Antibody, IgG			x																
2011810	СНІКМ	Chikungunya Antibody, IgM			x																
2011812	CHIKPAN	Chikungunya Antibodies, IgG and IgM			x																
2011815	OLIVES IGG	Allergen, Food, Olives IgG			x																
2011817	CHEDCHEES E	Allergen, Food, Cheddar Cheese IgG			x																
2011819	WHOLE EGG	Allergen, Food, Whole Egg, IgG			x																
2012166	DPYD	Dihydropyrimidine Dehydrogenase (DPYD)										x									
3000143	MS QUAD	Maternal Serum Screen, Alpha Fetoprotein, hCG, Estriol, and Inhibin A (Quad)										x									
3000144	MS AFP	Maternal Serum Screen, Alpha Fetoprotein										x									
3000145	MS FTS	Maternal Serum Screen, First Trimester, hCG, PAPP-A, NT										x									
3000146	MS SEQ1	Maternal Screening, Sequential, Specimen #1, hCG, PAPP-A, NT										x									



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3000147	MS INT1	Maternal Serum Screening, Integrated, Specimen #1, PAPP-A, NT										x									
3000462	IMM PLT	Immature PLT Fraction (Inactive as of 07/21/25)																			x
3000876	ASPERF IGG	Aspergillus fumigatus Antibody IgG			x																
3000894	HHACASCA DE	Hereditary Hemolytic Anemia Cascade																x			
3001518	3A4/3A5	CYP3A4 and CYP3A5										х									
3001524	CYP PANEL	Cytochrome P450 Genotyping Panel										x									
3001831	CTNT	Troponin T (cTnT) 5th Generation					x														
3002477	GP210 AB	Anti-gp210 Antibody, IgG					x														
3002478	SP100 AB	Anti-sp100 Antibody, IgG					х														
3002482	SP100GP21 0	Anti-sp100 and anti- gp210 Antibodies, IgG					x														
3002598	PETH	Phosphatidylethanol (PEth), Whole Blood, Quantitative			x																
3003043	NIPT NGSAN	Non-Invasive Prenatal Aneuploidy Screen by cell-free DNA Sequencing			x																
3003748	IBD-PAN	Inflammatory Bowel Disease Differentiation Panel					x														



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3003924	ALPHAGALP N	Allergen, Food, Alpha- Gal (galactose-alpha-1,3- galatose) Panel			x																
3003992	CARP IGG	Carbamylated Protein (CarP) Antibody, IgG					x														
3004255	CYP GD	Cytochrome P450 Genotyping Panel, with GeneDose Access										x									
3004273	CMAPFFPE	Cytogenomic Molecular Inversion Probe Array FFPE Tissue - Products of Conception										x									
3004275	FFPEARRAY	Cytogenomic Molecular Inversion Probe Array FFPE Tissue - Oncology										x									
3004471	PGX PSYCH	Pharmacogenetics Panel: Psychotropics										x									
3004833	COMPDRUG SP	Drug Profile, Expanded Targeted Panel by LC- MS/MS, Serum/Plasma					x														
3005060	Compdrug Ur	Drug Profile, Expanded Targeted Panel by LC- MS/MS, Urine					x														
3005425	STRPOST- MO	Chimerism, Posttransplant, Sorted Cells (Monocytes) (Inactive as of 7/21/25)																			x
3006066	TOXOCARA G	Toxocara Antibodies, IgG by ELISA				x	x		x												
3006366	PGXPSYC GD	Pharmacogenetics Panel: Psychotropics, with GeneDose Access										х									



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3016636	HPV PRMRY	HPV Primary Screen by PCR With Reflex to Cytology					x	x	x									x			
3016767	ANTI-PLA2R	Anti-Phospholipase A2 Receptor (PLA2R) Antibody, IgG by ELISA					x														
3017156	THROMRISK	Thrombotic Risk Reflex Panel			x																
3017747	HERPR CSF	Herpes Simplex Virus Type 1 and/or 2 Antibodies, IgG (CSF) With Reflex to Type 1 and 2 Glycoprotein G- Specific Ab, IgG			x																
3017866	DPYDUGT1A 1	Dihydropyrimidine Dehydrogenase (DPYD) and UPD Glucuronosyltransferase 1A1 (UGT1A1) Genotyping (Change effective as of 07/21/25: Refer to 3019841 in the July Hotline)																		x	
3017902	SC5B-9	SC5b-9			x																
3018799	TNUT PAN R	Allergen, Food, Tree Nuts With Reflex to Components, IgE			x																
3018866	COMBI PAN2	Dermatomyositis and Polymyositis Panel			x																
3018867	MYOS EXT2	Extended Myositis Panel			х																
3018868	POLY MY02	Polymyositis Panel			х																



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Test Number	Mnemonic	Test Name	New Test	Test Name Change	Specimen Requirements	Methodology	Performed/Reported	Note	Interpretive Data	Reference Interval	Component Charting Name	Component Change	Reflex Pattern	Result Type	Ask at Order Prompt	Numeric Map	Unit of Measure	CPT Code	Pricing Change	Inactivation w/ Replacement	Inactivation w/o Replacement
3018869	ILD PANEL2	Interstitial Lung Disease Autoantibody Panel			x																
3018870	DERM PAN2	Dermatomyositis Autoantibody Panel			x																
3018968	TP REQUEST	ThinPrep PAP Test (Standalone)	х																		
3018971	TA REQUEST	ThinPrep PAP Test With Reflex to HPV if ASCUS	x																		
3018973	TM REQUEST	ThinPrep PAP Test With Co-Test HPV	x																		
3019007	HFEPCR	Hemochromatosis (HFE) 3 Variants	x																		
3019269	MEASLESPC R	Measles Virus by Qualitative NAAT	x																		
3019336	MCAD_PCR	Medium Chain Acyl-CoA Dehydrogenase (ACADM) 2 Variants	x																		
3019342	RHD PCR	RhD Gene (RHD) Copy Number by PCR	x																		
3019353	ICOS_IHC	ICOS by Immunohistochemistry	x																		
3019466	HLA- B27PCR	Ankylosing Spondylitis (HLA-B27) Genotyping	x																		
3019471	THC CRT	THC Metabolite, with Ratio, Urine	x																		
3019538	H3K36MIHC	H3K36M by Immunohistochemistry	x																		
3019566	HBA FGA FE	Alpha Globin (HBA1 and HBA2) Sequencing and Deletion/Duplication, Fetal	x																		



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Additional ordering and billing information

Test Number	Mnemonic	Test Name	New Test	Test Name Change	Specimen Requirements	Methodology	Performed/Reported	Note	Interpretive Data	Reference Interval	Component Charting Name	Component Change	Reflex Pattern	Result Type	Ask at Order Prompt	Numeric Map	Unit of Measure	CPT Code	Pricing Change	Inactivation w/ Replacement	Inactivation w/o Replacement
3019581	BICARBON	Bicarbonate, Urine	х																		
3019583	BICARB BF	Bicarbonate, Body Fluid	х																		
3019585	MA CARBA5	Antimicrobial Susceptibility - Carbapenemase Detection by CARBA5	x																		
3019648	METHOTRE X	Methotrexate, Serum or Plasma	x																		
3019650	LACTATE P	Lactate, Plasma	х																		
3019652	TRBC1_IHC	TRBC1 by Immunohistochemistry	x																		
3019672	LACT CSF	Lactate, CSF	х																		
3019841	UGT1A1DPY D	UPD Glucuronosyltransferase 1A1 (UGT1A1) and Dihydropyrimidine Dehydrogenase (DPYD) Genotyping	x																		



Gamma Glutamyl Transferase, Serum or Plasma								
Specimen Bequirements:								
Patient Preparation:								
Collect:	Serum separator tube or plasma separator tube. Also acceptable: <u>Green (lithium heparin Pink (K2EDTA</u>).							
Specimen Preparation:	Allow serum tube to clot completely at room temperature. Separate serum or plasma from cells within 2 hours of collection. Transfer 1 mL serum or plasma to an ARUP Standard Transport Tube. (Min: 0.2 mL)							
Transport Temperature:	Refrigerated.							
Unacceptable Conditions:	Grossly hemolyzed specimens.							
Remarks:								
Stability:	After separation from cells: Ambient: 1 week; Refrigerated: 1 week; Frozen: 2 months							
Methodology:	Quantitative Enzymatic Assay							
Performed:	Sun-Sat							
Reported:	Within 24 hours							
Note:								
CPT Codes:	82977							
New York DOH Approval Status:	This test is New York DOH approved.							
Interpretive Data:								
Reference Interval:								
By report (reports may vary based	on instrumentation)							

Aldolase, Serum 0020012, ALDOLASE	
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube.
Specimen Preparation:	Allow specimen to clot completely at room temperature. Transfer 1 mL serum to an ARUP <u>standard transport</u> <u>tube.</u> Standard Transport Tube. (Min: 0.5 mL)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Specimen types other than serum. Hemolyzed specimens.
Remarks:	
Stability:	After separation from cells: Ambient: <u>24</u> 8 hours; Refrigerated: 5 days; Frozen: 6 months
Methodology:	Quantitative Enzymatic Assay
Performed:	Sun-Sat
Reported:	Within 24 hours
Note:	
CPT Codes:	82085
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	

Reference Interval:

0-30 days	6.0-32.0 U/L
1-5 months	3.0-12.0 U/L
6-35 months	3.5-10.0 U/L
3-6 years	2.7-8.8 U/L
7-17 years	3.3-9.7 U/L
18 years and older	1.2-7.6 U/L



HDL Cholesterol	
0020053, CH HDL	
Specimen Requirements:	
Patient Preparation:	
Collect:	Plasma separator tube or serum separator tube.
Specimen Preparation:	Allow specimen to clot completely at room temperature. Separate serum or plasma from cells ASAP or within 2 hours of collection. Transfer 1 mL serum or plasma to an ARUP <u>standard transport tube.</u> <u>Standard Transport Tube.</u>
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	
Remarks:	
Stability:	After separation from cells: Ambient: <u>48</u> 24 hours; Refrigerated: 1 week; Frozen: <u>3 months1 month</u>
Methodology:	Quantitative Detergent Solubilization/Enzymatic Assay
Performed:	Sun-Sat
Reported:	Within 24 hours
Note:	Assay interference (negative) may be observed when high concentrations of N-acetylcysteine (NAC) are present. Negative interference has also been reported with NAPQI (an acetaminophen metabolite), but only when concentrations are at or above those expected during acetaminophen overdose.
CPT Codes:	83718
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	
An HDL cholesterol less than 40 m An HDL cholesterol greater than 60	g/dL is low and constitutes a coronary heart disease risk factor. D mg/dL is a negative risk factor for coronary heart disease.
CHD Risk Factors	

- +1 Age: Men > 45
 - Women > 55 or premature menopause without estrogen therapy
- +1 Family history of premature CHD
- +1 Current smoking



- +1 Hypertension
- +1 Diabetes mellitus
- +1 Low HDL cholesterol: < 40 mg/dL
- -1 High HDL cholesterol: \geq 60 mg/dL

Reference Interval:

Desirable: 40-59 mg/dL



Copper, Serum or Plasma	
0020096, COPPER	
Specimen Requirements:	
Patient Preparation:	Diet, medication, and nutritional supplements may introduce interfering substances. Patients should be encouraged to discontinue nutritional supplements, vitamins, minerals, and <u>nonessential non-essential</u> over-the-counter medications (upon the advice of their physician).
Collect:	Royal blue (<u>no additive), royal</u> No <u>Additive), Royal</u> blue (K2EDTA), or <u>r</u> Royal blue (NaHep).
Specimen Preparation:	Separate from cells ASAP or within 2 hours of collection. Transfer 2 mL serum or plasma to an ARUP Trace Element-Free Transport Tube (ARUP supply #43116) available online through eSupply using ARUP Connect(TM) or contact ARUP Client Services at (800-)-522-2787- (Min: 0.5 mL). Do not use utensils (i.e., syringes, needles, or pipettes) in the collection or transfer of the sample, pour directly into transport tube.)
Transport Temperature:	Room temperature. Also acceptable: Refrigerated or frozen.
Unacceptable Conditions:	Specimens that are not separated from the red cells or clot within 2 hours. Specimens collected in containers other than specified. Specimens transported in containers other than specified.
Remarks:	
Stability:	Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Indefinitely
Methodology:	Quantitative Inductively Coupled Plasma-Mass Spectrometry
Performed:	Sun-Sat
Reported:	1-3 days
Note:	
CPT Codes:	82525
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	



Elevated results may be due to skin or collection-related contamination, including the use of a noncertified metal-free collection/transport tube. If contamination concerns exist due to elevated levels of serum/plasma copper, confirmation with a second specimen collected in a certified metal-free tube is recommended.

Serum copper may be elevated with infection, inflammation, stress, and copper supplementation. In females, elevated copper may also be caused by oral contraceptives and pregnancy (concentrations may be elevated up to 3 times normal during the third trimester).

Serum copper may be reduced by use of corticosteroids and zinc and by malnutrition or malabsorption.

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Age	Male	Female
0-10 years	75.0-153.0 ug/dL	75.0-153.0 ug/dL
11 years-12 years	64.0-132.0 ug/dL	64.0-132.0 ug/dL
13 years-18 years	57.0-129.0 ug/dL	57.0-129.0 ug/dL
19 years and older	70.0-140.0 ug/dL	80.0-155.0 ug/dL



Zinc, Serum or Plasma	
0020097, ZINC	
Specimen Requirements:	
Patient Preparation:	Diet, medication, and nutritional supplements may introduce interfering substances. Upon the advice of their physician, patients should be encouraged to discontinue nutritional supplements, vitamins, minerals, and nonessential over-the- counter medications for one week prior to sample draw.
Collect:	Royal blue (<u>no additive), royal</u> No Additive), Royal blue (K2EDTA), or <u>r</u> Royal blue (NaHep).
Specimen Preparation:	Separate from cells ASAP or within 2 hours of collection. Transfer 2 mL serum or plasma to an ARUP Trace Element-Free Transport Tube (ARUP supply #43116) available online through eSupply using ARUP <u>Connect(TM)orConnector</u> contact ARUP Client Services at (800-)-522-2787- (Min: 0.5 mL). Do not use <u>utensils (i.e., syringes, needles, or pipettes) in the collection or</u> <u>transfer of the sample, pour directly into transport tube.</u>)
Transport Temperature:	Room temperature. Also acceptable: Refrigerated or frozen.
Unacceptable Conditions:	Specimens that are not separated from the red cells or clot within 2 hours. Specimens collected in containers other than specified. Specimens transported in containers other than specified. Hemolyzed specimens.
Remarks:	
Stability:	Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Indefinitely
Methodology:	Quantitative Inductively Coupled Plasma-Mass Spectrometry
Performed:	Sun-Sat
Reported:	1-3 days
Note:	
CPT Codes:	84630
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	



Elevated results may be due to skin or collection-related contamination, including the use of a noncertified metal-free collection/transport tube. If contamination concerns exist due to elevated levels of serum/plasma zinc, confirmation with a second specimen collected in a certified metal-free tube is recommended.

Circulating zinc concentrations are dependent on albumin status and are depressed with malnutrition. Zinc may also be lowered with infection, inflammation, stress, oral contraceptives, and pregnancy. Zinc may be elevated with zinc supplementation or fasting. Elevated zinc concentrations may interfere with copper absorption.

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

60.0-120.0 µg/dL



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TEST CHANGE

Lead, <u>Whole </u> Blood (Venous)	
0020098, LEAD-WB	
Specimen Requirements:	
Patient Preparation:	
Collect:	Royal blue (K2EDTA), Royal blue (NaHep), or tan (K2EDTA).
Specimen Preparation:	Transport 3 or 6 mL whole blood in the original collection tube (royal blue). (Min: 0.5 mL) OR Transport 3 mL whole blood in the original collection tube (tan). (Min: 0.5 mL)
Transport Temperature:	Room temperature. Also acceptable: Refrigerated.
Unacceptable Conditions:	Serum. Specimens collected in tubes other than <u>r</u> Royal blue(K2EDTA), <u>r</u> Royal blue (NaHep), or tan (K2EDTA). Clotted specimens. Capillary pediatric EDTA collection tubes, refer to Lead, <u>Whole</u> Blood (Capillary) 0020745.
Remarks:	
Stability:	Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable
Methodology:	Quantitative Inductively Coupled Plasma-Mass Spectrometry (ICP-MS)
Performed:	Sun-Sat
Reported:	1-3 days
Note:	
CPT Codes:	83655
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	

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Reference intervals are based on the CDC's Blood Lead Reference Value (BLRV). Analysis performed by Inductively Coupled Plasma-Mass Spectrometry (ICP-MS).

Elevated results may be due to skin or collection-related contamination, including the use of a noncertified lead-free tube. If contamination concerns exist due to elevated levels of blood lead, confirmation with a second specimen collected in a certified lead-free tube is recommended.

Information sources for blood lead reference intervals and interpretive comments include the CDC's "Childhood Lead Poisoning Prevention: Recommended Actions Based on Blood Lead Level" and the "Adult Blood Lead Epidemiology and Surveillance: Reference Blood Lead Levels (BLLs) for Adults in the U.S." Thresholds and time intervals for retesting, medical evaluation, and response vary by state and regulatory body. Contact your State Department of Health and/or applicable regulatory agency for specific guidance on medical management recommendations.

Elevated results may be due to skin- or collection-related contamination, including the use of tubes that are not certified to be trace element free. If an elevated result is suspected to be due to contamination, confirmation with a second specimen collected in a certified trace element-free tube is recommended.

Methodology: Inductively Coupled Plasma-Mass Spectrometry (ICP-MS).

Concentration	Comment
5-19.9 ug/dL	Medical removal is recommended for pregnant women or those who are trying or may become pregnant. Adverse health effects are possible. Reduced lead exposure and increased blood lead monitoring are recommended.
20-69.9 ug/dL	Adverse health effects are indicated. Medical removal from lead exposure is required by OSHA if blood lead level exceeds 50 ug/dL. Prompt medical evaluation is recommended.
Greater than 69.9 ug/dL	Critical. Immediate medical

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	evaluation is
	recommended.
	Consider
	chelation therapy
	when symptoms
	of lead toxicity
	are present.
Children	
Concentration	Comment
3.5-19.9 ug/dL	Children under
	the age of 6 years
	are the most
	vulnerable to the
	harmful effects of
	Fourier constants
	Environmental
	exposure history
	to identify
	potential sources
	of lead. Biological
	and nutritional
	monitoring are
	recommended.
	Follow-up blood
	recommended.
20-44 9 µg/dl	Lead bazard
20 44.5 ug/uL	reduction and
	prompt medical
	evaluation are
	recommended.
	Contact a
	Pediatric
	Environmental
	Health Specialty
	control contor for
	quidance
	guidanoe.
Greater than 44.9	Unitediate
ug/u E	medical
	evaluation.
	including detailed
	neurological
	exam is
	recommended.
	Consider
	chelation therapy
	wnen symptoms
	are present
	Contact a
	Pediatric
	Environmental
	Health Specialty
	Unit or poison
	control center for
	assistance.

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Reference Interval:

<u>Test Nu</u>	mberEffecti	ve December 6, 2021	<u>Components</u>	Reference In	<u>terval</u>	 Inserted C	ells	
Age	Reference Internal		<u>Lead, Whole Blood</u> (Venous)	Less than or	equal to 3.4 μg/L	Inserted C	ells	
0-5 years	Less than or equal to 3.4 ug/dL							
6 year or above	Less than or equal to 4.9 ug/dL							
			Lead, Whole Blood					
			<u>(Venous)</u>	1				
				Concentration	Comment			
				<u>3.5-19.9 ug/dL</u>	Children under the age of 6			
					years are the			
					most			
					the harmful			
					effects of lead			
					exposure. Environmental			
					investigation			
					and exposure history to			
					identify			
					potential sources of lead.			
					Biological and			
					nutritional monitoring are			
					recommended.			
					Follow-up			
					monitoring is			
					recommended.			
				<u>20-44.9 ug/dL</u>	Lead hazard			
					prompt medical			
					evaluation are			
					<u>recommended.</u> Contact a			
					Pediatric			
					Environmental Health			
					Specialty Unit			
					or poison control center			
					for guidance.			
				Greater than	Critical.			
				44.9 ug/dL	Immediate medical			
					evaluation,			
					including detailed			
					neurological			
					<u>exam is</u>			

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Consider chelation therapy when symptoms of lead toxicity are present. Contact a Pediatric Environmental Health Specialty Unit or poison control center for assistance.
Concentration Comment
 5-19.9 ug/dL Medical removal is recommended for pregnant women or those who are trying or may become pregnant. Adverse health effects are possible. Reduced lead exposure and increased blood lead monitoring are recommended. 20-69.9 ug/dL Adverse health effects are indicated. Medical removal from lead exposure is required by OSHA if blood lead level exceeds 50 ug/dL. Prompt medical evaluation is recommended.
Greater than 69.9 ug/dL Immediate medical evaluation is recommended. Consider chelation therapy when symptoms of

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	lead toxicity are present.	
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LDL Cholesterol, Direct	
0020257, LDL D	
Specimen Requirements:	
Collect:	Plasma separator tube or serum separator tube. Also acceptable: Green (lithium heparin), <u>I</u> -avender (K2 EDTA or <u>K3EDTAK3-EDTA</u>), or <u>p</u> Pink (K2 EDTA).
Specimen Preparation:	Allow specimen to clot completely at room temperature. Separate serum or plasma from cells ASAP or within 2 hours of collection. Transfer 1 mL serum or plasma to an ARUP <u>standard transport tube.</u> Standard Transport Tube. (Min: 0.5 mL)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	
Remarks:	
Stability:	After separation from cells: Ambient: <u>24</u> 8 hours; Refrigerated: 7 days; Frozen: 12 months
Methodology:	Quantitative Detergent Solubilization/ Enzymatic Assay
Performed:	Sun-Sat
Reported:	Within 24 hours
Note:	
CPT Codes:	83721
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	
CHD Risk Factors: +1 Age: Men, 45 years and older <u>+1 Age:</u> Women, 55 years an	d older or premature menopause without estrogen therapy
 +1 Family history of premature CF +1 Current smoking +1 Hypertension +1 Diabetes mellitus +1 Low HDL <u>c</u>Cholesterol: 39 mg/d -1 High HDL <u>c</u>Cholesterol: 60 mg/d 	ID dL or less dL or greater



LDL Cholesterol: Therapeutic goal 99 mg/dL or less if CHD is present (Optional: 69 mg/dL or less). 129 mg/dL or less if no CHD and two or more risk factors. 159 mg/dL or less if no CHD. (Circulation 2004; 110:227-39)

Reference Interval:

Age	Desirable	Borderline	Higher Risk
0-19 years	109 mg/dL or less	110-129 mg/dL	130 mg/dL or greater
20 years and older	129 mg/dL or less (99 mg/dL or less if patient has CHD)	130-159 mg/dL	160 mg/dL or greater


Lipid Panel, Extended 0020468, CRISK E	
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube or plasma separator tube.
Specimen Preparation:	Allow specimen to clot completely at room temperature. Separate serum or plasma from cells ASAP or within 2 hours of collection. Transfer 1 mL serum or plasma to an ARUP standard transport tube. (Min: 0.5 mL)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Body fluid (refer to Cholesterol, Fluid, ARUP test code 0020714; Triglycerides, Fluid ARUP test code 0020713; and Chylomicron Screen, Body Fluid, ARUP test code 0098457).
Remarks:	
Stability:	After separation from cells: Ambient: 248 hours; Refrigerated: 75 days; Frozen: 3 months
Methodology:	Quantitative Spectrophotometry/Quantitative Enzymatic Assay
Performed:	Sun-Sat
Reported:	Within 24 hours
Note:	LDL-cholesterol is measured (not calculated) on this panel.
CPT Codes:	80061; 83721
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	



An HDL cholesterol less than 40 mg/dL is low and constitutes a coronary heart disease risk factor. An HDL cholesterol greater than 60 mg/dL is a negative risk factor for coronary heart disease.

Non-HDL cholesterol is a secondary target of therapy in persons with high serum triglycerides (greater than 199 mg/dL). The goal for non-HDL cholesterol in persons with high triglycerides is 30 mg/dL higher than their LDL cholesterol goal.

CHD Risk Factors

- +1 Age: Men, 45 years and older
- +1 Women, 55 years and older or premature menopause without estrogen therapy
- +1 Family history of premature CHD
- +1 Current smoking
- +1 Hypertension
- +1 Diabetes mellitus
- +1 Low HDL cholesterol: 39 mg/dL or less
- -1 High HDL cholesterol: 60 mg/dL or greater

20 years and older	Desirable	Borderline	Higher Risk	
Total Cholesterol	199 mg/dL or less	200-239 mg/dL	240 mg/dL or greater	
Triglycerides	149 mg/dL or less	150-199 mg/dL	200-499 mg/dL	
HDL Cholesterol	40 mg/dL or greater		39 mg/dL or less	
LDL Cholesterol	129 mg/dL or less (99 mg/dL or less if patient has CHD)	130-159 mg/dL	160 mg/dL or greater	
VLDL Cholesterol (calculated)	30 mg/dL or less			
0-19 years	Desirable	Borderline	High Risk	
Total Cholesterol	169 mg/dL or less	170-199 mg/dL	200 mg/dL or greater	
Triglycerides	149 mg/dL or less	150-199 mg/dL	200-499 mg/dL	
HDL Cholesterol	40 mg/dL or greater		39 mg/dL or less	
LDL-Cholesterol (measured)	109 mg/dL or less	110-129 mg/dL	130 mg/dL or greater	
VLDL Cholesterol (calculated)	30 mg/dL or less			

Reference Interval:

By Report



Acid Phosphatase, Total, Serun	n
0020544, ACS	
Specimen Requirements:	
Patient Preparation:	
Collect:	Plain red <u>or SST</u> -
Specimen Preparation:	Allow specimen to clot completely at room temperature. Transfer 1.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.5 mL)
Transport Temperature:	CRITICAL FROZEN. Separate specimens must be submitted when multiple tests are ordered.
Unacceptable Conditions:	Plasma. <u>NonfrozenNon-frozen</u> specimens. Hemolyzed specimens.
Remarks:	
Stability:	After separation from cells: <u>Room temperatureAmbient</u> : Unacceptable; Refrigerated: Unacceptable; Frozen: 1 month
Methodology:	Quantitative Enzymatic Assay
Performed:	Sun-Sat
Reported:	1-2 days
Note:	
CPT Codes:	84060
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	
Reference Interval:	
0.0-4.3 U/L	

Heavy Metals Panel 4, <u>Whole</u> Blood		
0020584, HY MET B4		
Specimen Requirements:		
Patient Preparation:	Diet, medication, and nutritional supplements may introduce interfering substances. Patients should be encouraged to discontinue nutritional supplements, vitamins, minerals, non- essential over-the-counter medications (upon the advice of their physician), and avoid shellfish and seafood for 48 to 72 hours.	
Collect:	Royal blue(K2EDTA) or Royal blue (NaHep).	
Specimen Preparation:	Transport 3 or 6 mL whole blood in the original collection tube. (Min: 0.5 mL)	
Transport Temperature:	Room temperature. Also acceptable: Refrigerated.	
Unacceptable Conditions:	Specimens collected in tubes other than Royal blue(K2EDTA) or Royal blue (NaHep). Specimens transported in containers other than Royal blue (K2EDTA) or Royal blue (NaHep). Trace Element-Free Transport Tube. Clotted specimens.	
Remarks:		
Stability:	Ambient: 1 week; Refrigerated: 1 week; Frozen: Unacceptable	
Methodology:	Quantitative Inductively Coupled Plasma-Mass Spectrometry (ICP-MS)	
Performed:	Sun-Sat	
Reported:	1-3 days	
Note:	Mercury is volatile; concentration may decrease over time. If the specimen is drawn and stored in the appropriate container, the arsenic, cadmium, and lead values do not change with time.	
CPT Codes:	82175; 83655; 83825; 82300	
New York DOH Approval Status:	This test is New York DOH approved.	
Interpretive Data:		
Refer to report.		
Reference Interval:		



Test Number	Components	Reference Inte	rval	
Turnser	Lead, <u>Whole</u> Blood (Venous)	Less than or ed	<u>μal to 3.4 μg/L</u>	
	Lead, Whole Blood (Venous)			
		Concentration	Comment	
		3.5-19.9 ug/dL	Children under the age of 6 years are the most vulnerable to the harmful effects of lead exposure. Environmental investigation and exposure history to identify potential sources of lead. Biological and nutritional monitoring are recommended. Follow-up blood lead monitoring is recommended.	
		20-44.9 ug/dL	Lead hazard reduction and prompt medical evaluation are recommended. Contact a Pediatric Environmental Health Specialty Unit or poison control center for guidance.	
		Greater than 44.9 ug/dL	Critical. Immediate medical evaluation, including detailed neurological exam is recommended. Consider chelation therapy when symptoms of lead toxicity are present. Contact a Pediatric Environmental Health Specialty Unit or poison control center for	
	Lead, Whole Blood (Venous)		assistance.	



	Concentration	Comment
	5-19.9 ug/dL	Medical removal is recommended for pregnant women or those who are trying or may become pregnant. Adverse health effects are possible. Reduced lead exposure and increased blood lead monitoring are recommended.
	20-69.9 ug/dL	Adverse health effects are indicated. Medical removal from lead exposure is required by OSHA if blood lead level exceeds 50 ug/dL. Prompt medical evaluation is recommended.
	Greater than 69.9 ug/dL	Critical. Immediate medical evaluation is recommended. Consider chelation therapy when symptoms of lead toxicity are present.
Lead, Blood (Venous)		· · · · · · · · · · · · · · · · · · ·
	Age	Reference Interval (ug/dL)
	0-5 years	Less than or equal to 3.4
	6 years or above	Less than or equal to 4.9
Arsenic <u>, Whole</u> Blood	Less than or ec	ual to 12 .0 μg/L
Mercury <u>. Whole</u> Blood	Less than or ec	jual to 10.0 μg/L
Cadmium, <u>Whole</u> Blood	Less than or ec	jual to 5.0 μg/L



Zinc Protoporphyrin (ZPP), Whole Blood Industrial 0020614, ZPP IND Specimen Requirements: Patient Preparation: Collect: Lavender (EDTA), royal blue (K2EDTA), royal blue (NaHep), tan (K2EDTA) or pink (K2EDTA). **Specimen Preparation:** Transport 3 or 6 mL whole blood in the original collection tube. (Min: 0.2 mL) Refrigerated. Transport Temperature: Unacceptable Conditions: Clotted, frozen, or hemolyzed specimens. Remarks: Stability: Ambient: 30 hours; Refrigerated: 5 weeks; Frozen: Unacceptable Methodology: Quantitative Hematofluorometry Performed: Mon-Fri Reported: 1-4 days Note: Elevated ZPP results are seen in early and late iron deficiency, the anemia of chronic disease, chronic lead poisoning, and erythropoietic protoporphyria. Elevated bilirubin or riboflavin and hemolyzed, clotted, or improperly aliquoted specimens may falsely increase the ZPP concentration. CPT Codes: 84202 New York DOH Approval Status: This test is New York DOH approved. Interpretive Data:

For occupational exposure to lead, OSHA requires ZPP whole blood concentration to be reported in units of ug/dL. For adults, conversion of ZPP to units of ug/dL assumes a hematocrit of 45%. This test was performed on the ProtoFluor Z system manufactured by Helena Laboratories. The result is not comparable to results obtained from extraction-based methods or from the AVIV ZPP system.

Reference Interval:

Test Number	Components	Reference Interval
	Zinc Protoporphyrin <u>(ZPP), Whole</u> , Blood	0-40 μg/dL



Zinc Protoporphyrin (ZPP) WholeBld Ratio 0-69 µmol ZPP/mol heme



TEST CHANGE

Lead, <u>Whole</u> Blood (Capillary)	
Specimen Bequirements:	
Patient Preparation:	Clean puncture site well with soap and water before collection procedure begins.
Collect:	Lavender microtainer (K2EDTA)
Specimen Preparation:	Invert specimen 10 times to prevent clot formation. Transport 0.5 mL whole blood in the original collection tube. (Min: 0.3 mL)
Transport Temperature:	Room temperature. Also acceptable: Refrigerated.
Unacceptable Conditions:	Specimens collected in tubes other than lavender microtainer (K2EDTAK[2]EDTA). Specimens transported in tubes other than trace element-free transport tubes or lavender microtainer (K2EDTAK[2]EDTA) tubes. Heparin anticoagulant. Clotted specimens. Venous whole blood, refer to Lead, Whole Blood (Venous) (ARUP test code 0020098).
Remarks:	
Stability:	Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable
Methodology:	Quantitative Inductively Coupled Plasma-Mass Spectrometry (ICP-MS)
Performed:	Sun-Sat
Reported:	1-3 days
Note:	
CPT Codes:	83655
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	



<u>Reference intervals are based on the CDC's Blood Lead Reference Value (BLRV). Analysis performed</u> by inductively coupled plasma-mass spectrometry (ICP-MS).

Elevated results may be due to skin or collection-related contamination, including the use of a noncertified lead-free collection/transport tube. If contamination concerns exist due to elevated levels of blood lead, confirmation with a venous specimen collected in a certified lead-free tube is recommended.

Repeat testing is recommended prior to initiating chelation therapy or conducting environmental investigations of potential lead sources. Repeat testing collections should be performed using a venous specimen collected in a certified lead-free collection tube.

Information sources for blood lead reference intervals and interpretive comments include the CDC's "Childhood Lead Poisoning Prevention: Recommended Actions Based on Blood Lead Level" and the "Adult Blood Lead Epidemiology and Surveillance: Reference Blood Lead Levels (BLLs) for Adults in the U.S." Thresholds and time intervals for retesting, medical evaluation, and response vary by state and regulatory body. Contact your State Department of Health and/or applicable regulatory agency for specific guidance on medical management recommendations.

<u>Capillary collections are prone to contamination from skin and from use of nontrace element-free</u> <u>collection tubes. Results above the reference interval should be confirmed with a venous specimen</u> <u>collected in a certified trace element-free tube.</u>

Methodology: Inductively Coupled Plasma-Mass Spectrometry (ICP-MS).

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Concentration	Comment
5-19.9 ug/dL	Medical removal is recommended for pregnant women or those who are trying or may become pregnant. Adverse health effects are possible. Reduced lead exposure and increased blood lead monitoring are recommended.
20-69.9 ug/dL	Adverse health effects are indicated. Medical removal from lead exposure is required by OSHA if blood lead level exceeds 50 ug/dL. Prompt medical



	evaluation is recommended
Greater than 69.9 ug/dL	Critical. Immediate medical
	evaluation is recommended. Consider
	when symptoms of lead toxicity are present.
Children	
Concentration	Comment
3.5-19.9 ug/dL	Children under the age of 6 years are the most vulnerable to the harmful effects of lead exposure. Environmental investigation and exposure history to identify potential sources of lead. Biological and nutritional monitoring are recommended. Follow-up blood lead monitoring is recommended.
20-44.9 ug/dL	Lead hazard reduction and prompt medical evaluation are recommended. Contact a Pediatric Environmental Health Specialty Unit or poison control center for guidance.
Greater than 44.9 ug/dL	Critical. Immediate medical evaluation, including detailed neurological exam is recommended. Consider chelation therapy when symptoms of lead toxicity are present. Contact a Pediatric



Environmental
Health Specialty
Unit or poison
control center for
assistance.

Reference Interval:

<u>Test</u> <u>Number</u>	<u>Components</u>	Reference Interval
	Lead, Whole Blood (Capillary)	Less than or equal to 3.4 μ g/dL

Effective December 6, 2021

0-5 years	Less than or equal to 3.4 ug/dL
6 years or above	Less than or equal to 4.9 ug/dL

Citric Acid, Urine					
0020852, CITRIC U					
Specimen Requirements:					
Patient Preparation:					
Collect:	24-hour urine. Refrigerate during collection. Also acceptable: Random urine.				
Specimen Preparation:	Adjust pH to less than or equal to 2 by adding 6M HCl. Transfer a 4 mL aliquot of urine to an ARUP <u>standard transport</u> <u>tubeStandard Transport Tube</u> . (Min: 0.5 mL) Record total volume, collection time interval, and pH on transport tube and test request form. Also acceptable: Specimens previously preserved with boric acid.				
Transport Temperature:	Refrigerated.				
Unacceptable Conditions:					
Remarks:					
Stability:	Ambient: 248 hours; Refrigerated: 2 weeks; Frozen: 1 month				
Methodology:	Quantitative Enzymatic Assay				
Performed:	Sun-Sat				
Reported:	Within 24 hours				
Note:					
CPT Codes:	82507				
New York DOH Approval Status:	This test is New York DOH approved.				

Interpretive Data:

Per 24h calculations are provided to aid interpretation for collections with a duration of 24 hours and an average daily urine volume. For specimens with notable deviations in collection time or volume, ratios of analytes to a corresponding urine creatinine concentration may assist in result interpretation.

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:



Test Number	Components	Reference Interval			
	<u>Creatinine</u> Citric Acid, Urine - per 24h	18 years and older: 320-1240 mg/d			
		<u>Age</u>	Male (mg/d)	Female (mg/d)	
		<u>3-8 years</u>	<u>140-700</u>	<u>140-700</u>	
		<u>9-12 years</u>	<u>300-1300</u>	<u>300-1300</u>	
		<u>13-17 years</u>	<u>500-2300</u>	<u>400-1600</u>	
		<u>18-50 years</u>	<u>1000-2500</u>	<u>700-1600</u>	
		51-80 years	800-2100	<u>500-1400</u>	
		<u>81 years and</u> older	<u>600-2000</u>	<u>400-1300</u>	
	Citric Acid/Creatinine Ratio, Urine	1 year and old mg/g.	er: greater than	or equal to 150	
	Citric Acid, Urine - per 24h		18 years and older: 320-1240 mg/d		



Cadmium Exposure Panel - OSHA

0025013, CD EXP

Specimen Requirements:				
Patient Preparation:	To avoid contamination, please collect specimens at the beginning of work shift. Blood and urine should be collected the same day. Urine: Diet, medication, and nutritional supplements may introduce interfering substances. Patients should be encouraged to discontinue nutritional supplements, vitamins, minerals, and non-essential over-the-counter medications (upon the advice of their physician). High concentrations of iodine may interfere with elemental testing. Collection of urine specimens from patients receiving iodinated or gadolinium-based contrast media should be avoided for a minimum of 72 hours post-exposure. Collection from patients with impaired kidney function should be avoided for a minimum of 14 days post contrast media exposure.			
Collect:	Royal blue (K2EDTA) or royal blue (NaHep). AND minimum 40 mL urine using spot technique (single void) in an open-top urine collection cup.			
Specimen Preparation:	Transfer specimens to the appropriate transport device using the Cadmium exposure kit, ARUP supply #16450, available online through eSupply using ARUP Connect(TM) or by contacting ARUP Client Services at (800) 522-2787. Blood: Transport 3 or 6 mL whole blood in the original collection tube. (Min: 0.5 mL) Urine for Beta-2-Microglobulin: Transfer 3 mL aliquot from original urine collection to an ARUP Standard Transport Tube. Adjust the pH of this specimen immediately after pouring off collection, so the pH is between 6 and 8. Use 1M HCl or 5 percent NaOH to adjust the urine pH. Label tube as beta2 Microglobulin. Freeze within one hour of collection. Urine for Cadmium: Transfer 7 mL aliquot from original urine collection to ARUP Trace Element-Free Transport Tubes (ARUP supply #43116). Available online through eSupply using ARUP Connect(TM) or by contacting ARUP Client Services at (800) 522-2787. (Min: 0.5 mL) Label tube as Cadmium. Urine for Creatinine: Transfer 2 mL aliquot from original urine collection to an ARUP Standard Transport Tube. (Min: 0.5 mL) Label tube as Creatinine.			
Transport Temperature:	Blood: Refrigerated. Urine for Beta-2-Microglobulin: Frozen Urine for Cadmium: Refrigerated. Urine for Creatinine:			
	nenigerateu.			

	(K2EDTA) or royal blue (NaHep). Specimens transported in containers other than royal blue (K2EDTA) or royal blue (NaHep) tube or trace element-free transport tube. Clotted specimens. Urine: Specimens transported in <u>nontracenon- trace</u> element <u>-</u> free transport tube (with the exception of the original device).Specimens collected within 72 hours after administration of iodinated or gadolinium-based contrast media. Specimens containing blood or fecal materials.
Remarks:	Record total volume and collection time interval on transport tube and on test request form.
Stability:	Blood: Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable Urine for Beta-2-Microglobulin: Ambient: 8 hours; Refrigerated: 48 hours; Frozen: 2 months Urine for Cadmium: Ambient: 1 week; Refrigerated: 2 weeks; Frozen: 1 year Urine for Creatinine: Ambient: 2 days; Refrigerated: 1 month; Frozen: 6 months
Methodology:	Quantitative Inductively Coupled Plasma-Mass Spectrometry (ICP-MS)/Spectrophotometry/Chemiluminescent Immunoassay(CLIA)
Performed:	Mon-Fri
Reported:	1-5 days
Note:	
CPT Codes:	82300 x2; 82232
New York DOH Approval Status:	This test is New York DOH approved.

Interpretive Data:

Blood cadmium levels can be used to monitor acute toxicity and, in combination with cadmium urine and <u>B-</u>³2 microglobulin, is the preferred method for monitoring occupational exposure. Symptoms associated with cadmium toxicity vary based upon route of exposure and may include tubular proteinuria, fever, headache, dyspnea, chest pain, conjunctivitis, rhinitis, sore throat, and cough. Ingestion of cadmium in high concentration may cause vomiting, diarrhea, salivation, cramps, and abdominal pain.

Urine cadmium levels can be used to assess cadmium body burden. In chronic exposures, the kidneys are the primary target organ. Symptoms associated with cadmium toxicity vary based upon route of exposure and may include tubular proteinuria, fever, headache, dyspnea, chest pain, conjunctivitis, rhinitis, sore throat, and cough. Ingestion of cadmium in high concentration may cause vomiting, diarrhea, salivation, cramps, and abdominal pain.

Urine \underline{B} - $\beta 2 \underline{m}$ \underline{M} icroglobulin is an early marker of irreversible kidney damage and disease.



Urine creatinine values less than 20 mg/dL represent very dilute urine and collections should be repeated.

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

CADMIUM ACTION LEVELS BEGINNING JANUARY 1999 (Federal Register 1999, Std. CFR, Part 1910. 1027 Appendix

A)

Components	A	В	С
Cadmium, Urine (ug/g CRT)	{<=} 3	> 3 to {<=} 7	>7
Cadmium, Blood (ug/L)	0-5	> 5 to {<=} 10	> 10
β2 Microglobulin, Urine (ug/g CRT)	{<=} 300	> 300 to {<=} 750	> 750*
Monitor	Annual	Semiannual	Quarterly
Medical Exam	Biennial	Annual	Semiannual
Reassess Cadmium exposure in less than two weeks		Discretionary	Mandatory removal

*If an employee's β2 Microglobulin level is above 750 ug/g CRT, in order for mandatory medical removal to be required, either the employee's CdU level must also be >3 ug/g CRT or CdB level must also be >5 ug/L. The determination of discretionary or mandatory removal is made by the examining physician consistent with the medical surveillance specifications in the Federal Register 42456 to 42463. References: 1. US Department of Labor (2004). Cadmium. Occupational Safety and Health Administration. 3136-06R. 2. US Department of Labor (1999). Cadmium. Occupational Safety and Health Standards. 1910.1027

Reference Interval:

Test Number	Components	Reference Interval
	Cadmium, Urine - ratio to CRT	0.0-3.2 μg/g CRT
	Cadmium, Urine - per volume	0.0-1.0 μg/L
	Beta-2-Microglobulin, ratio to CRT	0-300 μg/g CRT
	Beta-2-Microglobulin, Urine	0-300 μg/L
	Cadmium, <u>Whole</u> Blood	Less than or equal to 5.0 μ g/L



Lead, Industrial <u>, Whole Blood</u> -Exposure Panel, Adults				
0025016, LEAD-IND				
Specimen Requirements:				
Patient Preparation:	Collect from patient aged 16 years or older.			
Collect:	Royal blue(K2EDTA), Royal blue (NaHep) or tan (K2EDTA).			
Specimen Preparation:	Transport 3 or 6 mL whole blood (royal blue) (Min: 0.5 mL) OR Transport 3 mL whole blood (tan) (Min: 0.5 mL)			
Transport Temperature:	Refrigerated.			
Unacceptable Conditions:	Serum. Specimens collected in tubes other than Royal blue(K2EDTA), Royal blue (NaHep), or tan (K2EDTA). Hemolyzed or clotted specimens.			
Remarks:				
Stability:	Ambient: 30 hours; Refrigerated: 5 weeks; Frozen: Unacceptable			
Methodology:	Quantitative Inductively Coupled Plasma-Mass Spectrometry (ICP-MS)/Hematofluorometry			
Performed:	Sun-Sat			
Reported:	1-5 days			
Note:				
CPT Codes:	83655; 84202			
New York DOH Approval Status:	This test is New York DOH approved.			
Interpretive Data:				



Interpretive Data

<u>Reference intervals are based on the CDC's Blood Lead Reference Value (BLRV). Analysis performed</u> by Inductively Coupled Plasma-Mass Spectrometry (ICP-MS).

Elevated results may be due to skin or collection-related contamination, including the use of a noncertified lead-free collection/transport tube. If contamination concerns exist due to elevated levels of blood lead, confirmation with a second specimen collected in a certified lead-free tube is recommended.

Reference interval and interpretive comments are based on the CDC's "Childhood Lead Poisoning Prevention: Recommended Actions Based on Blood Lead Level" and the "Adult Blood Lead Epidemiology and Surveillance: Reference Blood Lead Levels (BLLs) for Adults in the U.S." Thresholds and time intervals for retesting, medical evaluation, and response vary by state and regulatory body. Actions described by OSHA in 1978 and finalized in 1983 are shown below. Contact your State Department of Health and/or applicable regulatory agency for specific guidance on medical management recommendations. Actions described by OSHA in 1978 and finalized in 1983 are shown below.

Elevated results may be due to skin- or collection-related contamination, including the use of tubes that are not certified to be trace element free. If an elevated result is suspected to be due to contamination, confirmation with a second specimen collected in a certified trace element-free tube is recommended.

Concentration	Comment	
5-19.9 ug/dL	Medical removal is recommended for pregnant women or those who are trying or may become pregnant. Adverse health effects are possible. Reduced lead exposure and increased blood lead monitoring are recommended.	
20-69.9 ug/dL	Adverse health effects are indicated. Medical removal from lead exposure is required by OSHA if blood lead level exceeds 50 ug/dL. Prompt medical evaluation is	

Methodology: Inductively Coupled Plasma-Mass Spectrometry (ICP-MS).



	recommended.	
Greater than 69.9	Critical.	
ug/dL	Immediate	
	medical	
	evaluation is	
	recommended.	
	Consider	
	chelation therapy	
	when symptoms	
	of lead toxicity	
	are present.	

"Occupational Safety and Health Standards: Lead (1983). 29 CFR Part 1910.1025 App C" Action required for workers with Elevated Lead Values OSHA, Occupational Exposure to Lead, 1978

No. of Tests	Lead	Action Required
1	Greater than equal to 40.0 ug/dL	Notification of worker in writing; medical examination of worker and consultation.
3 (average)	Greater than or equal to 50.0 ug/dL	Removal of worker from job with potential lead exposure.
1	Greater than or equal to 60.0 ug/dL	Removal of worker from job with potential lead exposure.
2	Less than 40.0 ug/dL	Reinstatement of worker in job with potential lead exposure is based upon symptoms and medical evaluation.

OSHA requirements in effect since 1978 call for the measurement of whole blood lead and zinc protoporphyrins (ZPP) (NCCLS document C42-A, Nov. 1996) to evaluate the occupational exposure to lead. OSHA requires ZPP whole blood testing to be reported in units of ug/dL. For adults, conversion of ZPP units of ug/dL whole blood assumes a hematocrit of 45 percent. Conversion factor: umol/mol heme x 0.584= ug/dL.

Reference Interval:



Test Number	Components	Reference Interval
	Lead, Industrial, Whole Blood	Less than or equal to $3.4.9 \ \mu$ g/dL
	Zinc Protoporphyrin <u>(ZPP), Whole</u> , Blood	0-40 μg/dL
	Zinc Protoporphyrin (ZPP) WholeBld Ratio	0-69 μmol ZPP/mol heme



Selenium, Serum or Plasma 0025023, SE S	
Specimen Requirements:	
Patient Preparation:	Diet, medication, and nutritional supplements may introduce interfering substances. Patients should be encouraged to discontinue nutritional supplements, vitamins, minerals, and <u>nonessentialnon-essential</u> over-the-counter medications (upon the advice of their physician).
Collect:	Royal blue (<u>no additive), royal</u> No <u>Additive), Royal</u> blue (K2EDTA), or <u>r</u> Royal blue (NaHep).
Specimen Preparation:	Separate from cells ASAP or within 2 hours of collection. Transfer 2 mL serum or plasma to an ARUP Trace Element-Free Transport Tube (ARUP supply #43116) available online through eSupply using ARUP Connect(TM) or contact ARUP Client Services at (800-)-522-2787- (Min: 0.5 mL). Do not use utensils (i.e., syringes, needles, or pipettes) in the collection or transfer of the sample, pour directly into transport tube.)
Transport Temperature:	Room temperature. Also acceptable: Refrigerated or frozen.
Unacceptable Conditions:	Specimens that are not separated from the red cells or clot within 2 hours. Specimens collected in containers other than specified. Specimens transported in containers other than specified.
Remarks:	
Stability:	Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Indefinitely
Methodology:	Quantitative Inductively Coupled Plasma-Mass Spectrometry
Performed:	Sun-Sat
Reported:	1-3 days
Note:	
CPT Codes:	84255
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	



Elevated results may be due to contamination from skin or other collection-related issues, including the use of a noncertified metal-free collection/transport tube. If contamination concerns exist due to elevated levels of serum/plasma selenium, confirmation with a second specimen collected in a certified metal-free tube is recommended.

Serum selenium levels can be used in the determination of deficiency or toxicity. Plasma and serum contains 75 percent of the selenium measured in whole blood and reflects recent dietary intake. Selenium deficiency can occur endemically or as a result of sustained TPN or restricted diets and has been associated with cardiomyopathy and may exacerbate hypothyroidism. Selenium toxicity is relatively rare. Excess intake of selenium can result in symptoms consistent with selenosis and include gastrointestinal upset, hair loss, white blotchy nails, and mild nerve damage.

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

23.0-190.0 μg/L

Cobalt, Serum or Plasma	
0025037, COBALT S	
Specimen Requirements:	
Patient Preparation:	Diet, medication, and nutritional supplements may introduce interfering substances. Patients should be encouraged to discontinue nutritional supplements, vitamins, minerals, and <u>nonessentialnon-essential</u> over-the-counter medications (upon the advice of their physician).
Collect:	Royal blue (<u>no additive), royal</u> No Additive), Royal blue (K2EDTA), or <u>r</u> Royal blue (NaHep).
Specimen Preparation:	Separate from cells ASAP or within 2 hours of collection. Transfer 2 mL serum or plasma to an ARUP Trace Element-Free Transport Tube (ARUP supply #43116) available online through eSupply using ARUP Connect(TM) or contact ARUP Client Services at (800-)-522-2787- (Min: 0.5 mL). Do not use utensils (i.e., syringes, needles, or pipettes) in the collection or transfer of the sample, pour directly into transport tube.)
Transport Temperature:	Room temperature. Also acceptable: Refrigerated or frozen.
Unacceptable Conditions:	Specimens collected in containers other than specified. Specimens transported in containers other than specified.
Remarks:	
Stability:	Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Indefinitely
Methodology:	Quantitative Inductively Coupled Plasma-Mass Spectrometry
Performed:	Sun-Sat
Reported:	1-3 days
Note:	
CPT Codes:	83018
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	
Elevated results may be due to skin noncertified metal-free collection/r	n or collection-related contamination, including the use of a transport tube. If contamination concerns exist due to elevated

levels of serum/plasma cobalt, confirmation with a second specimen collected in a certified metal-



free tube is recommended.

Serum cobalt levels can be used in the assessment of occupational exposure or toxic ingestion. Symptoms associated with cobalt toxicity vary based on route of exposure, and may include cardiomyopathy, allergic dermatitis, pulmonary fibrosis, cough, and dyspnea.

Serum cobalt levels can be significantly higher in patients with metal-on-metal total hip replacement implants than in control patients without metal implants. Serum cobalt levels may be increased in asymptomatic patients with metal-on-metal prosthetics and should be considered in the context of the overall clinical scenario. Whole blood is the specimen type recommended by the U.S. Food and Drug Administration for assessing the risks of metal-on-metal hip implants in symptomatic patients.-

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Less than or equal to 1.0 $\mu\text{g/L}$



Entamoeba histolytica (amebiasis), Antibody, IgG

0050070, AMB	
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube.
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 1 mL serum to an ARUP <u>standard transport</u> <u>tubeStandard Transport Tube</u> . (Min: 0.1 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of the acute specimens. Mark specimens plainly as "acute" or "convalescent."
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Contaminated, heat-inactivated, hemolyzed, or severely lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)
Methodology:	Semi-Quantitative Enzyme-Linked Immunosorbent Assay (ELISA)
Performed:	Tue , Fri
Reported:	1- <u>8</u> 5 days
Note:	In the case of extraintestinal complications, a positive antibody can indicate amebiasis even though stool findings are negative.
CPT Codes:	86753
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	

Seroconversion between acute and convalescent sera is considered strong evidence of recent infection. The best evidence for infection is a significant change on two appropriately timed specimens where both tests are done in the same laboratory at the same time.

Reference Interval:

Effective February 6, 2017



8 U or less: Negative - No significant level of detectable *E. histolytica* IgG antibody.

9 - 11 U: Equivocal - Repeat testing in 10-14 days may be helpful.

12 U or greater: Positive - IgG antibody to *E. histolytica* detected, suggestive of a current or past infection.



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O050165, CMV IGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum Separator Tube (SST).
Specimen Preparation:	Allow specimen to clot completely at room temperature. Separate from cells ASAP or within 2 hours of collection. Transfer 1 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.5 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of the acute specimens.
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Contaminated, heat-inactivated, <u>icteric, or</u> grossly hemolyzed specimens.
Remarks:	Label specimens plainly as "acute" or "convalescent."
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (Avoid repeated freeze/thaw cycles)
Methodology:	Semi-Quantitative Chemiluminescent Immunoassay (CLIA)
Performed:	Sun-Sat
Reported:	Within 24 hours
Note:	
CPT Codes:	86644
New York DOH Approval Status:	This test is New York DOH approved.

Interpretive Data:

In immunocompromised patients, CMV serology (IgG or IgM antibody titers) may not be reliable and may be misleading in the diagnosis of acute or reactivation CMV disease. The preferred method for diagnosis is culture of virus and/or demonstration of viral antigen in peripheral white cells (buffy coat), bronchoalveolar lavage (BAL) cells, or tissue biopsies.

This test should not be used for blood donor screening, associated re-entry protocols, or for screening <u>human cell, tissues</u>, <u>Human Cell, Tissues</u> and <u>c</u>Cellular<u>-</u> and <u>tissue-based</u> <u>products</u><u>Tissue-Based Products</u> (HCT/P).



The best evidence for current infection is a significant change on two appropriately timed specimens, where both tests are done in the same laboratory at the same time.

Reference Interval:

0.59 U/mL or less:	Not Detected.
0.60-0.69 U/mL:	Indeterminate - Repeat testing in 10-14 days may be helpful.
0.70 U/mL or greater:	Detected.



Effective Date: July 21, 2025

TEST CHANGE

Epstein-Barr Virus Antibody to Early D Antigen (EA-D), IgG

. 0050225, EBV EAD

Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube (SST).
Specimen Preparation:	Allow specimen to clot completely at room temperature. Separate serum from cells ASAP or within 2 hours of collection. Transfer 2 mL serum to an ARUP standard transport tube. (Min: 0.5 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of the acute specimens.
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Contaminated, heat-inactivated, icteric, or grossly hemolyzed specimens.
Remarks:	Label specimens plainly as "acute" or "convalescent."
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)
Methodology:	Semi-Quantitative Chemiluminescent Immunoassay
Performed:	Sun-Sat
Reported:	1-2 days
Note:	EBV EA-D values obtained with different manufacturers' assay methods may not be used interchangeably. The magnitude of the reported EBV EA-D level cannot be correlated to an endpoint titer.
CPT Codes:	86663
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	
	*
	8.9 U/mL or less: Not Detected 9.0-10.9 U/mL: Indeterminate - Repeat testing in 10- 14 days may be helpful. 11.0 U/mL or greater: Detected

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Effective Date: July 21, 2025

Reference Interval:

Effective February 19, 2013

8.9 U/mL or less: Not Detected 9.0-10.9 U/mL: Indeterminate - Repeat testing in 10-14 days may be helpful. 11.0 U/mL or greater: Detected **Deleted Cells**

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Effective Date: July 21, 2025

TEST CHANGE

Epstein-Barr Virus Antibody to Viral Capsid Antigen, IgG

0050235, EBV G

Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube (SST).
Specimen Preparation:	Allow specimen to clot completely at room temperature. Separate from cells ASAP or within 2 hours of collection. Transfer 2 mL serum to an ARUP standard transport tube. (Min: 0.5 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of the acute specimens.
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Contaminated, heat-inactivated, icteric, or grossly hemolyzed specimens.
Remarks:	Label specimens plainly as "acute" or "convalescent."
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)
Methodology:	Semi-Quantitative Chemiluminescent Immunoassay
Performed:	Sun-Sat
Reported:	1-2 days
Note:	EBV IgG values obtained with different manufacturers' assay methods may not be used interchangeably. The magnitude of the reported EBV IgG level cannot be correlated to an endpoint titer.
CPT Codes:	86665
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	A
	<u>17.9 U/mL or less: Not Detected</u> 18.0-21.9 U/mL: Indeterminate. Repeat testing in 10-14 days may be helpful. 22.0 U/mL or greater: Detected

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Inserted Cells



Effective Date: July 21, 2025

Reference Interval:

Effective February 19, 2013

17.9 U/mL or less: Not Detected 18.0-21.9 U/mL: Indeterminate. Repeat testing in 10-14 days may be helpful. 22.0 U/mL or greater: Detected **Deleted Cells**

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Effective Date: July 21, 2025

TEST CHANGE

Epstein-Barr Virus Antibody to Viral Capsid Antigen, IgM

0050240, EBV M

Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube (SST).
Specimen Preparation:	Allow specimen to clot completely at room temperature. Separate from cells ASAP or within 2 hours of collection. Transfer 2 mL serum to an ARUP standard transport tube. (Min: 0.5 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of the acute specimens.
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Contaminated, heat-inactivated, icteric, or grossly hemolyzed specimens.
Remarks:	Label specimens plainly as "acute" or "convalescent."
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)
Methodology:	Semi-Quantitative Chemiluminescent Immunoassay
Performed:	Sun-Sat
Reported:	1-2 days
Note:	EBV IgM values obtained with different manufacturers' assay methods may not be used interchangeably. The magnitude of the reported EBV IgM level cannot be correlated to an endpoint titer.
CPT Codes:	86665
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	

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Inserted Cells



Effective Date: July 21, 2025

Reference Interval:

Effective February 19, 2013

Deleted Cells

35.9 U/mL or less: Not Detected 36.0-43.9 U/mL: Indeterminate. Repeat testing in 10-14 days may be helpful. 44.0 U/mL or greater: Detected

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Effective Date: July 21, 2025

TEST CHANGE

Epstein-Barr Virus Antibody to Nuclear Antigen, IgG

specimen requirements:	
Patient Preparation:	
Collect:	Serum separator tube (SST).
Specimen Preparation:	Allow specimen to clot completely at room temperature. Separate from cells ASAP or within 2 hours of collection. Transfer 2 mL serum to an ARUP standard transport tube. (Min: 0.5 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of the acute specimens.
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Contaminated, heat-inactivated, icteric, or grossly hemolyzed specimens.
Remarks:	Label specimens plainly as "acute" or "convalescent."
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)
Methodology:	Semi-Quantitative Chemiluminescent Immunoassay
Performed:	Sun-Sat
Performed: Reported:	Sun-Sat 1-2 days
Performed: Reported: Note:	Sun-Sat 1-2 days EBNA values obtained with different manufacturers' assay methods may not be used interchangeably. The magnitude of the reported EBNA level cannot be correlated to an endpoint titer.
Performed: Reported: Note: CPT Codes:	Sun-Sat 1-2 days EBNA values obtained with different manufacturers' assay methods may not be used interchangeably. The magnitude of the reported EBNA level cannot be correlated to an endpoint titer. 86664
Performed: Reported: Note: CPT Codes: New York DOH Approval Status:	Sun-Sat 1-2 days EBNA values obtained with different manufacturers' assay methods may not be used interchangeably. The magnitude of the reported EBNA level cannot be correlated to an endpoint titer. 86664 This test is New York DOH approved.
Performed: Reported: Note: CPT Codes: New York DOH Approval Status: Interpretive Data:	Sun-Sat 1-2 days EBNA values obtained with different manufacturers' assay methods may not be used interchangeably. The magnitude of the reported EBNA level cannot be correlated to an endpoint titer. 86664 This test is New York DOH approved.

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Inserted Cells


A nonprofit enterprise of the University of Utah and its Department of Pathology

Effective Date: July 21, 2025

Reference Interval:

Effective February 19, 2013

17.9 U/mL or less: Not Detected 18.0-21.9 U/mL: Indeterminate. Repeat testing in 10-14 days may be helpful. 22.0 U/mL or greater: Detected **Deleted Cells**

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Haptoglobin	
0050280, HAPTO	
Specimen Requirements:	
Patient Preparation:	Fasting specimen preferred.
Collect:	Plasma separator tube or serum separator tube. Also acceptable: Green (lithium heparin), or pink (K2EDTA).
Specimen Preparation:	Allow specimen to clot completely at room temperature. Separate serum or plasma from cells ASAP or within 2 hours of collection. Transfer 1 mL serum or plasma to an ARUP <u>standard transport tube.Standard Transport Tube.</u> (Min: 0.5 mL)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Grossly hemolyzed specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 3 months; Refrigerated: 8 months; Frozen: <u>3</u> + months
Methodology:	Quantitative Immunoturbidimetry
Performed:	Sun-Sat
Reported:	Within 24 hours
Note:	
CPT Codes:	83010
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	
Reference Interval:	
30-200 mg/dL	



Herpes Simplex Virus Type 1 an 0050293, HERPES	nd/or 2 Antibodies, IgG
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube. Also acceptable: Serum from umbilical cord blood.
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 1 mL of serum to an ARUP <u>standard transport</u> <u>tubeStandard Transport Tube</u> . (Min: 0.5 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of the acute specimens. Mark specimens plainly as "acute" or "convalescent."
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Urine. CSF (refer to Herpes Simplex Virus Type 1 and/or 2 Antibodies, IgG, CSF, ARUP test code 0050394). Contaminated, heat-inactivated, icteric, or hemolyzed specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)
Methodology:	Semi-Quantitative Chemiluminescent Immunoassay
Performed:	Sun-Sat
Reported:	Within 24 hours
Note:	
CPT Codes:	86694
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	
The best evidence for current infect	tion is a significant change on two appropriately timed

specimens, where both tests are done in the same laboratory at the same time.

False positive results are possible. Consider additional testing for HSV-2, particularly if the result for HSV-2 is </= 3.0 IV.



0.89 IV or less:	Not Detected.
0.90-1.09 IV:	Indeterminate - Repeat testing in 10-14 days may be helpful.
1.10 IV or greater:	Detected.



Herpes Simplex Virus Type 1 Glycoprotein G-Specific Antibody, IgG by ELISA, CSF

0050379, HERPICSF	
Specimen Requirements:	
Patient Preparation:	
Collect:	CSF.
Specimen Preparation:	Transfer 0.5 mL CSF to an ARUP Standard Transport Tube. (Min: 0.2 mL)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Specimen types other than CSF. Contaminated, heat- inactivated, <u>icteric,</u> or hemolyzed specimens.
Remarks:	Indicate source on test request form.
Stability:	Ambient: 8 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Semi-Quantitative <u>Chemiluminescent Immunoassay</u> (<u>CLIA)</u> Enzyme-Linked Immunosorbent Assay
Performed:	Mon, Wed, Fri
Reported:	1-5 days
Note:	
CPT Codes:	86695
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.

Interpretive Data:

Individuals infected with HSV may not exhibit detectable IgG antibody to type-specific HSV antigens 1 and 2 in the early stages of infection. Detection of antibody presence in these cases may only be possible using a nontype-specific screening test.

The detection of antibodies to herpes simplex virus in CSF may indicate central nervous system infection. However, consideration must be given to possible contamination by blood or transfer of serum antibodies across the blood-brain barrier.

Fourfold or greater rise in CSF antibodies to herpes on specimens at least 4 weeks apart are found in 74-94 percent of patients with herpes encephalitis. Specificity of the test based on a single CSF testing is not established. Presently PCR is the primary means of establishing a diagnosis of herpes encephalitis.



This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

0.89 IV or less	Negative - No significant level of detectable IgG antibody to HSV type 1 glycoprotein G.	
0.90-1.10 IV	Equivocal - Questionable presence of IgG antibody to HSV type 1. Repeat testing in 10-14 days may be helpful.	
1.11 IV or greater	Positive - IgG antibody to HSV type 1 glycoprotein G detected, which may indicate a current or past infection.	



Herpes Simplex Virus Type 1 and/or 2 Antibodies, IgG, CSF

0050394, HER1/2CSF	
Specimen Requirements:	
Patient Preparation:	
Collect:	CSF.
Specimen Preparation:	Transfer 1 mL CSF to an ARUP <u>standard transport</u> <u>tube.Standard-Transport Tube.</u> (Min: 0.5 mL)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Specimens types other than CSF. Contaminated, heat- inactivated <u>, icteric,</u> or hemolyzed specimens.
Remarks:	Indicate source on test request form.
Stability:	Ambient: 8 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Semi-Quantitative Chemiluminescent Immunoassay
Performed:	Sun-Sat
Reported:	Within 24 hours
Note:	
CPT Codes:	86694
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.

Interpretive Data:

The detection of antibodies to herpes simplex virus in CSF may indicate central nervous system infection. However, consideration must be given to possible contamination by blood or transfer of serum antibodies across the blood-brain barrier.

Fourfold or greater rise in CSF antibodies to herpes on specimens at least 4 weeks apart are found in 74-94% of patients with herpes encephalitis. Specificity of the test based on a single CSF testing is not established. Presently PCR is the primary means of establishing a diagnosis of herpes encephalitis-

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.



False positive results are possible. Consider additional testing for HSV-2, particularly if the result for HSV-2 is <u>less than or equal to</u> = 3.0 IV.

0.89 IV or less	Negative - No significant level of detectable HSV IgG antibody.	
0.90-1.09 IV	Equivocal - Questionable presence of IgG antibodies. Repeat testing in 10-14 days may be helpful.	
1.10 IV or greater	Positive - IgG antibody to HSV detected which may indicate a current or past HSV infection.	



Toxoplasma 0050521, TOX	gondii Antibodies, O PAN	IgG and IgM
Specimen Req	uirements:	
Patient Pre	paration:	
Collect:		Serum <u>separator tube</u> Separator Tube (SST).
Specimen F	Preparation:	Allow specimen to clot completely at room temperature. Separate from cells ASAP or within 2 hours of collection. Transfer 2 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.5 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of the acute specimens.
Transport T	emperature:	Refrigerated.
Unacceptak	ole Conditions:	Plasma or urine. Contaminated, heat-inactivated <u>, icteric,</u> or grossly hemolyzed specimens.
Remarks:		Label specimens plainly as "acute" or "convalescent."
Stability:		After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)
Methodology:		Semi-Quantitative Chemiluminescent Immunoassay (CLIA)
Performed:		Sun-Sat
Reported:		Within 24 hours
Note:		This test should not be used for blood donor screening, associated re-entry protocols, or for screening <u>human cell,</u> <u>tissues,Human Cell, Tissues</u> and <u>c</u> Cellular <u>-</u> and <u>tissue-based</u> <u>productsTissue-Based Products</u> (HCT/P).
CPT Codes:		86777; 86778
New York DOH	Approval Status:	This test is New York DOH approved.
Interpretive Da	ita:	
Component	Interpretation	
Toxoplasma gondii Antibody, IgG	7.1 IU/mL or less: Not Detected. 7.2-8.7 IU/mL: Indeterminate: Repeat testing in	



1

10-14	days may
be hel	pful. 8.8
IU/mL	or greater:
Detec	ted.
boxoplasma ondii Antibody, M Detec AU/m Indete Repea 10-14 be hel AU/m Detec Signif of Tox gondii antibc and m a curr recent Howe levels antibc occas persis than 1	I/mL or lot ted. 8.0-9.9 L: rminate:

Test Number	Components	Reference Interval
	Toxoplasma gondii Ab, IgM	7.9 AU/mL or less
	<u>Toxoplasma gondii Ab, IgG</u>	<u><=7.1</u>

Rubella Antibody, IgM	
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum <u>separator tube</u> Separator Tube (SST).
Specimen Preparation:	Allow specimen to clot completely at room temperature. Separate from cells ASAP or within 2 hours of collection. Transfer 1 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.5 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of acute specimens.
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Contaminated, heat-inactivated, <u>icteric, or</u> grossly hemolyzed specimens.
Remarks:	Label specimens plainly as "acute" or "convalescent."-
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (Avoid repeated freeze/thaw cycles)
Methodology:	Semi-Quantitative Chemiluminescent Immunoassay (CLIA)
Performed:	Sun-Sat
Reported:	Within 24 hours
Note:	
CPT Codes:	86762
New York DOH Approval Status:	This test is New York DOH approved.

Interpretive Data:

Testing immediately post-exposure is of no value without a later convalescent specimen. While the presence of IgM antibodies suggests current or recent infection, low levels of IgM antibodies may occasionally persist for more than 12 months post-infection or immunization.

The magnitude of the measured result is not indicative of the amount of antibody present. Reference Interval:



19.9 AU/mL or less:	Not Detected.
20.0 - 24.9 AU/mL:	Indeterminate - Repeat testing in 10-14 days may be helpful.
25.0 AU/mL or greater:	Detected - IgM antibody to rubella detected, which may indicate a current or recent infection or immunization.



Rubella Antibodies, IgG and IgI 0050552, RUBE G/M	N
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube.
Specimen Preparation:	Allow specimen to clot completely at room temperature. Separate serum from cells ASAP or within 2 hours of collection. Transfer 2 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.5 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of the acute specimens. Mark specimens plainly as "acute" or "convalescent."
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Plasma or urine. Contaminated, heat-inactivated, <u>icteric, or</u> hemolyzed specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)
Methodology:	Semi-Quantitative Chemiluminescent Immunoassay (CLIA)
Performed:	Sun-Sat
Reported:	Within 24 hours
Note:	
CPT Codes:	86762 x2
New York DOH Approval Status:	This test is New York DOH approved.

Interpretive Data:

Testing immediately post-exposure is of no value without a later convalescent specimen. While the presence of IgM antibodies suggests current or recent infection, low levels of IgM antibodies may occasionally persist for more than 12 months post-infection or immunization.

The best evidence for current infection is a significant change on two appropriately timed specimens, where both tests are done in the same laboratory at the same time.

The magnitude of the measured result is not indicative of the amount of antibody present.



Component	Reference Interval
Rubella Antibody, IgG	Less than 9 IU/mL: Not Detected. 9-9.9 IU/mL: Indeterminate: Repeat testing in 10-14 days may be helpful. 10 IU/mL or greater: Detected.
Rubella Antibody, IgM	19.9 AU/mL or less: Not Detected. 20.0 - 24.9 AU/mL: Indeterminate: Repeat testing in 10-14 days may be helpful. 25.0 AU/mL or greater: Detected: IgM antibody to rubella detected, which may indicate a current or recent infection or immunization.

Test Number	Components	Reference Interval
	Rubella Antibody IgM	19.9 AU/mL or less
	Rubella Antibody IgG	< <u>=8.9</u>



Cytomegalovirus Antibody, IgM 0050553, CMV IGM	1
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum <u>separator tube</u> Separator Tube (SST).
Specimen Preparation:	Allow specimen to clot completely at room temperature. Separate from cells ASAP or within 2 hours of collection. Transfer 1 mL serum to an ARUP Standard Transport Tube. (Min: 0.5 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of the acute specimens.
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Contaminated, heat-inactivated <u>, icteric,</u> or grossly hemolyzed specimens.
Remarks:	Label specimens plainly as "acute" or "convalescent."-
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (Avoid repeated freeze/thaw cycles)
Methodology:	Semi-Quantitative Chemiluminescent Immunoassay (CLIA)
Performed:	Sun-Sat
Reported:	Within 24 hours
Note:	
CPT Codes:	86645
New York DOH Approval Status:	This test is New York DOH approved.

Interpretive Data:

A negative result does not rule out primary infection, please correlate clinically. CMV serology is not useful for the evaluation of active or reactivated infection in immunocompromised patients. Molecular diagnostic tests (i.e., PCR) are preferred in these cases.

This test should not be used for blood donor screening, associated <u>reentry</u> protocols, or for screening <u>human cell, tissues, Human Cell, Tissues</u> and <u>c</u>Cellular_ and <u>tissue-based</u> <u>productsTissue-Based Products</u> (HCT/P).



29.9 AU/mL or less:	Not Detected.
30.0-34.9 AU/mL:	Indeterminate - Repeat testing in 10-14 days may be helpful.
35.0 AU/mL or greater:	Detected - IgM antibody to CMV detected, which may indicate a current or recent infection. However, low levels of IgM antibodies may occasionally persist for more than 12 months post-infection.



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TEST CHANGE

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l oxoplasma gondii Antibody, lo 0050557, TOXEIGM	gM
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum Separator Tube (SST).
Specimen Preparation:	Allow specimen to clot completely at room temperature. Separate from cells ASAP or within 2 hours of collection. Transfer 1 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.5 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of the acute specimens.
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Contaminated, heat-inactivated, <u>icteric, or</u> grossly hemolyzed specimens.
Remarks:	Label specimens plainly as "acute" or "convalescent."
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (Avoid repeated freeze/thaw cycles)
Methodology:	Semi-Quantitative Chemiluminescent Immunoassay (CLIA)
Performed:	Sun-Sat
Reported:	Within 24 hours
Note:	
CPT Codes:	86778
New York DOH Approval Status:	This test is New York DOH approved.

Interpretive Data:

This test is performed using the DiaSorin LIAISON. As suggested by the CDC, any indeterminate or detected *Toxoplasma gondii* IgM result should be retested in parallel with a specimen collected 1-3 weeks later. Further confirmation may be necessary using a different test from another reference laboratory specializing in toxoplasmosis testing where an IgM ELISA should be ordered. Caution should be exercised in the use of IgM antibody levels in prenatal screening. Any *Toxoplasma gondii* IgM in pregnant patients that have also been confirmed by a second reference laboratory should be evaluated by amniocentesis and PCR testing for *Toxoplasma gondii*.

For male and <u>nonpregnantnon-pregnant</u> female patients with indeterminate or detected



Toxoplasma gondii IgM results, PCR may also be useful if a specimen can be collected from an affected body site.

This test should not be used for blood donor screening, associated re-entry protocols, or for screening <u>human cell, tissues</u>, <u>Human Cell, Tissues</u> and <u>c</u>Cellular_ and <u>tissue-based</u> <u>productsTissue-Based Products</u> (HCT/P).

For additional information, refer to the CDC website: www.cdc.gov/parasites/toxoplasmosis/health_professionals/index.html.

The magnitude of the measured result is not indicative of the amount of antibody present.

7.9 AU/mL or less:	Not Detected.
8.0-9.9 AU/mL:	Indeterminate - Repeat testing in 10-14 days may be helpful.
10.0 AU/mL or greater:	Detected - Significant level of Toxoplasma gondii lgM antibody detected and may indicate a current or recent infection. However, low levels of lgM antibodies may occasionally persist for more than 12 months post-infection.



Saccharomyces cerevisiae Antibodies, IgG & IgA

0050564, SC PAN	
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube.
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.3 mL)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Contaminated, heat-inactivated, hemolyzed, or severely lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)
Methodology:	Semi-Quantitative Enzyme-Linked Immunosorbent Assay <u>(ELISA)</u>
Performed:	<u>Mon, Wed, Fri</u> Sun-Sat
Reported:	1-2 <u>-4</u> days
Note:	This test may be a useful tool for distinguishing ulcerative colitis (UC) from Crohn disease (CD) in patients with suspected inflammatory bowel disease.
CPT Codes:	86671 x2
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data	

Saccharomyces cerevisiae IgG antibodies are found in 60-70% of Crohn disease (CD) patients and 10-15% of ulcerative colitis (UC) patients. *Saccharomyces cerevisiae* IgA antibodies are found in about 35% of CD patients but less than 1% in UC patients. Detection of both *Saccharomyces* IgG and IgA antibodies in the same serum specimen is highly specific for CD.



Test Number	Components	Reference Inte	rval
	S. cerevisiae Antibody, IgG		
		20.0 Units or less	Negative
		20.1-24.9 Units	Equivocal
		25.0 Units or greater	Positive
	S. cerevisiae Antibody, IgA		
		20.0 Units or less	Negative
		20.1-24.9 Units	Equivocal
		25.0 Units or greater	Positive



Gastric Parietal Cell Antibody, IgG		
0050596, PCA		
Specimen Requirements:		
Patient Preparation:		
Collect:	Serum separator tube.	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.25 mL)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Urine or plasma. Contaminated, heat-inactivated, grossly hemolyzed, or severely lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)	
Methodology:	Semi-Quantitative Enzyme-Linked Immunosorbent Assay	
Performed:	<u>Sun, Tue, Thu</u> Mon, Wed-Sat	
Reported:	1-3 days	
Note:	Most patients with pernicious anemia have parietal cell antibodies. However, the fact that such antibodies are found with increased frequency in unaffected family members, as well as in patients with other autoimmune diseases, suggests these antibodies do not cause disease by themselves.	
CPT Codes:	83516	
New York DOH Approval Status:	This test is New York DOH approved.	

Interpretive Data:

In the context of vitamin B12 deficiency, the presence of gastric parietal cell antibodies (PCA) and/or intrinsic factor antibodies in association with macrocytic anemia is considered diagnostic for pernicious anemia (PA). However, the presence of gastric PCAs alone is not specific for PA. Gastric PCAs may occur with increased frequency in unaffected family members, a small percentage of healthy individuals, and patients with other autoimmune diseases, such as autoimmune thyroiditis.



Component	Interpretation
Parietal Cell Antibody, IgG	0.0-20.0 Units Negative 20.1- 24.9 Units Equivocal 25.0 Units or greater Positive

Test Number	Components	Reference Interval
	Gastric Parietal Cell Antibody, IgG	24.9 Units or less



Epstein-Barr 0050600, EBV	Virus Antibody Pa PAN	anel I
Specimen Req	uirements:	
Patient Prep	paration:	
Collect:		Serum separator tube (SST).
Specimen P	reparation:	Allow specimen to clot completely at room temperature. Separate from cells ASAP or within 2 hours of collection. Transport 2 mL serum to an ARUP standard transport tube. (Min: 0.5 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of acute specimens.
Transport T	emperature:	Refrigerated.
Unacceptab	le Conditions:	Contaminated, heat-inactivated, icteric, or grossly hemolyzed specimens.
Remarks:		Label specimens plainly as "acute" or "convalescent."
Stability:		After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (Avoid repeated freeze/thaw cycles)
Methodology:		Semi-Quantitative Chemiluminescent Immunoassay
Performed:		Sun-Sat
Reported:		1-2 days
Note:		
CPT Codes:		86665 x2; 86664; 86663
New York DOH	Approval Status:	This test is New York DOH approved.
Interpretive Da	ta:	
Component	Interpretation	
Epstein-Barr Virus Antibody to Viral Capsid Antigen, IgG	17.9 U/mL or less: Not Detected 18.0- 21.9 U/mL: Indeterminate. Repeat testing in 10-14 days may be helpful. 22.0 U/mL or greater: Detected	



Epstein-Barr Virus Antibody to Viral Capsid Antigen, gM	35.9 U/mL or less: Not Detected 36.0- 43.9 U/mL: Indeterminate. Repeat testing in 10-14 days may be helpful. 44.0 U/mL or greater:
Epstein-Barr Virus	Detected 17.9 U/mL or
Antibody to Nuclear Antigen, gG	less: Not Detected 18.0- 21.9 U/mL: Indeterminate. Repeat testing in 10-14 days may be helpful. 22.0 U/mL or greater: Detected
Epstein-Barr Virus Antibody to Early D Antigen (EA-D), gG	8.9 U/mL or less: Not Detected 9.0- 10.9 U/mL: Indeterminate - Repeat testing in 10-14 days may be helpful. 11.0 U/mL or greater: Detected

Test Number	Components	Reference Interval
	EBV Antibody to Early (D) Antigen IgG	<u><=8</u> 10.9 U/mL or less
	EBV Antibody to Viral Capsid Antigen IgG	< <u>=17</u> 21.9 U/mL or less
	EBV Antibody to Viral Capsid Antigen IgM	< <u>=35</u> 43.9 U/mL or less
	EBV Antibody to Nuclear Antigen IgG	< <u>=17</u> 21.9 U/mL or less



Epstein-Barr 0050602, EBV	Virus Antibody P PAN 2	anel II
Specimen Req	uirements:	
Patient Prep	paration:	
Collect:		Serum separator tube (SST).
Specimen P	reparation:	Allow specimen to clot completely at room temperature. Separate from cells ASAP or within 2 hours of collection. Transfer 2 mL serum to an ARUP standard transport tube. (Min: 0.5 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of the acute specimens.
Transport T	emperature:	Refrigerated.
Unacceptab	le Conditions:	Contaminated, heat-inactivated, icteric, or grossly hemolyzed specimens.
Remarks:		Label specimens plainly as "acute" or "convalescent."
Stability:		After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (Avoid repeated freeze/thaw cycles).
Methodology:		Semi-Quantitative Chemiluminescent Immunoassay
Performed:		Sun-Sat
Reported:		1-2 days
Note:		
CPT Codes:		86665 x2
New York DOH	Approval Status:	This test is New York DOH approved.
Interpretive Da	ta:	
Component	Interpretation	
Epstein-Barr Virus Antibody to Viral Capsid Antigen, IgG	17.9 U/mL or less: Not Detected 18.0- 21.9 U/mL: Indeterminate. Repeat testing in 10-14 days may be helpful. 22.0 U/mL or greater: Detected	



Test Number	Components	Reference Interval
	EBV Antibody to Viral Capsid Antigen IgG	<u><=17</u> 21.9 U/mL or less
	EBV Antibody to Viral Capsid Antigen IgM	< <u>=35</u> 43.9 U/mL or less



TEST CHANGE

Cytomegalovir 0050622, CMV F	us Antibodies, Ig PAN	JG and IgM
Specimen Requi	irements:	
Patient Prepa	aration:	
Collect:		Serum <u>separator tube</u> Separator Tube (SST).
Specimen Pre	eparation:	Allow specimen to clot completely at room temperature. Separate from cells ASAP or within 2 hours of collection. Transfer 2 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.5 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of the acute specimens.
Transport Ter	mperature:	Refrigerated.
Unacceptable	e Conditions:	Plasma or urine. Contaminated, hemolyzed, <u>icteric,</u> or heat- inactivated specimens.
Remarks:		Label specimens plainly as "acute" or "convalescent."
Stability:		After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)
Methodology:		Semi-Quantitative Chemiluminescent Immunoassay (CLIA)
Performed:		Sun-Sat
Reported:		1-2 days
Note:		This test should not be used for blood donor screening, associated re-entry protocols, or for screening <u>human cell,</u> <u>tissues,Human Cell, Tissues</u> and <u>c</u> Cellular <u>-</u> and <u>tissue-based</u> <u>productsTissue-Based Products</u> (HCT/P).
CPT Codes:		86644; 86645
New York DOH A	Approval Status:	This test is New York DOH approved.
Interpretive Data	a:	
Component Ir	nterpretation	
Cytomegalovirus 0 Antibody, IgG le D 0	0.59 U/mL or ess: Not Detected. 0.60- 0.69 U/mL: ndeterminate:	



	Repeat testing in 10-14 days may be helpful. 0.70 U/mL or greater: Detected.
Cytomegaloviru Antibody, IgM	IS 29.9 AU/mL or less: Not Detected. 30.0- 34.9 AU/mL: Indeterminate: Repeat testing in 10-14 days may be helpful. 35.0 AU/mL or greater: Detected: IgM antibody to CMV detected, which may indicate a current or recent infection. However, low levels of IgM antibodies may occasionally persist for more than 12 months post-infection.

Reference Interval:

Test Number	Components	Reference Interval
	CMV Antibody Ig <u>G</u> M	<u><=0.59</u> 29.9 AU/mL or less



Endomysial Antibody, IgA by IFA	Α
0050736, EMAR TITER	
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube <u>or red tube</u> .
Specimen Preparation:	Separate from cells ASAP or within 2 hours of collection. Transfer 1 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.15 mL)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Plasma. Severely lipemic, contaminated, or hemolyzed specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid freeze/thaw cycles)
Methodology:	Semi-Quantitative Indirect Fluorescent Antibody
Performed:	Mon-Fri
Reported:	1-5 days
Note:	
CPT Codes:	86231
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	
The endomysial antigen has been in transglutaminase.	dentified as the protein cross-linking enzyme known as tissue
Reference Interval:	
Less than 1:10	



TEST CHANGE

Toxoplasma gondii Antibody, Ig 0050770, TOXEIGG	gG
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum Separator Tube (SST).
Specimen Preparation:	Allow specimen to clot completely at room temperature. Separate from cells ASAP or within 2 hours of collection. Transfer 1 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.5 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of the acute specimens.
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Contaminated, heat-inactivated, <u>icteric, or</u> grossly hemolyzed specimens.
Remarks:	Label specimens plainly as "acute" or "convalescent."-
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (Avoid repeated freeze/thaw cycles)
Methodology:	Semi-quantitative Chemiluminescent Immunoassay (CLIA)
Performed:	Sun-Sat
Reported:	Within 24 hours
Note:	
CPT Codes:	86777
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	
The best evidence for current infect	ction is a significant change on two appropriately timed

The magnitude of the measured result is not indicative of the amount of antibody present.

This test should not be used for blood donor screening, associated re-entry protocols, or for screening human cell, tissues, Human Cell, Tissues and cCellular- and tissue-based products Tissue-Based Products (HCT/P).



Effective March 3, 2014

7.1 IU/mL or less:	Not Detected.
7.2-8.7 IU/mL:	Indeterminate - Repeat testing in 10-14 days may be helpful.
8.8 IU/mL or greater:	Detected.

Rubella Antibody, IgG 0050771, RUBEIGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Serum Separator Tube (SST).	
Specimen Preparation:	Allow specimen to clot completely at room temperature. Separate from cells ASAP or within 2 hours of collection. Transfer 1 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.5 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of the acute specimens.	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Contaminated, heat-inactivated, <u>icteric,</u> or grossly hemolyzed specimens.	
Remarks:	Label specimens plainly as "acute" or "convalescent."	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (Avoid repeated freeze/thaw cycles)	
Methodology:	Semi-quantitative Chemiluminescent Immunoassay (CLIA)	
Performed:	Sun-Sat	
Reported:	Within 24 hours	
Note:		
CPT Codes:	86762	
New York DOH Approval Status:	This test is New York DOH approved.	
Interpretive Data:		
The best evidence for current infection is a significant change on two appropriately timed specimens, where both tests are done in the same laboratory at the same time.		

The magnitude of the measured result is not indicative of the amount of antibody present. Reference Interval:



Less than 9 IU/mL:	Not Detected.
9-9.9 IU/mL:	Indeterminate - Repeat testing in 10-14 days may be helpful.
10 IU/mL or greater:	Detected.

TORCH Antib 0050772, TOR	odies, IgG CH IGG	
Specimen Req	uirements:	
Patient Pre	paration:	
Collect:		Serum Separator Tube (SST).
Specimen Preparation:		Allow specimen to clot completely at room temperature. Separate from cells ASAP or within 2 hours of collection. Transfer 2 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 1 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of the acute specimens.
Transport Temperature:		Refrigerated.
Unacceptable Conditions:		Refer to individual components.
Remarks:		Mark specimens plainly as "acute" or "convalescent."
Stability:		After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)
Methodology:		Semi-Quantitative Chemiluminescent Immunoassay (CLIA)
Performed:		Sun-Sat
Reported:		Within 24 hours
Note:		This test should not be used for blood donor screening, associated re-entry protocols, or for screening <u>human cell</u> , <u>tissues</u> , <u>Human Cell</u> , <u>Tissues</u> and <u>c</u> Cellular <u>-</u> and <u>tissue-based</u> <u>productsTissue-Based Products</u> (HCT/P).
CPT Codes:		86644; 86694; 86762; 86777
New York DOH	Approval Status:	This test is New York DOH approved.
Interpretive Da	ata:	
Component	Interpretation	
Cytomegalovirus Antibody, IgG	0.59 U/mL or less: Not Detected. 0.60- 0.69 U/mL: Indeterminate: Repeat testing in	



	10-14 days may be helpful. 0.70 U/mL or greater: Detected.
Herpes Simplex Virus Type 1 and/or 2 Antibodies, IgG	0.89 IV or less: Not Detected. 0.90-1.09 IV: Indeterminate: Repeat testing in 10-14 days may be helpful. 1.10 IV or greater: Detected.
Rubella Antibody, IgG	Less than 9 IU/mL: Not Detected. 9-9.9 IU/mL: Indeterminate: Repeat testing in 10-14 days may be helpful. 10 IU/mL or greater: Detected.
Toxoplasma gondii Antibody, IgG	7.1 IU/mL or less: Not Detected. 7.2-8.7 IU/mL: Indeterminate: Repeat testing in 10-14 days may be helpful. 8.8 IU/mL or greater: Detected.



Treponema pallidum Antibody by TP-PA 0050777, MHA				
Specimen Requirements:				
Patient Preparation:				
Collect:	Serum separator tube or plasma separator tube.			
Specimen Preparation:	Separate serum or plasma from cells ASAP or within 2 hours of collection. Transfer 1 mL serum or plasma to an ARUP standard transport tube. (Min: 0.4 mL)			
Transport Temperature:	Refrigerated.			
Unacceptable Conditions:	CSF or other body fluids.			
Remarks:				
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)			
Methodology:	Qualitative Particle Agglutination			
Performed:	<u>Sun-Sat</u> Mon-Fri			
Reported:	1-4 days			
Note:	TP-PA is a helpful diagnostic aid for the patient with a reactive reagin test, but presents with atypical signs of primary, secondary, or late syphilis. TP-PA compares favorably with the FTA test, but appears slightly less sensitive in cases of untreated early primary syphilis. In late syphilis, the agreement with FTA is 99%. VDRL is the preferred test for cerebrospinal fluid. Treponemal tests (TP-PA or FTA) are not recommended for CSF. FTAs on CSF may be tested, but TP-PA cannot be tested on CSF.			
CPT Codes:	86780			
New York DOH Approval Status:	This test is New York DOH approved.			
Interpretive Data:				
Reference Interval:				
Nonreactive				




Diphtheria, Tetanus, and H. Influenzae b Antibodies, IgG

0050779, DTH	
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube <u>or red tube</u> .
Specimen Preparation:	Transfer 1.5 mL serum to an ARUP <u>standard transport</u> <u>tubeStandard Transport Tube</u> . (Min: 0.45 mL) Acute and convalescent specimens must be labeled as such. Clearly mark specimens as "Pre-Vaccine" or "Post-Vaccine."", Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of the acute specimens.
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Plasma or other body fluids. Contaminated, hemolyzed, or severely lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)
Methodology:	Quantitative Multiplex Bead Assay
Performed:	Sun-Sat
Reported:	1-3 days
Note:	
CPT Codes:	86317 x3

New York DOH Approval Status: This test is New York DOH approved.

Interpretive Data:

Responder status is determined according to the ratio of a one-month, post-vaccination sample to pre-vaccination concentration of IgG antibodies as follows:

Diphtheria and tetanus:

1. If the post-vaccination concentration is less than 1.0 IU/mL, the patient is considered a nonresponder.

2. If the post-vaccination concentration is greater than or equal to 1.0 IU/mL, a patient with a ratio of less than 1.5 is a nonresponder, and a ratio of 1.5 to less than 3.0 is a weak responder, and a ratio of 3.0 or greater is a good responder.

3. If the pre-vaccination concentration is greater than 1.0 IU/mL, it may be difficult to assess the



response based on a ratio alone. A post-vaccination concentration above 2.5 IU in this case is usually adequate.

Haemophilus influenza B:

1. If the post-vaccination concentration is < 3.0 μ g/mL, the patient is considered to be a nonresponder.

2. If the post-vaccination concentration is $3.0 \frac{\mu g/mL}{m}$, a patient with a ratio of 4 is a good responder, a ratio of 2-4 is weak responder, and a ratio of < 2 is considered a nonresponder.

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Diphtheria and tetanus: Antibody concentration of > 0.1 IU/mL is usually considered protective for diphtheria or tetanus.

Haemophilus influenzae type B:

< 1.0 μ g/mL = Antibody concentration not protective.

> 1.0 μ g/mL = Antibody to *H. influenzae* type B detected. Suggestive of protection.



SSA-60 (Ro60)

lgG

(ENA) Antibody,

SSB (La) (ENA)

Antibody, IgG

29 AU/mL or Less

29 AU/mL or less

Negative 30-40 AU/mL Equivocal 41 AU/mL or

Negative 30-40 AU/mL Equivocal

41 AU/mL or greater Positive

TEST CHAN	GE	
Extractable N	luclear Antigen A	ntibodies (SSA 52, SSA 60, and SSB)
0050791, SSA	/SSB	
Specimen Req	uirements:	
Patient Pre	paration:	
Collect:		Serum separator tube <u>or red tube</u> .
Specimen F	reparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 1 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.3 mL)
Transport T	emperature:	Refrigerated.
Unacceptab	le Conditions:	Plasma or other body fluids. Contaminated, hemolyzed, or severely lipemic specimens.
Remarks:		
Stability:		After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)
Methodology:		Semi-Quantitative Multiplex Bead Assay
Performed:		Sun-Sat
Reported:		1-2 days
Note:		
CPT Codes:		86235 x3
New York DOH	Approval Status:	This test is New York DOH approved.
Interpretive Data:		
Component	Interpretation	
SSA-52 (Ro52) (ENA) Antibody, IgG	29 AU/mL or Less Negative 30-40 AU/mL Equivocal 41 AU/mL or greater Positive	



greater Positive

Reference Interval:

Test Number	Components	Reference Interval
	SSA-52 (Ro52) (ENA) Antibody, IgG	40 AU/mL or less
	SSB (La) (ENA) Antibody, IgG	40 AU/mL or less
	SSA-60 (Ro60) (ENA) Antibody, IgG	40 AU/mL or less



Phosphatidylserine Antibodies, IgG, IgM, and IgA

0050905, PHOS AB	
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube.
Specimen Preparation:	Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.</u> Standard Transport Tube. (Min: 0.25 mL)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Contaminated, heat-inactivated, hemolyzed, or severely lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 month
Methodology:	Semi-Quantitative Enzyme-Linked Immunosorbent Assay
Performed:	<u>Mon</u> Sun, Tue, Wed, Fri , Sat
Reported:	1-4 days
Note:	
CPT Codes:	86148 x3

New York DOH Approval Status: This test is New York DOH approved.

Interpretive Data:

IgG and/or IgM antibodies to phosphatidylserine (aPS) may be associated with a positive test for anti-cardiolipin autoantibodies (aCL) and risk for obstetric antiphospholipid syndrome (APS). Strong clinical correlation is recommended in the absence of lupus anticoagulant, IgG and/or IgM cardiolipin and/or beta2 glycoprotein antibodies.

Isolated presence of IgM or IgG antibodies to aPS may have questionable clinical significance for APS and/or SLE.

If results are positive, repeat testing with two or more specimens drawn at least 12 weeks apart to demonstrate persistence of antibodies.

Results should not be used alone for diagnosis and must be interpreted in light of APS-specific clinical manifestations and/or other criteria phospholipid antibody tests.



Reference Interval:

Test Number	Components	Reference Interval
	Phosphatidylserine Antibody IgG	Less than 16 GPS
	Phosphatidylserine Antibody IgM	Less than 22 MPS
	Phosphatidylserine Antibody IgA	Less than 20 APS



TEST CHANGE

Smooth Muscle Antibody, IgG Titer		
0051244, *ASM TITER		
Specimen Requirements:		
Patient Preparation:		
Collect:	Serum separator tube <u>or red tube.</u>	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 1 mL serum to an ARUP <u>standard transport</u> <u>tube.</u> Standard Transport Tube. (Min: 0.15 mL)	
Transport Temperature:	Refrigerated	
Unacceptable Conditions:	Contaminated, hemolyzed, or severly lipemic specimens.	
Remarks:		
Stability:	Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Semi-Quantitative Indirect Fluorescent Antibody	
Performed:	Sun-Sat	
Reported:	1-3 days	
Note:		
CPT Codes:	86256	

New York DOH Approval Status: This test is New York DOH approved.

Interpretive Data:

Component
Smooth Muscle Antibody, IgG Titer

Reference Interval:



Test Number	Components	Reference Interval
	Smooth Muscle Ab, IgG Titer	Less than 1:20



Galactosemia (GALT) 9 Mutations, Fetal 0051270, GALTDNA FE		
Specimen Requirements:		
Patient Preparation:		
Collect:	Fetal: Cultured Amniocytes, Cultured CVS , or Direct Amniotic Fluid (direct) AND Maternal Whole Blood Specimen: Lavender (EDTA), pink (K2EDTA), or yellow (ACD solution A or B).	
Specimen Preparation:	Cultured Amniocytes or Cultured CVS: Transfer cultured amniocytes or cultured CVS to two T-25 flasks at 80 percent confluence. (Min: one T-25 flask at 80 percent confluence)). Backup cultures must be retained at the client's institution until testing is complete. If ARUP receives a sample below the minimum confluence, Cytogenetics Grow and Send (ARUP test code 0040182) will be added on by ARUP, and additional charges will apply. If clients are unable to culture specimens, Cytogenetics Grow and Send should be added to the initial order. <u>Direct Amniotic Fluid: 10 mL</u> Maternal Whole Blood Specimen: 2 mL whole blood (Min: 1 mL).	
Transport Temperature:	Cultured Amniocytes or Cultured CVS: CRITICAL ROOM TEMPERATURE. Must be received within 48 hours of collection due to viability of cells. <u>Direct Amniotic Fluid: Ship room</u> <u>temperature.</u> Maternal Whole Blood Specimen: Room temperature	
Unacceptable Conditions:		
Remarks:		
Stability:	Cultured Amniocytes or Cultured CVS: Room temperature: 48 hours; Refrigerated: Unacceptable; Frozen: Unacceptable <u>Direct Amniotic Fluid: Room temperature: 48 hours;</u> <u>Refrigerated: 72 hours; Frozen: Unacceptable</u> _Maternal Whole Blood Specimen: Room temperature: 7 days; Refrigerated: 1 month; Frozen: Unacceptable	
Methodology:	Polymerase Chain Reaction (PCR <u>)</u> Extensions	
Performed:	Sun-Sat	
Reported:	5-7 days	
Note:	This test is offered to individuals with a known familial mutation(s).	



CPT Codes:

81401; 81265 Fetal Cell Contamination (FCC)

New York DOH Approval Status: This test is New York DOH approved.

Interpretive Data:

Refer to report. Background Information for Galactosemia (GALT) 9 Mutations:

Characteristics: Affected infants present at 3-14 days old with poor feeding, vomiting, diarrhea, jaundice, lethargy progressing to coma, and abdominal distension with hepatomegaly usually followed by progressive liver failure. Patients with galactosemia are also at increased risk for E. coli or other Gram negative neonatal sepsis. Diagnosis is made by measuring GALT enzyme activity in red blood cells.

Incidence: Approximately 1 in 30,000 to 60,000 for classic galactosemia in Caucasians; varies in other populations.

Inheritance: Autosomal recessive.

Penetrance: 100 percent for severe GALT mutations.

Cause: Mutations in the GALT gene.

Mutations Tested: Seven GALT gene mutations (Q188R, S135L, K285N, T138M, L195P, Y209C, and IVS2-2 A>G) and two variants (N314D and L218L).

Clinical Sensitivity: Approaches 80 percent in Caucasians but reduced in other ethnic groups. Methodology: Polymerase chain reaction followed by single nucleotide extension (SNE) and capillary electrophoresis.

Analytical Sensitivity: 99 percent for mutations listed.

Limitations: GALT gene mutations, other than the 9 targeted, will not be detected. Diagnostic errors can occur due to rare sequence variations.

Reference Interval:

By report



Prothrombin Antibody, IgG 0051302, PROTHROM G	
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube (SST) OR lt. blue (sodium citrate)
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum (Min: 0.3 mL) OR 0.5 mL citrate plasma (Min: 0.3 mL) to an ARUP standard transport tube.
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Contaminated, heat-inactivated, clots, fibrin, gross red blood cells, severely lipemic, severely hemolyzed, or severely icteric specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Semi-Quantitative Enzyme-Linked Immunosorbent Assay (ELISA)
Performed:	<u>FriMon</u>
Reported:	1-8 days
Note:	
CPT Codes:	83516
New York DOH Approval Status:	This test is New York DOH approved.

Interpretive Data:

IgG antibodies to prothrombin may be a risk factor for either venous or arterial thrombosis in antiphospholipid syndrome (APS). Strong clinical correlation is recommended in the absence of lupus anticoagulant, IgG and/or IgM cardiolipin and/or beta2 glycoprotein antibodies.

If results are positive, repeat testing with two or more specimens drawn at least 12 weeks apart to demonstrate persistence of antibodies.

Results should not be used alone for diagnosis and must be interpreted in light of APS-specific clinical manifestations and/or other criteria phospholipid antibody tests. Reference Interval:



Effective Date: July 21, 2025

Effective 5/21/2018 Less than 20 Units



Ashkenazi Jewish Diseases, 16 Genes

0051	415	A.JP
0001	TIU	,

Specimen Requirements:	
Patient Preparation:	
Collect:	 Whole blood: Lavender (EDTA), pink (K2EDTA), or yellow (ACD solution A or B). Whole blood: Lavender (EDTA), pink (K 2 EDTA), or yellow (ACD solution A or B). Fetal specimens: Cultured amniocytes: Two T-25 flasks at 80 percent confluency. OR cultured CVS: Two T-25 flasks at 80 percent confluency. If the client is unable to culture, order ARUP test Cytogenetics Grow and Send (test code 0040182) in addition to this test and ARUP will culture upon receipt (culturing fees will apply). If you have any questions, contact ARUP's Genetics Processing at 800-522-2787 ext. 3301. AND maternal whole blood: lavender (K2 or K3EDTA), pink (K2EDTA), or yellow (ACD solution A or B).
Specimen Preparation:	Whole blood: Transport 3 mL whole blood. (Min: 1 mL)—Fetal Specimens: Cultured amniocytes OR cultured CVS: Transport two T-25 flasks at 80 percent confluency filled with culture media. Backup cultures must be retained at the client's institution until testing is complete. AND maternal whole blood: transport 2 mL whole blood (min: 1 mL).
Transport Temperature:	Whole blood: Refrigerated. Whole blood: Refrigerated. Fetal specimens: Cultured amniocytes OR cultured CVS: CRITICAL ROOM TEMPERATURE. Must be received within 48 hours of shipment due to lability of cells AND maternal whole blood: room temperature. Also acceptable: refrigerated.
Unacceptable Conditions:	Plasma or serum. Specimens collected in sodium heparin or lithium heparin tubes. Frozen specimens in glass collection tubes.
Remarks:	
Stability:	Whole blood: Ambient: 72 hours; Refrigerated: 1 week; Frozen: unacceptable-Fetal specimens : Ambient: 48 hours; Refrigerated: Unacceptable; Frozen: Unacceptable-AND Maternal whole blood: Ambient: 72 hours; Refrigerated: 1 week. Frozen: Unacceptable.
Methodology:	Polymerase Chain Reaction (PCR)/Fluorescence Monitoring



Performed:	Varies	
Reported:	5-10 days	
Note:	Cystic fibrosis (CF) carrier testing is NOT included as part of this panel. Please order Cystic Fibrosis (CFTR) Expanded Variant Panel (ARUP test code 2013661) to assess CF carrier status. <u>Any submitted fetal specimens will have Maternal Cell</u> <u>Contamination, Fetal Sample, added on by ARUP. Additional charges will apply.</u>	
CPT Codes:	81401, 81209, 81200, 81260, 81242, 81251, 81250, 81479, 81205, 81290, 81400, 81330, 81255	
New York DOH Approval Status:	This test is New York DOH approved.	
Interpretive Data:		
Refer to report. Refer to report		
Counseling and informed consent available online.	are recommended for genetic testing. Consent forms are	
Reference Interval:		



Epstein-Barr Virus Antibody to Viral Capsid Antigen, IgG and IgA

TEST CHANGE

0051627, EBV PAN 3 Specimen Requirements: Patient Preparation: Collect: Serum separator tube (SST). **Specimen Preparation:** Allow specimen to clot completely at room temperature. Separate from cells ASAP or within 2 hours of collection. Transfer 2 mL serum to an ARUP standard transport tube. (Min: 0.5 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of the acute specimens. Transport Temperature: Refrigerated. Unacceptable Conditions: Contaminated, heat-inactivated, icteric, or grossly hemolyzed specimens. Remarks: Label specimens plainly as "acute" or "convalescent." Stability: After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 month (Avoid repeated freeze/thaw cycles). Methodology: Enzyme-Linked Immunosorbent Assay/Semi-Quantitative Chemiluminescent Immunoassay Performed: Tue **Reported:** 1-8 days Note: CPT Codes: 86665 x2 New York DOH Approval Status: This test is New York DOH approved. Interpretive Data: Component Interpretation Epstein-Barr Virus 17.9 U/mL or Antibody to Viral less: Not Capsid Antigen, Detected 18.0lgG 21.9 U/mL: Indeterminate. Repeat testing in 10-14 days may be helpful. 22.0 U/mL or greater:



	Detected
Epstein-Barr Virus Antibody to Viral Capsid Antigen, IgA	8 U or less: Not Detected 9-11 U: Indeterminate - Repeat testing in 10-14 days may be helpful. 12 U or greater: Detected

Reference Interval:

Test Number	Components	Reference Interval
	EBV Antibody to Viral Capsid Antigen IgG	<u><=17</u> 21.9 U/mL or less
	EBV Antibody To Viral Capsid Antigen IgA	8 U or less



Herpes Simplex Virus Type 1 and/or 2 Antibodies, IgG with Reflex to Type 1 and 2 Glycoprotein G-Specific Ab, IgG

0051708, HERPR PAN2

Specimen Requirements:

Patient Preparation:	
Collect:	Serum separator tube.
Specimen Preparation:	Transfer 1 mL serum to an ARUP <u>standard transport</u> <u>tube</u> <u>Standard Transport Tube</u> . (Min: 0.5 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of acute specimens. Mark specimens plainly as "acute" or "convalescent."
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Plasma or urine. Contaminated, heat-inactivated, hemolyzed <u>,</u> icteric, or severely lipemic specimens.
Remarks:	
Stability:	Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Semi-Quantitative Chemiluminescent Immunoassay
Performed:	Sun-Sat
Reported:	1-2 days
Note:	If HSV 1 and/or 2 IgG is 1.10 IV or greater, then HSV 1 gG- Specific IgG and HSV 2 gG-Specific IgG will be added. Additional charges apply.
CPT Codes:	86694; if reflexed, add 86695; 86696
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	

False positive results are possible. Consider additional testing for HSV-2, particularly if the result for HSV-2 is less than or equal to </= 3.0 IV.

Component	Interpretation
Herpes Simplex	0.89 IV or less:
Virus Type 1	Not Detected.
and/or 2	0.90-1.09 IV:
Antibodies, IgG by	Indeterminate <u>.</u>
Chemiluminescent	Repeat testing in
Immunoassay	10-14 days may



be helpful. 1.10 IV	
or greater:	
Detected.	

Reference Interval:



Effective Date: July 21, 2025

TEST CHANGE

Allergen, Insects and Venom, White-Faced Hornet IgG

00	55405, WH F IGG		
Sp	ecimen Requirements:		
	Patient Preparation:		
	Collect:	<u>Plain red or serum</u> Serum se	parator tube <u>(SST)</u> -
	Specimen Preparation:	Separate serum from cells A Transfer 0.5 mL serum to ar <u>tube.Standard Transport Tu</u> <u>"Allergen Specimen Collecti</u> <u>www.aruplab.com/testing/r</u>	ASAP or within 2 hours of collection. n ARUP <u>standard transport</u> be. (Min: 0.252 mL) <u>Refer to</u> on Instructions" at resources/specimen.
	Transport Temperature:	Refrigerated.	
	Unacceptable Conditions:	Hemolyzed, icteric, or lipem	ic specimens.
	Remarks:		
	Stability:	After separation from cells: weeks; Frozen: 1 year	Ambient: 48 hours; Refrigerated: 2
M	ethodology:	Quantitative ImmunoCAP FI	luorescent Enzyme Immunoassay
Pe	rformed:	Sun	
Re	ported:	1-8 days	
No	ote:	The units of measure mcg/r 1 mg/L = 1000 mcg/1000 m	mL and mgA/L are interchangeable. 1L
CF	PT Codes:	86001	
Ne	w York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Int	erpretive Data:		
Va an	lues less than 2.00 mcg/mL rep tibody.	resent absent or undetectable	e levels of allergen-specific IgG
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.		termined by ARUP Laboratories. It ninistration. This test was ical purposes.	
Re	ference Interval:		
Te	st Number	Components	Reference Interval

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Effective Date: July 21, 2025

Allergen, Insect, WhFaced Less than 8.47 mcg/mL Hornet Ven IgG

Inserted Cells Inserted Cells

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Effective Date: July 21, 2025

TEST CHANGE

Allergen, Insects and Venom, Yellow Jacket IgG

0055410, YEL J IGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Plain red or serumSerum separator tube (SST)	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to</u> "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.)	
Transport Temperature:	Refrigerated	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL	
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL rep antibody.	resent absent or undetectable levels of allergen-specific IgG	
This test was developed and its pe has not been cleared or approved l performed in a CLIA certified labor	rformance characteristics determined by ARUP Laboratories. It by the US Food and Drug Administration. This test was atory and is intended for clinical purposes.	
Reference Interval:		
Test Number	Components Reference Interval	

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Less than 6.26 mcg/mL

Effective Date: July 21, 2025

Allergen, Insect, YellowJacket Venom IgG Inserted Cells Inserted Cells

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Effective Date: July 21, 2025

TEST CHANGE

Allergen, Insects and Venom, Paper Wasp IgG

00	055415, PAP-W IGG		
Sp	pecimen Requirements:		
	Patient Preparation:		
	Collect:	Plain red or serumSerum se	eparator tube <u>(SST)</u>
	Specimen Preparation:	Separate serum from cells A Transfer 0.5 mL serum to an <u>tube.Standard Transport Tu</u> <u>"Allergen Specimen Collecti</u> www.aruplab.com/testing/n	ASAP or within 2 hours of collection. n ARUP <u>standard transport</u> the. (Min: 0.252 mL). <u>Refer to</u> ton Instructions" at resources/specimen.
	Transport Temperature:	Refrigerated	
	Unacceptable Conditions:	Hemolyzed, icteric, or lipem	ic specimens.
	Remarks:		
	Stability:	After separation from cells: weeks; Frozen: 1 year	Ambient: 48 hours; Refrigerated: 2
М	Methodology: Quantitative ImmunoCAP Fluorescent Enzyme Immunoass		luorescent Enzyme Immunoassay
Pe	erformed:	Sun	
Re	eported:	1-8 days	
No	ote:	The units of measure mcg/r 1 mg/L = 1000 mcg/1000 m	mL and mgA/L are interchangeable. nL
CF	PT Codes:	86001	
Ne	ew York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible	
In	terpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.			
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.		stermined by ARUP Laboratories. It ninistration. This test was ical purposes.	
Re	eference Interval:		
Te	st Number (<u>Components</u>	Reference Interval

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Less than 7.47 mcg/mL

Effective Date: July 21, 2025

Allergen, Insect, Paper Wasp Venom IgG

Inserted Cells Inserted Cells

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Allergen, Insects and Venom, Yellow Hornet IgG 0055420, YE F IGG Specimen Requirements: Patient Preparation: Collect: Plain red or serumSerum separator tube (SST) **Specimen Preparation:** Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP standard transport tube.Standard Transport Tube. (Min: 0.252 mL). Refer to "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen. Transport Temperature: Refrigerated Unacceptable Conditions: Hemolyzed, icteric, or lipemic specimens. Remarks: Stability: After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay Methodology: Performed: Sun **Reported:** 1-8 days Note: The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL CPT Codes: 86001 Specimens from New York clients will be sent out to a New New York DOH Approval Status: York DOH approved laboratory, if possible. Interpretive Data: Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody. This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes. **Reference Interval:**

<u>Test</u>	<u>Components</u>	Reference Interval
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Number		
	<u>Allergen, Insect, Yellow Hornet IgG</u>	Less than 6.87 mcg/mL

Less than 6.87 mcg/mL



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Effective Date: July 21, 2025

TEST CHANGE

Antimicrobial Susceptibility, AFB/Mycobacteria

0060217, MA AFB	
Specimen Requirements:	
Patient Preparation:	
Collect:	Actively growing isolate in pure culture.
Specimen Preparation:	Transport sealed container with pure <u>isolateculture</u> on solid or liquid media. Place each <u>isolatespecimen</u> in an individually sealed bag.
Transport Temperature:	Room temperature <u>Submit M. tuberculosis complex isolates</u> <u>If</u> culture is suspected of being a microorganism identified on the IATA list as an infectious substance affecting humans, submit specimen according to Infectious Substance, Category A, shipping guidelines.
Unacceptable Conditions:	Mixed <u>isolateseultures</u> or nonviable organisms. <u>M. tuberculosis</u> complex isolatesOrganisms submitted on an agar plate.
Remarks:	
Stability:	Ambient: 2 weeks; Refrigerated: 2 weeks; Frozen: unacceptable2 weeks
Methodology:	Broth Macrodilution/Broth Microdilution
Performed:	Sun-Sat
Reported:	Varies
Note:	AFB susceptibility testing is billed at the panel level. Charges will vary based on organism identified. An additional handling fee will be billed for all organisms submitted that are not in pure culture as indicated in the specimen requirements. If species identification is not provided or if incorrect identification is provided, identification will be performed at ARUP. Additional charges apply. M. tuberculosis complex isolates mono-resistant to pPyrazinamide (PZA) will be further identified to species by PCR at an additional charge. An additional charge will be added for drug requests that are not tested at ARUP and require sendout.
CPT Codes:	CPT codes vary based on method
New York DOH Approval Status:	This test is New York DOH approved.

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Effective Date: July 21, 2025

Interpretive Data:

	•					Inserted Cells
	Test Name	<u>Methodology</u>	Drugs Tested	<u>CPT</u> Code		Inserted Cells
	<u>Antimicrobial</u> <u>Susceptibility -</u> AFB/Mycobacterium	MGIT960 Broth Macro dilution	The interpretation provided is based on results for the following drugs at the stated	<u>87188</u> <u>x4</u>		
	tuberculosis Primary Panel		concentrations: Drugs tested: Ethambutol: 5.0 ug/mL; Isoniazid: 0.1 ug/mL (0.4 ug/mL if resistant to 0.1 ug/mL); Rifampin: 1.0 ug/mL. This procedure screens isolates of M. tuberculosis complex for drug resistance. The procedure does not use serial dilutions to provide quantitative MIC values. Single critical concentrations for each antimycobacterial agent used have been defined by the United States Public Health Service.			
	Antimicrobial Susceptibility - AFB/Mycobacterium	Agar proportion and Broth	Note: If M. tuberculosis complex isolate is resistant to rifampin or any two primary	<u>87190</u> <u>x6,</u> 87188		
	tuberculosis Secondary Panel	dilution	drugs, a secondary panel is available as a send-out test. The interpretation provided is based on testing for the following drugs at the stated concentrations: Drugs tested: Amikacin: 6 ug/mL; capreomycin: 10 ug/mL; cycloserine: 60 ug/mL; ethionamide: 10 ug/mL; kanamycin: 6 ug/mL; PAS: 8 ug/mL; streptomycin at a low level (2.0 ug/mL) and a high level (4.0 ug/mL). Levofloxacin and moxifloxacin are tested at 2.4 and 8 ug/mL	X3		
	<u>Antimicrobial</u> <u>Susceptibility -</u> <u>AFB/Mycobacteria</u>	Broth Microdilution	See organism-specific panels below.	<u>87186</u>		
	Mycobacterium aviumintracellularae Complex	Broth Microdilution	Drugs tested: Amikacin, clarithromycin, linezolid, moxifloxacin. Clofazimine at request only Clarithromycin. Because MIC results do not predict clinical response and may be misleading, rifampin, rifabutin, and ethambutol MICs are not tested.	<u>87186</u>		
	Rapid Growing Mycobacteria	Broth Microdilution	Drugs tested: Amikacin, cefoxitin, ciprofloxacin, clarithromycin, clofazimine,	<u>87186</u>		

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		doxycycline, imipenem, linezolid, moxifloxacin, tigecycline, tobramycin (M. chelonae only), and trimethoprim/sulfamethoxazole (TIMP/SXT). Extended 14-day incubation is performed on isolates initially susceptible to clarithromycin to detect Erm- dependent inducible macrolide resistance. Extended drugs at an additional charge: bedaguiline, omadacycline, and eravacycline	
Miscellaneous Slowly Growing Non-tuberculosis Mycobacteria (NTM, non-fastidious species)	<u>Broth</u> <u>Microdilution</u>	Drugs tested: Amikacin, ciprofloxacin, clarithromycin, doxycycline, linezolid, moxifloxacin, rifabutin, rifampin, streptomycin and trimethoprim/sulfamethoxazole (TIMP/SXT). Selective reporting by organism. CLSI recommends that isolates of M. kansasii be tested against rifampin and clarithromycin only. Rifampin- susceptible isolates are also susceptible isolates are also susceptible to rifabutin. If the isolate is rifampin-resistant, the following secondary drugs will also be reported: Amikacin, ciprofloxacin, rifabutin, streptomycin and trimethoprim- sulfamethoxazole. M. marinum isolates are tested against amikacin, ciprofloxacin, clarithromycin, doxycycline, moxifloxacin, rifabutin, rifampin, and trimethoprim- sulfamethoxazole. Slowly- growing NTM other than M. kansasii and M. marinum are tested against amikacin, ciprofloxacin, clarithromycin, linezolid, moxifloxacin, rifabutin, rifampin, streptomycin, and trimethoprim- sulfamethoxazole.	
Miscellaneous Slowly Growing Non-tuberculosis Mycobacteria (NTM, fastidious species)		Susceptibility testing is not available for M. haemophilum, M. genavense, and M. ulcerans	

Reference Interval:

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Available	Test Name	Methodology	Reference Interval/Drugs	CPT Code
Separately			Tested	07165
JUDU344	Antimicrobial Susceptibility- AFB/Mycobacterium tuberculosis Primary Panel	WG11 200	I the interpretation provided is based on results for the following drugs at the stated concentrations: Drugs tested: Ethambutol: 5.0 ug/mL; Isoniazid: 0.1 ug/mL (0.4 ug/mL); Pyrazinamide: 100 ug/mL; Rifampin: 1.0 ug/mL This procedure screens isolates of M. tuberculosis complex for drug resistance. The procedure does not use serial dilutions to provide quantitative MIC values. Single critical concentrations for each antimycobacterial agent used have been defined by the United States Public Health Service.	87188.x 4
	Antimicrobial Susceptibility - AFB/Mycobacterium tuberculosis Secondary Panel	Agar proportion and Broth dilution	Effective February 21, 2012 Note: If M. tuberculosic isolate is resistant to rifampin or any two primary drugs, a secondary panel will be performed as a send-out test. The interpretation provided is based on testing for the following drugs at the stated concentrations: Drugs tested: Amikacin: 6. ug/mL; capreomycin: 10. ug/mL; cycloserine: 60. ug/mL; ethionamide: 10. ug/mL; kanamycin: 6. ug/mL; kanamycin: 6. ug/mL; kanamycin: 6. ug/mL; kanamycin: 6. ug/mL; hevel (2.0. ug/mL). Levofloxacin and moxifloxacin are tested at 2, 4 and 8. ug/mL	87190 x6, 87188 x3
	Antimicrobial Susceptibility - AFB/Mycobacteria	Broth Microdilution	See organism-specific panels below.	87186
	Mycobacterium avium- intracellularae Complex	Broth Microdilution	Effective-April/1/2022 Drugs tested: Amikacin, clarithromycin, linezolid, moxifloxacin. Clarithromycin. Because MIC results do not predict-clinical response and may be misleading, rifampin, rifabutin, and ethambutol MICs are not tested.	87186
	Rapid Growing Mycobacteria	Broth Microdilution	Effective April 1, 2022 Drugs tested: Amikacin, cefoxitin, ciprofloxacin, clarithromycin, doxycycline, imipenem,	87186

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ole A a A a A a A a A a A a A b A a A b A a A b A <td< th=""><th>linezolid, moxifloxacin, tigecycline, tobramycin (M. chelonae-only), and trimethoprim/sulfamethoxazole (TMP/SXT). Extended 14-day incubation is performed on isolates initially susceptible to clarithromycin to detect Erm(41)-dependent inducible macrolide resistance except Mycobacterium species with a nonfuctional Erm(41) gene. Effective April 1, 2022 Drugs tested: Amikacin, ciprofloxacin, clarithromycin, doxycycline, linezolid, moxifloxacin, rifabutin, rifampin, streptomycin and trimethoprim/sulfamethoxazole (TMP/SXT). Selective reporting by organism. CLS1 recommends that isolates of M. kansasii be tested against rifampin and clarithromycin only. Rifampin- susceptible isolates are also susceptible to rifabutin. If the isolate is rifampin-resistant, the following secondary drugs will also be reported: Amikacin, ciprofloxacin, rifabutin, streptomycin and trimethoprim- sulfamethoxazole. M. marinum isolates are tested against amikacin, ciprofloxacin, clarithromycin, doxycycline, moxifloxacin, rifabutin, rifampin, and trimethoprim- sulfamethoxazole. Slowly- growing NTM other than M. kansasii and M. marinum are tested against amikacin, ciprofloxacin, clarithromycin, linezolid, moxifloxacin, rifabutin, rifampin, streptomycin, and trimethoprim- sulfamethoxazole.</th><th>Broth Microdilution</th><th>Other Slowly- Growing Non- tuberculosis Mycobacteria (NTM)</th><th></th></td<>	linezolid, moxifloxacin, tigecycline, tobramycin (M. chelonae-only), and trimethoprim/sulfamethoxazole (TMP/SXT). Extended 14-day incubation is performed on isolates initially susceptible to clarithromycin to detect Erm(41)-dependent inducible macrolide resistance except Mycobacterium species with a nonfuctional Erm(41) gene. Effective April 1, 2022 Drugs tested: Amikacin, ciprofloxacin, clarithromycin, doxycycline, linezolid, moxifloxacin, rifabutin, rifampin, streptomycin and trimethoprim/sulfamethoxazole (TMP/SXT). Selective reporting by organism. CLS1 recommends that isolates of M. kansasii be tested against rifampin and clarithromycin only. Rifampin- susceptible isolates are also susceptible to rifabutin. If the isolate is rifampin-resistant, the following secondary drugs will also be reported: Amikacin, ciprofloxacin, rifabutin, streptomycin and trimethoprim- sulfamethoxazole. M. marinum isolates are tested against amikacin, ciprofloxacin, clarithromycin, doxycycline, moxifloxacin, rifabutin, rifampin, and trimethoprim- sulfamethoxazole. Slowly- growing NTM other than M. kansasii and M. marinum are tested against amikacin, ciprofloxacin, clarithromycin, linezolid, moxifloxacin, rifabutin, rifampin, streptomycin, and trimethoprim- sulfamethoxazole.	Broth Microdilution	Other Slowly- Growing Non- tuberculosis Mycobacteria (NTM)	
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Bile Acids, Total						
Specimen Requirements:						
Patient Preparation:	Patient should fast for 8 hours prior to collection.					
Collect:	Serum separator tube or plasma separator tube. Also acceptable : Lavender (EDTA), green (lithium heparin)					
Specimen Preparation:	Allow specimen to clot completely at room temperature before centrifugation. Transfer 1 mL serum to an ARUP <u>standard</u> <u>transport tube.</u> Standard Transport Tube. (Min: 0.5 mL)					
Transport Temperature:	Refrigerated.					
Unacceptable Conditions:	Body fluids. Hemolyzed specimens.					
Remarks:						
Stability:	After separation from cells: Ambient: <u>24</u> 8 hours; Refrigerated: 2 weeks; Frozen: 3 months					
Methodology:	Quantitative Enzymatic Assay					
Performed:	Sun-Sat					
Reported:	Within 24 hours					
Note:						
CPT Codes:	82239					
New York DOH Approval Status:	This test is New York DOH approved.					
Interpretive Data:						
Reference interval applies to fasting specimens.						
Reference Interval:	Reference Interval:					
0-10 μmol/L						



Vitamin A (Retinol), Serum or Plasma 0080525. VIT A				
Specimen Requirements:				
Patient Preparation:	Patient should fast for 12 hours and abstain from alcohol for 24 hours prior to collection.			
Collect:	Green (sodium or lithium heparin), plasma separator tube, or serum separator tube. Also acceptable: Lavender (EDTA) or pink (K2EDTA).			
Specimen Preparation:	Separate serum or plasma within 1 hour of collection. Transfer 1 mL serum or plasma to an ARUP Standard Transport Tube immediately. (Min: 0.2 mL) Avoid hemolysis.			
Transport Temperature:	Refrigerated.			
Unacceptable Conditions:	Whole blood or body fluids other than serum or plasma.			
Remarks:				
Stability:	After separation from cells: Ambient: <u>unacceptable</u> 3-hours; Refrigerated: 1 month; Frozen: 1 year			
Methodology:	Quantitative High Performance Liquid Chromatography (HPL			
Performed:	Sun-Sat			
Reported:	1-4 days			
Note:	Serum retinol is typically maintained until hepatic stores are almost depleted. Values greater than 0.30 mg/L represent adequate liver stores, whereas values less than 0.10 mg/L indicate deficiency. Samples that come in contact with plastic tubing or have been exposed to excessive light may show low results. Vitamin A toxicity occurs when retinol concentration exceeds the capacity of retinol binding protein (RBP). Individuals with compromised renal function can retain RBP and may, therefore, have moderate retinol elevations. Drugs which interfere with vitamin A analysis include probucol (Lorelco). This assay does not measure other vitamin A metabolites such as retinaldehyde and retinoic acid.			
CPT Codes:	84590			
New York DOH Approval Status:	This test is New York DOH approved.			



Interpretive Data:

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Test Number	Components	Reference Interval		
	Vitamin A (Retinol)			
		Age	Reference Interval	
		0-1 month 2 months-12 years 13-17 years 18 years and older	0.18-0.50 mg/L 0.20-0.50 mg/L 0.26-0.70 mg/L 0.30-1.20 mg/L	
	Vitamin A (Retinyl Palmitate)	0-150 years: 0-	0.10 mg/L	



Allergen, Food, Almond IgG					
0090284, ALMOND IGG					
Specimen Requirements:					
Patient Preparation:					
Collect:	Plain red or serum Serum separator tube (SST)).				
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>25</u> 2 mL) <u>Refer to</u> <u>"Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>				
Transport Temperature:	Refrigerated.				
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.				
Remarks:					
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year				
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay				
Performed:	Sun				
Reported:	1-8 days				
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL				
CPT Codes:	86001				
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.				
Interpretive Data:					
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.					
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was					

nas not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Effective November 19, 2012


Less than 15.21 mcg/mL



Allergen, Food, Banana IgG	
0090286, BANANA IGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Plain red or serumSerum separator tube (SST)).
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>252 mL). Refer to</u> <u>"Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.	
This test was developed and its pe	rformance characteristics determined by ARUP Laboratories. It

nas not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:



Less than 46.11 mcg/mL



Allergen, Food, Garlic IgG	
0090287, GARLIC IGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Plain red or serum Serum separator tube (SST)).
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to</u> <u>"Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL repr antibody.	resent absent or undetectable levels of allergen-specific IgG
This test was developed and its pe	rformance characteristics determined by ARUP Laboratories. It

has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:



Less than 12.71 mcg/mL



Allergen, Food, Gluten IgG	
Specimen Bequirements:	
Patient Prenaration	
Collect:	Plain red or serum Serum separator tube (SST)).
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.252 mL). Refer to "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.	
This test was developed and its per has not been cleared or approved b	rformance characteristics determined by ARUP Laboratories. It wy the US Food and Drug Administration. This test was

nas not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:



Less than 47.41 mcg/mL



Allergen, Food, Whey IgG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Plain red or serum Serum separator tube (SST)
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.252 mL). Refer to "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.
Transport Temperature:	Refrigerated
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL repr antibody. This test was developed and its pe has not been cleared or approved to performed in a CLIA certified labora	resent absent or undetectable levels of allergen-specific IgG rformance characteristics determined by ARUP Laboratories. It by the US Food and Drug Administration. This test was atory and is intended for clinical purposes.
Reference Interval:	

Less than 88.61 mcg/mL





Allergen, Fungi and Molds, Fusarium proliferatum/moniliforme IgG 0093454, FUS M IGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Plain red or serumSerum separator tube (SST)).	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>25</u> 2 mL). <u>Refer to</u> <u>"Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL	
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its pe has not been cleared or approved b	rformance characteristics determined by ARUP Laboratories. It by the US Food and Drug Administration. This test was	

performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Less than 46.41 mcg/mL





Allergen, Food, Mushroom IgG	
Specimen Bequirements:	
Patient Prenaration:	
Collect:	Serum separator tube (SST).
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.252 mL). Refer to "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL repr antibody.	resent absent or undetectable levels of allergen-specific IgG
This test was developed and its pe	rformance characteristics determined by ARUP Laboratories. It

has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:



Less than 15.61 mcg/mL



Allergen, Food, Coffee IgG	
0097302, COFFEE IGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Plain red or serum separator tube.
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>25</u> 2 mL). <u>Refer to</u> "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.}
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL repr antibody.	resent absent or undetectable levels of allergen-specific IgG
This test was developed and its pe	rformance characteristics determined by ARUP Laboratories. It

has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:



Less than 13.51 mcg/mL



Allergen, Fungi and Molds, Can 0097304, CANDIDAIGG	dida albicans IgG
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube (SST).
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to</u> "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.}
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL repr antibody.	esent absent or undetectable levels of allergen-specific IgG
This test was developed and its per	formance characteristics determined by ARUP Laboratories. It

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Less than 84.01 mcg/mL





Allergen, Fungi and Molds, Aure 0097305, AUREO IGG	eobasidium pullulans IgG
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube (SST).
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>25</u> 2 mL). <u>Refer to</u> "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.}
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL repr antibody.	esent absent or undetectable levels of allergen-specific IgG
This test was developed and its per	formance characteristics determined by ARUP Laboratories. It

This test was developed and its performance characteristics determined by ARUP Laboratories. has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Less than 17.51 mcg/mL





Allergen, Food, Onion IgG	
0097306, ONION IGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube (SST).
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>252 mL). Refer to</u> "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL repr antibody.	esent absent or undetectable levels of allergen-specific IgG
This test was developed and its pe	rformance characteristics determined by ARUP Laboratories. It

This test was developed and its performance characteristics determined by ARUP Laboratories. If has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:



Less than 17.01 mcg/mL



Allergen, Fungi and Molds, Rhiz 0097307, RHIZO IGG	opus nigricans IgG
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube (SST).
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>25</u> 2 mL). <u>Refer to</u> "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL repr antibody.	esent absent or undetectable levels of allergen-specific IgG
This test was developed and its per	formance characteristics determined by ARUP Laboratories. It

has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Less than 8.41 mcg/mL





Allergen, Stemphylium herbarum/botryosum, IgG		
0097308, STEMPHBIGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Plain red or serum Serum separator tube (SST)).	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum or plasma to an ARUP standard transport tube.Standard Transport Tube. (Min: 0.252 mL). Refer to "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL	
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its pe has not been cleared or approved by	rformance characteristics determined by ARUP Laboratories. It by the US Food and Drug Administration. This test was	

nas not been cleared or approved by the US Food and Drug Administration. This test performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Less than 115.01 mcg/mL





Allergen, Fungi and Molds, Phoma betae IgG		
0097309, PHOMAB IGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Serum separator tube (SST).	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>25</u> 2 mL). <u>Refer to</u> "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL	
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its performance characteristics determined by ARUP Laboratories. It		

has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Less than 12.91 mcg/mL





Allergen, Fungi and Molds, Penicillium chrysogenum/notatum IgG 0097310, PENI N IGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Serum separator tube (SST).	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>25</u> 2 mL). <u>Refer to</u> "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.}	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL	
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was		

performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Less than 55.51 mcg/mL





Allergen, Fungi and Molds, Helminthosporium halodes/Setomelanomma rostrata IgG

0097313, HELMINIGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Serum separator tube (SST).	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.252 mL). Refer to <u>"Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL	
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was		

performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Less than 131.01 mcg/mL





TEST CHANGE

Allergen, Fungi and Molds, Clad 0097314, CLADO IGG	losporium IgG	
Specimen Requirements:		
Patient Preparation:		
Collect:	Serum separator tube (SST).	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to</u> "Allergen Specimen Collection Instructions" at <u>www.aruplab.com/testing/resources/specimen.</u>)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL	
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its performance characteristics determined by ARUP Laboratories. It		

has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Less than 86.11 mcg/mL





Allergen, Food, Egg Yolk IgG		
0097315, EGGYOLKIGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Serum separator tube (SST).	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>252 mL). Refer to</u> <u>"Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL	
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was		

Reference Interval:

Effective November 19, 2012

performed in a CLIA certified laboratory and is intended for clinical purposes.



Less than 19.61 mcg/mL



Allergen, Fungi and Molds, Mucor racemosus IgG 0097316, MUCOR IGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Serum separator tube (SST).	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.252 mL). Refer to "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.}	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL	
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was		

performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Less than 9.77 mcg/mL




Allergen, Food, Rice IgG 0097323, RICE IGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube (SST).
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>25</u> 2 mL). <u>Refer to</u> "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL repr antibody.	esent absent or undetectable levels of allergen-specific IgG
This test was developed and its per	rformance characteristics determined by ARUP Laboratories. It

has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Less than 16.71 mcg/mL





Allergen, Food, Wheat IgG 0097636, WHEAT IGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Serum separator tube.	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to</u> "Allergen Specimen Collection Instructions" at <u>www.aruplab.com/testing/resources/specimen.</u>)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:		
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its per has not been cleared or approved b performed in a CLIA certified labora	rformance characteristics determined by ARUP Laboratories. It by the US Food and Drug Administration. This test was atory and is intended for clinical purposes.	
Reference Interval:		
Effective August 20, 2012		

Less than 60.20 mcg/mL





Allergen, Food, Potato (White) IgG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Plain red or serumSerum separator tube (SST)).
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.252 mL). Refer to "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.)
Transport Temperature:	Refrigerated
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 1 week; Refrigerated: 1 month; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000mcg/1000mL.
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.	
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was	

performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Less than 6.09 mcg/mL



Allergen, Food, Rye IgG	
0097642, RYE IGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube (SST <u>)</u>).
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>252 mL). Refer to</u> <u>"Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL repr antibody.	resent absent or undetectable levels of allergen-specific IgG
This test was developed and its pe	rformance characteristics determined by ARUP Laboratories. It

has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Less than 26.71 mcg/mL





Allergen, Food, Soybean IgG	
0097643, SOY IGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Plain red or serumSerum separator tube (SST)-
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum at to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>25</u> 2 mL). <u>Refer to</u> "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL repr antibody.	resent absent or undetectable levels of allergen-specific IgG
This test was developed and its per has not been cleared or approved be performed in a CLIA certified labora	rformance characteristics determined by ARUP Laboratories. It by the US Food and Drug Administration. This test was atory and is intended for clinical purposes.
Reference Interval:	
Effective August 20, 2012	

Less than 5.30 mcg/mL



Allergen, Tomato IgG	
0097644, TOMATO IGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube (SST <u>)</u> .
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>252 mL). Refer to</u> <u>"Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.	
This test was developed and its pe	rformance characteristics determined by ARUP Laboratories. It

has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Less than 7.20 mcg/mL





Allergen, Food, Orange IgG	
0097647, ORANGE IGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Collect: Serum separator tube (SST <u>)</u> .
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to</u> "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL repr antibody.	resent absent or undetectable levels of allergen-specific IgG
This test was developed and its per	rformance characteristics determined by ARUP Laboratories. It

has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:



Less than 8.65 mcg/mL



Allergen, Food, Peanut IgG 0097648, PEANUT IGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Plain red or serum Serum separator tube (SST)-	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to</u> "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:		
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its per has not been cleared or approved b performed in a CLIA certified labora	rformance characteristics determined by ARUP Laboratories. It by the US Food and Drug Administration. This test was atory and is intended for clinical purposes.	
Reference Interval:		
Effective August 20, 2012		

Less than 6.80 mcg/mL



Allergen, Food, Pork IgG	
0097649, PORK IGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube (SST <u>)</u> .
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>252 mL). Refer to</u> "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL repr antibody.	esent absent or undetectable levels of allergen-specific IgG
This test was developed and its pe	rformance characteristics determined by ARUP Laboratories. It

has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:



Less than 7.92 mcg/mL



Allergen, Food, Lettuce IgG	
0097651, LETT IGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube (SST <u>)</u> .
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>25</u> 2 mL). <u>Refer to</u> <u>"Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.	
This test was developed and its per	rformance characteristics determined by ARUP Laboratories. It

has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:



Less than 11.31 mcg/mL

Allergen, Food, Malt IgG	
0097652, MALT IGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Collect: Serum separator tube (SST)).
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>252 mL). Refer to</u> <u>"Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL repr antibody.	resent absent or undetectable levels of allergen-specific IgG
This test was developed and its pe	rformance characteristics determined by ARUP Laboratories. It

has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:



Less than 22.31 mcg/mL



Allergen, Food, Casein (Cow's Milk) IgG		
0097653, CASEIN IGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Plain red or serum Serum separator tube (SST)-	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to</u> <u>"Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL	
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was		

Reference Interval:

Effective August 20, 2012

performed in a CLIA certified laboratory and is intended for clinical purposes.



Less than 38.70 mcg/mL



Allergen, Food, Oat IgG		
0097654, OAT IGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Plain red or serum Serum separator tube (SST)-	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>25</u> 2 mL). <u>Refer to</u> "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:		
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.		
Reference Interval:		
Effective August 20, 2012		

Less than 13.30 mcg/mL





Allergen, Food, Chicken IgG		
0097656, CHICK IGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Serum separator tube (SST <u>)</u> .	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>25</u> 2 mL). <u>Refer to</u> "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL	
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was		

performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:



Less than 6.25 mcg/mL



Allergen, Food, Chocolate IgG		
0097657, CHOCO IGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Plain red or serum separator tube <u>(SST)</u> -	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to</u> <u>"Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL	
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its performance characteristics determined by ARUP Laboratories. It		

has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:



Less than 20.41 mcg/mL



Allergen, Food, Corn IgG		
0097658, CORN IGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Plain red or serum Serum separator tube (SST)-	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to</u> <u>"Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL	
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Effective May 21, 2012



Less than 10.5 mcg/mL



Allergen, Food, Egg White IgG		
0097659, EGG IGG		
Detient Dres enstiant		
Patient Preparation:		
Collect:	Plain red or serumSerum separator tube (SST)-	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>25</u> 2 mL). <u>Refer to</u> <u>"Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL	
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was		

performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Effective August 20, 2012



Less than 15.70 mcg/mL



Allergen, Food, Baker's Yeast IgG		
0097706, YEAST IGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Serum separator tube (SST).	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>25</u> 2 mL). <u>Refer to</u> <u>"Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL	
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was		

nas not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:


Less than 11.41 mcg/mL



Allergen, Food, Barley IgG			
0097707, BARLEY IGG			
Specimen Requirements:			
Patient Preparation:			
Collect:	Serum separator tube (SST <u>)</u>).		
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>25</u> 2 mL). <u>Refer to</u> "Allergen Specimen Collection Instructions" at <u>www.aruplab.com/testing/resources/specimen.</u>)		
Transport Temperature:	Refrigerated.		
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.		
Remarks:			
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year		
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay		
Performed:	Sun		
Reported:	1-8 days		
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL		
CPT Codes:	86001		
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.		
Interpretive Data:			
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.			
This test was developed and its performance characteristics determined by ARUP Laboratories. It			

has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Effective November 19, 2012



Less than 20.31 mcg/mL



Allergen, Food, Beef IgG		
0097708, BEEF IGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Serum separator tube (SST <u>)</u> .	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>25</u> 2 mL). <u>Refer to</u> <u>"Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL	
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was		

nas not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Effective November 19, 2012



Less than 22.01 mcg/mL



Allergen, Fungi and Molds, Alternaria tenuis IgG 0097773, ALTER IGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Serum separator tube (SST).	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.252 mL). Refer to "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL	
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL repr antibody.	resent absent or undetectable levels of allergen-specific IgG	
This test was developed and its performance characteristics determined by APLID I shoretorics. It		

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Less than 16.71 mcg/mL



Keppra (Levetiracetam) 0098627, KEPPRA				
Specimen Requirements:				
Patient Preparation:	Timing of specimen collection: <u>PredosePre-dose</u> (trough) draw <u>at</u> -At steady state concentration.			
Collect:	Plain red. Also acceptable: Lavender (EDTA), pink (K2EDTA), or green (sodium or lithium heparin).			
Specimen Preparation:	Separate serum or plasma from cells ASAP or within 2 hours of collection. Transfer 1 mL serum or plasma to an ARUP <u>standard transport tube.</u> Standard Transport Tube. (Min: 0.3 mL)			
Transport Temperature:	Refrigerated.			
Unacceptable Conditions:	Serum or plasma separator tubes. Grossly hemolyzed specimens.			
Remarks:				
Stability:	After separation from cells: Ambient: 7 days; Refrigerated: 1 week; Frozen: 1 month			
Methodology:	Quantitative Enzyme Immunoassay <u>(EIA)</u>			
Performed:	Sun-Sat			
Reported:	Within 24 hours			
Note:				
CPT Codes:	80177			
New York DOH Approval Status:	This test is New York DOH approved.			

Interpretive Data:

Pharmacokinetics of levetiracetam are affected by renal function. Adverse effects may include somnolence, weakness, headache and vomiting.

This levetiracetam (Keppra) immunoassay uses the ARK Diagnostics reagents, which has known cross-reactivity with the drug brivaracetam (Briviact) and may report inaccurate results. Patients transitioning from levetiracetam to brivaracetam or those who are using both medications should not monitor drug concentrations with the ARK Diagnostics assay. These patients should be monitored using a validated chromatographic methodology that distinguishes between drugs to determine drug concentrations.



Reference Interval:

Effective February 22, 2022

HOTLINE NOTE: There is a numeric map change associated with this test. Refer to the Hotline Test Mix for interface build information.

Chromium, Serum				
0098830, CR S				
Specimen Requirements:				
Patient Preparation:	Diet, medication, and nutritional supplements may introduce interfering substances. Patients should be encouraged to discontinue nutritional supplements, vitamins, minerals, and <u>nonessentialnon-essential</u> over-the-counter medications (upon the advice of their physician).			
Collect:	Royal <u>blue (no additive</u> Blue (No Additive).			
Specimen Preparation:	Separate from cells ASAP or within 2 hours of collection. Transfer 2 mL serum to an ARUP Trace Element-Free Transport Tube (ARUP supply #43116) available online through eSupply using ARUP Connect(TM) or contact ARUP Client Services at (800-)-522-2787- (Min: 0.5 mL). Do not use utensils (i.e., syringes, needles, or pipettes) in the collection or transfer of the sample, pour directly into transport tube.)			
Transport Temperature:	Room temperature. Also acceptable: Refrigerated or frozen.			
Unacceptable Conditions:	Plasma. Royal <u>b</u> Blue (EDTA) or separator tubes. Specimens that are not separated from the clot within 2 hours. Specimens transported in tubes other than specified.			
Remarks:				
Stability:	Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Indefinitely			
Methodology:	Quantitative Inductively Coupled Plasma-Mass Spectrometry			
Performed:	Sun-Sat			
Reported:	1-3 days			
Note:				
CPT Codes:	82495			
New York DOH Approval Status:	This test is New York DOH approved.			
Interpretive Data:				
Elevated results may be due to ski noncertified metal-free collection/	n or collection-related contamination, including the use of a transport tube. If contamination concerns exist due to elevated			

levels of serum chromium, confirmation with a second specimen collected in a certified metal-free



tube is recommended.

Serum chromium levels can be significantly higher in patients with metal-on-metal total hip replacement implants than in control patients without metal implants. Serum chromium levels may be increased in asymptomatic patients with metal-on-metal prosthetics and should be considered in the context of the overall clinical scenario. Whole blood is the specimen type recommended by the U.S. Food and Drug Administration for assessing the risks of metal-on-metal hip implants in symptomatic patients.

Symptoms associated with chromium toxicity vary based on route of exposure and dose, and may include dermatitis, impairment of pulmonary function, gastroenteritis, hepatic necrosis, bleeding, and acute tubular necrosis.-

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Less than or equal to 5.0 μ g/L



TEST CHANGE

Arsenic, <u>Whole</u> Blood 0099045, ARS B			
Specimen Requirements:			
Patient Preparation:	Diet, medication, and nutritional supplements may introduce interfering substances. Patient should be encouraged to discontinue nutritional supplements, vitamins, minerals, and non-essential over-the-counter medications (upon the advice of their physician) and avoid shellfish and seafood for 48 to 72 hours.		
Collect:	Royal blue (K2EDTA) or royal blue (NaHep).		
Specimen Preparation:	Transport 3 or 6 mL whole blood in the original collection tube. (Min: 0.5 mL)		
Transport Temperature:	Room temperature. Also acceptable: Refrigerated.		
Unacceptable Conditions:	Specimens collected in tubes other than royal blue (K2EDTA) or royal blue (NaHep). Specimens transported in containers other than royal blue (K2EDTA) or royal blue (NaHep) tube or trace element-free transport tube. Clotted specimens.		
Remarks:			
Stability:	Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable		
Methodology:	Quantitative Inductively Coupled Plasma-Mass Spectrometry (ICP-MS)		
Performed:	Sun-Sat		
Reported:	1-3 days		
Note:			
CPT Codes:	82175		

New York DOH Approval Status: This test is New York DOH approved.

Interpretive Data:

Elevated results may be due to skin or collection-related contamination, including the use of a noncertified metal-free collection/transport tube. If contamination concerns exist due to elevated levels of blood arsenic, confirmation with a second specimen collected in a certified metal-free tube is recommended.

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A nonprofit enterprise of the University of Utah and its Department of Pathology

Effective Date: July 21, 2025

Potentially toxic ranges for blood arsenic: Greater than or equal to 600 ug#g/L.

Blood arsenic is for the detection of recent exposure poisoning only. Blood arsenic levels in healthy subjects vary considerably with exposure to arsenic in the diet and the environment. A 24-hour urine arsenic is useful for the detection of chronic exposure.

Elevated results may be due to skin- or collection-related contamination, including the use of tubes that are not certified to be trace element free. If an elevated result is suspected to be due to contamination, confirmation with a second specimen collected in a certified trace element-free tube is recommended.

Methodology: Inductively Coupled Plasma-Mass Spectrometry (ICP-MS).

Reference Interval:

Tes	<u>t Number</u>	<u>Components</u>	Reference Interval		
ļ	Arsenic, Whole Blood	Less than or equal to 12.0 μg/L			Inserted Cells

Inserted Cells

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Cobalt, <u>Whole</u> Blood				
0099231, COBALT B				
Specimen Requirements:				
Patient Preparation:	Diet, medication, and nutritional supplements may introduce interfering substances. Patients should be encouraged to discontinue nutritional supplements, vitamins, minerals, and non-essential over-the-counter medications (upon the advice of their physician).			
Collect:	Royal blue (K2EDTA) or royal blue (NaHep).			
Specimen Preparation:	Transport 3 or 6 mL whole blood in the original collection tube. (Min: 0.5 mL)			
Transport Temperature:	Room temperature. Also acceptable: Refrigerated.			
Unacceptable Conditions:	Specimens collected in tubes other than royal blue (K2EDTA) or royal blue (NaHep). Clotted specimens.			
Remarks:				
Stability:	Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable			
Methodology:	Quantitative Inductively Coupled Plasma-Mass Spectrometry (ICP-MS)			
Performed:	Sun-Sat			
Reported:	1-3 days			
Note:				
CPT Codes:	83018			
New York DOH Approval Status:	This test is New York DOH approved.			
Intermenting Dates				

Interpretive Data:

Elevated results may be due to skin or collection-related contamination, including the use of a noncertified metal-free collection/transport tube. If contamination concerns exist due to elevated levels of blood cobalt, confirmation with a second specimen collected in a certified metal-free tube is recommended.

Blood cobalt levels can be used in the assessment of occupational exposure or toxic ingestion. Symptoms associated with cobalt toxicity vary based on route of exposure and may include cardiomyopathy, allergic dermatitis, pulmonary fibrosis, cough and dyspnea. Blood is the preferred



specimen type for evaluating metal ion release from metal-on-metal joint arthroplasty.

Elevated results may be due to skin- or collection-related contamination, including the use of tubes that are not certified to be trace element free. If an elevated result is suspected to be due to contamination, confirmation with a second specimen collected in a certified trace element-free tube is recommended.

Methodology: Inductively Coupled Plasma-Mass Spectrometry (ICP-MS).

Reference Interval:

Less than or equal to 0.5-3.9 µg/L



Ribosomal P Protein Antibody	
0099249, RIBPP	
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube <u>or red tube</u> .
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 1 mL serum to an ARUP <u>standard transport</u> <u>tube.</u> Standard Transport Tube. (Min: 0.2 mL)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Plasma or other body fluids. Bacterially contaminated or severely lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)
Methodology:	Semi-Quantitative Multiplex Bead Assay
Performed:	Sun-Sat
Reported:	1-3 days
Note:	
CPT Codes:	83516

New York DOH Approval Status: This test is New York DOH approved.

Interpretive Data:

Autoantibodies reacting with cytoplasmic ribosomes are highly specific for systemic lupus erythematosus. Ribosomal-P antibodies are found in approximately 12% of patients with systemic lupus erythematosus (SLE) and in 90% of patients with lupus psychosis; titers often increase more than fivefold during and before active phases of psychosis.

Component	Interpretation
Ribosomal P	29 AU/mL or less
Protein Antibody	Negative 30-40
	AU/mL Equivocal
	41 AU/mL or
	greater Positive

Reference Interval:



Test Number	Components	Reference Interval		
	Ribosome P Antibody, IgG	0-40 AU/mL		

Aluminum, Serum				
0099266, AL S				
Specimen Requirements:				
Patient Preparation:	Diet, medication, and nutritional supplements may introduce interfering substances. Patient should be encouraged to discontinue nutritional supplements, vitamins, minerals, and <u>nonessentialnon-essential</u> over-the-counter medications (upon the advice of their physician).			
Collect:	Royal Blue (No Additive).			
Specimen Preparation:	Separate from cells ASAP or within 2 hours of collection. Transfer 2 mL serum to an ARUP Trace Element-Free Transport Tube (ARUP supply #43116) available online through eSupply using ARUP Connect(TM) or contact ARUP Client Services at (800-)-522-2787- (Min: 0.5 mL). Do not use utensils (i.e., syringes, needles, or pipettes) in the collection or transfer of the sample, pour directly into transport tube.)			
Transport Temperature:	Room temperature. Also acceptable: Refrigerated or frozen.			
Unacceptable Conditions:	Plasma. Specimens that are not separated from the red cells or clot within 2 hours. Specimens collected in containers other than specified. Specimens transported in containers other than specified.			
Remarks:				
Stability:	Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Indefinitely (If the specimen is drawn and stored in the appropriate container, the trace element values do not change with time.)			
Methodology:	Quantitative Inductively Coupled Plasma-Mass Spectrometry			
Performed:	Tue, Thu, Sat			
Reported:	1-4 days			
Note:				
CPT Codes:	82108			
New York DOH Approval Status:	This test is New York DOH approved.			
Interpretive Data:				



Serum aluminum greater than 50.0 $\mu\text{g/L}$ is consistent with overload and may correlate with toxicity.

Elevated results may be due to skin or collection-related contamination, including the use of a noncertified metal-free collection/transport tube. If contamination concerns exist due to elevated levels of serum aluminum, confirmation with a second specimen collected in a certified metal-free tube is recommended.

Reference Interval:

0.0-15.0 μg/L

Manganese, Whole Blood	
0099272, MANG WB	
Specimen Requirements:	
Patient Preparation:	Diet, medication, and nutritional supplements may introduce interfering substances. Patient should be encouraged to discontinue nutritional supplements, vitamins, minerals, and non-essential over-the-counter medications (upon the advice of their physician).
Collect:	Royal blue (K2EDTA) or royal blue (NaHep).
Specimen Preparation:	Transport 3 or 6 mL whole blood in the original collection tube. (Min: 0.5 mL)
Transport Temperature:	Room temperature. Also acceptable: Refrigerated.
Unacceptable Conditions:	Specimens collected in tubes other than royal blue (K2EDTA) or royal blue (NaHep). Specimens transported in containers other than a royal blue (K2EDTA) or royal blue (NaHep) tube or trace element-free transport tube. Clotted specimens.
Remarks:	
Stability:	Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable
Methodology:	Quantitative Inductively Coupled Plasma-Mass Spectrometry (ICP-MS)
Performed:	Sun-Sat
Reported:	1-3 days
Note:	
CPT Codes:	83785
New York DOH Approval Status:	This test is New York DOH approved.

Interpretive Data:

Elevated results may be due to skin<u></u> or collection-related contamination, including the use of <u>tubes</u> <u>that are not certified to be trace element</u> <u>a noncertified metal-free</u><u>-collection/transport tube</u>. If <u>an</u> <u>elevated result is suspected to be due to</u> contamination-concerns exist due to elevated levels of <u>blood manganese</u>, confirmation with a second specimen collected in a certified <u>trace</u> <u>element</u><u>metal</u>-free tube is recommended.



Methodology: Inductively Coupled Plasma-Mass Spectrometry (ICP-MS).

Reference Interval:

4.2-16.5 μg/L

Mercury, Whole Blood	
0099305, HG B	
Specimen Requirements:	
Patient Preparation:	Diet, medication, and nutritional supplements may introduce interfering substances. Patient should be encouraged to discontinue nutritional supplements, vitamins, minerals, and non-essential over-the-counter medications (upon the advice of their physician), and avoid shellfish and seafood for 48 to 72 hours.
Collect:	Royal blue (K2EDTA) or royal blue (NaHep).
Specimen Preparation:	Transport 3 or 6 mL whole blood in the original collection tube. (Min: 0.5 mL)
Transport Temperature:	Room temperature. Also acceptable: Refrigerated.
Unacceptable Conditions:	Specimens collected in tubes other than royal blue (K2EDTA) or royal blue (NaHep). Specimens transported in containers other than royal blue (K2EDTA) or royal blue (NaHep) tube or trace element-free transport tube. Clotted specimens.
Remarks:	
Stability:	Ambient: 1 week; Refrigerated: 1 week; Frozen: Unacceptable
Methodology:	Quantitative Inductively Coupled Plasma-Mass Spectrometry (ICP-MS)
Performed:	Sun-Sat
Reported:	1-3 days
Note:	Mercury is volatile; concentration may decrease over time.
CPT Codes:	83825
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	
Elevated results may be due to skir	or collection-related contamination, including the use of a

noncertified metal-free collection/transport tube. If contamination concerns exist due to elevated levels of blood mercury, confirmation with a second specimen collected in a certified metal-free tube is recommended.



Blood mercury levels predominantly reflect recent exposure and are most useful in the diagnosis of acute poisoning as blood mercury concentrations rise sharply and fall quickly over several days after ingestion. Blood concentrations in unexposed individuals rarely exceed 20 ugµg/L. The provided reference interval relates to inorganic mercury concentrations. Dietary and nonoccupational non-occupational exposure to organic mercury forms may contribute to an elevated total mercury result. Clinical presentation after toxic exposure to organic mercury may include dysarthria, ataxia, and constricted vision fields with mercury blood concentrations from 20 to 50 ugµg/L.

Elevated results may be due to skin- or collection-related contamination, including the use of tubes that are not certified to be trace element free. If an elevated result is suspected to be due to contamination, confirmation with a second specimen collected in a certified trace element-free tube is recommended.

Methodology: Inductively Coupled Plasma-Mass Spectrometry (ICP-MS).

Reference Interval:

Less than or equal to 10.0 $\mu\text{g/L}$

Nickel, Serum	
0099452, NICKEL	
Specimen Requirements:	
Patient Preparation:	Diet, medication, and nutritional supplements may introduce interfering substances. Patients should be encouraged to discontinue nutritional supplements, vitamins, minerals, and <u>nonessential</u> non-essential over-the-counter medications (upon the advice of their physician).
Collect:	Royal <u>blue (no additive</u> Blue (No Additive).
Specimen Preparation:	Separate from cells ASAP or within 2 hours of collection. Transfer 2 mL serum-within 2 hours of collection to an ARUP Trace Element-Free Transport Tube (ARUP supply #43116) available online through eSupply using ARUP Connect(TM) or contact ARUP Client Services at (800-)-522-2787- (Min: 0.5 mL). Do not use utensils (i.e., syringes, needles, or pipettes) in the collection or transfer of the sample, pour directly into transport tube.)
Transport Temperature:	Room temperature. Also acceptable: Refrigerated or frozen.
Unacceptable Conditions:	Plasma. Specimens that are not separated from clot, within 2 hours. Separator tubes or <u>royal blue</u> Royal-Blue (EDTA). Specimens transported in tubes other than specified. Hemolyzed specimens.
Remarks:	
Stability:	Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Indefinitely
Methodology:	Quantitative Inductively Coupled Plasma-Mass Spectrometry
Performed:	Sun-Sat
Reported:	1-3 days
Note:	
CPT Codes:	83885
New York DOH Approval Status: Interpretive Data:	This test is New York DOH approved.



Elevated results may be due to skin or collection-related contamination, including the use of a noncertified metal-free collection/transport tube. If contamination concerns exist due to elevated levels of serum nickel, confirmation with a second specimen collected in a certified metal-free tube is recommended.

Serum nickel testing is intended to detect potentially toxic exposure.

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Less than or equal to 10.0 $\mu\text{g/L}$

Heavy Metals Panel 3, <u>Whole</u> Blood			
0099470, HY MET B			
Specimen Requirements:			
Patient Preparation:	Diet, medication, and nutritional supplements may introduce interfering substances. Patients should be encouraged to discontinue nutritional supplements, vitamins, minerals, non- essential over-the-counter medications (upon the advice of their physician), and avoid shellfish and seafood for 48 to 72 hours.		
Collect:	Royal blue (K2EDTA) or Royal blue (NaHep).		
Specimen Preparation:	Transport 3 or 6 mL whole blood in the original collection tube. (Min: 0.5 mL)		
Transport Temperature:	Room temperature. Also acceptable: Refrigerated.		
Unacceptable Conditions:	Specimens collected in tubes other than Royal blue (K2EDTA) or Royal blue (NaHep). Specimens transported in containers other than Royal blue (K2EDTA) or Royal blue (NaHep) or Trace Element-Free Transport Tube. Clotted specimens.		
Remarks:			
Stability:	Ambient: 1 week; Refrigerated: 1 week; Frozen: Unacceptable		
Methodology:	Quantitative Inductively Coupled Plasma-Mass Spectrometry (ICP-MS)		
Performed:	Sun-Sat		
Reported:	1-4 days		
Note:	Mercury is volatile; concentration may decrease over time. If the specimen is drawn and stored in the appropriate container, the arsenic and lead values do not change with time.		
CPT Codes:	82175; 83655; 83825		
New York DOH Approval Status:	This test is New York DOH approved.		
Interpretive Data:			
Refer to report.			
Reference Interval:			



Test Number	Components	Reference Inte	rval	
Turnser	Lead, <u>Whole</u> Blood (Venous)	Less than or ed	<u>μal to 3.4 μg/L</u>	
	Lead, Whole Blood (Venous)			
		Concentration	Comment	
		3.5-19.9 ug/dL	Children under the age of 6 years are the most vulnerable to the harmful effects of lead exposure. Environmental investigation and exposure history to identify potential sources of lead. Biological and nutritional monitoring are recommended. Follow-up blood lead monitoring is recommended.	
		20-44.9 ug/dL	Lead hazard reduction and prompt medical evaluation are recommended. Contact a Pediatric Environmental Health Specialty Unit or poison control center for guidance.	
		Greater than 44.9 ug/dL	Critical. Immediate medical evaluation, including detailed neurological exam is recommended. Consider chelation therapy when symptoms of lead toxicity are present. Contact a Pediatric Environmental Health Specialty Unit or poison control center for	
	Lead, Whole Blood (Venous)		assistance.	



	Concentration	Comment
	5-19.9 ug/dL	Medical removal is recommended for pregnant women or those who are trying or may become pregnant. Adverse health effects are possible. Reduced lead exposure and increased blood lead monitoring are recommended.
	20-69.9 ug/dL	Adverse health effects are indicated. Medical removal from lead exposure is required by OSHA if blood lead level exceeds 50 ug/dL. Prompt medical evaluation is recommended.
	Greater than 69.9 ug/dL	Critical. Immediate medical evaluation is recommended. Consider chelation therapy when symptoms of lead toxicity are present.
Lead, Blood (Venous)	1	
	Age	Reference
	0-5 years	Less than or equal to 3.4
	6 years or above	Less than or equal to 4.9
Arsenic <u>, Whole</u> Blood	Less than or ec	jual to 12 .0 μg/L
Mercury <u>, Whole</u> Blood	Less than or ec	jual to 10.0 μg/L

Bismuth, <u>Whole</u> Blood	
0099478, BS B	
Specimen Requirements:	
Patient Preparation:	Diet, medication, and nutritional supplements may introduce interfering substances. Patients should be encouraged to discontinue nutritional supplements, vitamins, minerals, and non-essential over-the-counter medications (upon the advice of their physician).
Collect:	Royal blue (K2EDTA) or royal blue (NaHep).
Specimen Preparation:	Transport 3 or 6 mL whole blood in the original collection tube. (Min: 0.5 mL)
Transport Temperature:	Room temperature. Also acceptable: Refrigerated.
Unacceptable Conditions:	Specimens collected in tubes other than royal blue (K2EDTA) or royal blue (NaHep). Specimens transported in containers other than a royal blue (K2EDTA) or royal blue (NaHep) tube or trace element-free transport tube. Clotted specimens.
Remarks:	
Stability:	Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable
Methodology:	Quantitative Inductively Coupled Plasma-Mass Spectrometry (ICP=MS)
Performed:	Sun-Sat
Reported:	1-3 days
Note:	
CPT Codes:	83018
New York DOH Approval Status:	This test is New York DOH approved.

Interpretive Data:

Elevated results may be due to skin<u></u> or collection-related contamination, including the use of <u>tubes</u> <u>that are not certified to be trace element</u> <u>a noncertified metal-free</u><u>-collection/transport tube</u>. If <u>an</u> <u>elevated result is suspected to be due to</u> contamination-concerns exist due to elevated levels of <u>blood bismuth</u>, confirmation with a second specimen collected in a certified <u>trace elementmetal-</u> free tube is recommended.



Methodology: Inductively Coupled Plasma-Mass Spectrometry (ICP-MS).

Reference Interval:

Less than or equal to 5.0 $\mu\text{g/L}$

Jo-1 Antibody, IgG	
0099592, ANTI-JO	
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator <u>tube or red tube</u> tubes.
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 1 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.2 mL)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Plasma or other body fluids.
Remarks:	
Stability:	After separation from the cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)
Methodology:	Semi-Quantitative Multiplex Bead Assay
Performed:	Sun-Sat
Reported:	1-3 days
Note:	Presence of Jo-1 antibody is found in patients with pure polymyositis, pure dermatomyositis, or myositis associated with another rheumatic disease or with interstitial lung disease.
CPT Codes:	86235
New York DOH Approval Status:	This test is New York DOH approved.

Interpretive Data:

Presence of Jo-1 (antihistidyl transfer RNA [t-RNA] synthetase) antibody is associated with polymyositis and may also be seen in patients with dermatomyositis. Jo-1 antibody is associated with pulmonary involvement (interstitial lung disease), Raynaud phenomenon, arthritis, and mechanic's hands (implicated in antisynthetase syndrome).

Component	Interpretation
Jo-1 Antibody,	29 AU/mL or less
igo	AU/mL Equivocal
	41 AU/mL or
	greater Positive



Reference Interval:

Test Number	Components	Reference Interval
	Jo-1 (Histidyl-tRNA Synthetase) Ab, IgG	40 AU/mL or less

Thallium, Whole Blood 0099610, THALB	
Specimen Requirements:	
Patient Preparation:	Diet, medication, and nutritional supplements may introduce interfering substances. Patients should be encouraged to discontinue nutritional supplements, vitamins, minerals, and <u>nonessentialnon-essential</u> over-the-counter medications (upon the advice of their physician).
Collect:	Royal blue (K2EDTA) or <u>r</u> Royal blue (NaHep).
Specimen Preparation:	Transport 6 mL whole blood in the original collection tube. (Min: 0.5 mL)
Transport Temperature:	Room temperature. Also acceptable: Refrigerated.
Unacceptable Conditions:	Specimens collected in tubes other than <u>r</u> Royal blue (K2EDTA) or <u>r</u> Royal blue (NaHep). Specimens transported in containers other than a <u>r</u> Royal blue (K2EDTA) or <u>r</u> Royal blue (NaHep) tube or <u>trace element-free transport tube</u> . <u>Trace Element-Free</u> <u>Transport Tube</u> . Heparin anticoagulant. Clotted specimens.
Remarks:	
Stability:	Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable
Methodology:	Quantitative Inductively Coupled Plasma-Mass Spectrometry (ICP-MS)
Performed:	Sun-Sat
Reported:	1-3 days
Note:	
CPT Codes:	83018
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	
Elevated results may be due to skil noncertified metal-free collection/ levels of blood thallium, confirmati tube is recommended.	n or collection-related contamination, including the use of a transport tube. If contamination concerns exist due to elevated ion with a second specimen collected in a certified metal-free



Blood thallium levels reflect recent exposure as thallium has a biological half-life of approximately 2 to 4 days. Blood levels greater than 100 ugµg/L are considered toxic and greater than 300 ugµg/L indicate severe ingestion. Reported symptoms after After severe thallium poisonings, reported symptoms have varying times of onset and include gastroenteritis, multiorganmulti-organ failure and neurologic injury. Peripheral neuropathy and alopecia are well-documented effects of acute and chronic exposure. Human health effects from low level thallium exposure are unknown.

Elevated results may be due to skin- or collection-related contamination, including the use of tubes that are not certified to be trace element free. If an elevated result is suspected to be due to contamination, confirmation with a second specimen collected in a certified trace element-free tube is recommended.

<u>Methodology: Inductively Coupled Plasma-Mass Spectrometry (ICP-MS).</u> -level thallium exposure are unknown.

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Less than or equal to 2.0 $\mu\text{g/L}$

TEST CHANGE

Cadmium, <u>Whole</u> Blood	
0099675, CADMIUM B	
Specimen Requirements:	
Patient Preparation:	Diet, medication, and nutritional supplements may introduce interfering substances. Patients should be encouraged to discontinue nutritional supplements, vitamins, minerals, non- essential over-the-counter medications (upon the advice of their physician).
Collect:	Royal blue (K2EDTA) or royal blue (NaHep).
Specimen Preparation:	Transport 3 or 6 mL whole blood in the original collection tube. (Min: 0.5 mL)
Transport Temperature:	Room temperature. Also acceptable: Refrigerated.
Unacceptable Conditions:	Specimens collected in tubes other than royal blue (K2EDTA) or royal blue (NaHep). Specimens transported in containers other than royal blue (K2EDTA) or royal blue (NaHep) tube or trace element-free transport tube. Clotted specimens.
Remarks:	
Stability:	Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable
Methodology:	Quantitative Inductively Coupled Plasma-Mass Spectrometry (ICP-MS)
Performed:	Sun-Sat
Reported:	1-3 days
Note:	
CPT Codes:	82300
New York DOH Approval Status:	This test is New York DOH approved.

Interpretive Data:

Elevated results may be due to skin or collection-related contamination, including the use of a noncertified metal-free collection/transport tube. If contamination concerns exist due to elevated levels of blood cadmium, confirmation with a second specimen collected in a certified metal-free tube is recommended.

Blood cadmium levels can be used to monitor acute toxicity and, in combination with cadmium



urine and B-2 microglobulin, is the preferred method for monitoring occupational exposure. Symptoms associated with cadmium toxicity vary based upon route of exposure and may include tubular proteinuria, fever, headache, dyspnea, chest pain, conjunctivitis, rhinitis, sore throat, and cough. Ingestion of cadmium in high concentration may cause vomiting, diarrhea, salivation, cramps, and abdominal pain.

Elevated results may be due to skin- or collection-related contamination, including the use of tubes that are not certified to be trace element free. If an elevated result is suspected to be due to contamination, confirmation with a second specimen collected in a certified trace element-free tube is recommended.

Methodology: Inductively Coupled Plasma-Mass Spectrometry (ICP-MS).

Reference Interval:

Less than or equal to 5.0 μ g/L


Hirsutism Evaluation Panel	
2001763, HIRSUTISM	
Specimen Requirements:	
Patient Preparation:	Collect between 6-10 a.m.
Collect:	Serum separator tube <u>, or serum from plain red</u> -
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 2.5 mL serum or plasma to an ARUP Standard Transport Tube. (Min: 1.5 mL)
Transport Temperature:	Frozen.
Unacceptable Conditions:	Hemolyzed specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 2 hours; Refrigerated: 48 hours; Frozen: 2 months
Methodology:	Quantitative Chemiluminescent Immunoassay (CLIA)/Electrochemiluminescent Immunoassay (ECLIA)/Liquid Chromatography-Tandem Mass Spectrometry
Performed:	Sun-Sat
Reported:	1-5 days
Note:	
CPT Codes:	82157; 82627; 84403; 84270
New York DOH Approval Status:	This test is New York DOH approved.
Internetive Deter	

Interpretive Data:

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.



Test Number	Components	Reference Inte	rval	
	DHEAS			
		Age	Male (ug/dL)	Female (ug/dL)
		0-6 days	108-607	108-607
		7-30 days	32-431	32-431
		1-5 months	3-124	3-124
		6-35 months	0-33	0-29
		3-6 years	0-44	0-47
		7-9 years	5-115	5-94
		10-14 years	22-332	22-255
		15-19 years	88-483	63-373
		20-24 years	211-492	148-407
		25-34 years	160-449	99-340
		35-44 years	89-427	61-337
		45-54 years	44-331	35-256
		55-64 years	52-295	19-205
		65-74 years	34-249	9-246
		75 years and older	16-123	12-154
		Tanner Stage I	7-209	7-126
		Tanner Stage II	28-260	13-241
		Tanner Stage III	39-390	32-446
		Tanner Stage IV & V	81-488	65-371
	Testosterone by Mass Spec			



	Age	Male (ng/dL)	Female (ng/dL)
	Premature (26-28 weeks)	59-125	5-16
	Premature (31-35 weeks)	37-198	5-22
	Newborn	75-400	20-64
	1-5 months	14-363	Less than 20
	6-24 months	Less than 37	Less than 9
	2-3 years	Less than 15	Less than 20
	4-5 years	Less than 19	Less than 30
	6-7 years	Less than 13	Less than 7
	8-9 years	2-8	1-11
	10-11 years	2-165	3-32
	12-13 years	3-619	6-50
	14-15 years	31-733	6-52
	16-17 years	158-826	9-58
	18-39 years	300-1080	9-55
	40-59 years	300-890	9-55
	60 years and older	300-720	5-32
	Premenopausal (18 years and older)	Not Applicable	9-55
	Postmenopausal	Not Applicable	5-32
	Tanner Stage I	2-15	2-17
	Tanner Stage II	3-303	5-40
	Tanner Stage III	10-851	10-63
	Tanner Stage IV-V	162-847	11-62
Testosterone, Free by Mass Spec			



	Age	Male (pg/mL)	Female (pg/mL)
	1-6 years	Less than 0.6	Less than 0.6
	7-9 years	0.1-0.9	0.6-1.8
	10-11	0.1-6.3	0.1-3.5
	12-13	0.5-98.0	0.9-6.8
	14-15	3-138.0	1.2-7.5
	16-17	38.0-173.0	1.2-9.9
	18 years and older	47-244	Not Applicable
	18-30	Not Applicable	0.8-7.4
	31-40	Not Applicable	1.3-9.2
	41-51	Not Applicable	1.1-5.8
	Postmenopausal	Not Applicable	0.6-3.8
	Tanner Stage I	Less than or equal to 3.7	Less than 2.2
	Tanner Stage II	0.3-21	0.4-4.5
	Tanner Stage III	1.0-98.0	1.3-7.5
	Tanner Stage IV	35.0-169.0	1.1-15.5
	Tanner Stage V	41.0-239.0	0.8-9.2
Sex Hormone Binding Globulin			
	Age	Male (nmol/L)	Female (nmol/L)
	1-30 days	13-85	14-60
	31-364 days	70-250	60-215
	1-3 years	50-180	60-190
	4-6 years	45-175	55-170
	7-9 years	28-190	35-170
	10-12 years	23-160	17-155
	13-15 years	13-140	11-120
	16-17 years	10-60	19-145
	18-49 years	17-56	25-122
	50 years and older	19-76	17-125
	Tanner Stage I	26-186	30-173
	Tanner Stage II	22-169	16-127
	Tanner Stage III	13-104	12-98
	Tanner Stage IV	11-60	14-151
	Tanner Stage V	11-71	23-165
Androstenedione by TMS			



	Age	Male (ng/mL)	Female (ng/mL)	
	Premature Infants (26-28 weeks Day 4)	0.92-2.82	0.92-2.82	
	Premature Infants (31-35 weeks Day 4)	0.80-4.46	0.80-4.46	
	Full Term Infants (1-7 days)	0.20-2.90	0.20-2.90	
	8-30 days	0.18-0.80	0.18-0.80	
	1-5 months	0.06-0.68	0.06-0.68	
	6-24 months	0.03-0.15	Less than 0.15	
	2-3 years	Less than 0.11	Less than 0.16	
	4-5 years	0.02-0.17	0.02-0.21	
	6-7 years	0.01-0.29	0.02-0.28	
	8-9 years	0.03-0.30	0.04-0.42	
	10-11 years	0.07-0.39	0.09-1.23	
	12-13 years	0.10-0.64	0.24-1.73	
	14-15 years	0.18-0.94	0.39-2.00	
	16-17 years	0.30-1.13	0.35-2.12	
	18-39 years	0.33-1.34	0.26-2.14	
	40 years and older	0.23-0.89	0.13-0.82	
	Premenopausal	Not Applicable	0.26-2.14	
	Postmenopausal	Not Applicable	0.13-0.82	
	Tanner Stage I	0.04-0.32	0.05-0.51	
	Tanner Stage II	0.08-0.48	0.15-1.37	
	Tanner Stage III	0.14-0.87	0.37-2.24	
	Tanner Stage IV-V	0.27-1.07	0.35-2.05	





Congenital Adrenal Hyperplasia Panel, 11-Beta Hydroxylase Deficiency			
2002282, CAH 11-B HYDROX			
Specimen Requirements:			
Patient Preparation:	Collect between 6-10 a.m.		
Collect:	Serum separator tube or green (sodium or lithium heparin). Also acceptable: Pink (K2EDTA <u>) or serum from plain red.)-</u>		
Specimen Preparation:	Separate serum or plasma from cells ASAP or within 2 hours of collection. Transfer 1.2 mL serum or plasma to an ARUP Standard Transport Tube. (Min: 0.7 mL)		
Transport Temperature:	Refrigerated.		
Unacceptable Conditions:			
Remarks:			
Stability:	Ambient: 24 hours; Refrigerated: 1 week; Frozen: 6 months		
Methodology:	Quantitative High Performance Liquid Chromatography- Tandem Mass Spectrometry		
Performed:	Sun-Sat		
Reported:	1-8 days		
Note:			
CPT Codes:	82157; 83498; 84403; 82634; 82626		
New York DOH Approval Status:	This test is New York DOH approved.		
Interpretive Data:			

Free or bioavailable testosterone measurements may provide supportive information.

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.



Test Number	Components	Reference Inter	rval	
	Testosterone by Mass Spec			
		Age	Male (ng/dL)	Female (ng/dL)
		Premature (26-28 weeks)	59-125	5-16
		Premature (31-35 weeks)	37-198	5-22
		Newborn	75-400	20-64
		1-5 months	14-363	Less than 20
		6-24 months	Less than 37	Less than 9
		2-3 years	Less than 15	Less than 20
		4-5 years	Less than 19	Less than 30
		6-7 years	Less than 13	Less than 7
		8-9 years	2-8	1-11
		10-11 years	2-165	3-32
		12-13 years	3-619	6-50
		14-15 years	31-733	6-52
		16-17 years	158-826	9-58
		18-39 years	300-1080	9-55
		40-59 years	300-890	9-55
		60 years and older	300-720	5-32
		Premenopausal (18 years and older)	Not Applicable	9-55
		Postmenopausal	Not Applicable	5-32
		Tanner Stage I	2-15	2-17
		Tanner Stage II	3-303	5-40
		Tanner Stage III	10-851	10-63
		Tanner Stage IV-V	162-847	11-62
	11-Deoxycortisol, HPLC-MS/MS			



		Age	Male (ng/dL)	Female (ng/dL)	
		Premature (26-28 weeks)	110-1376	110-1376	
		Premature (29-36 weeks)	70-455	70-455	
		Full Term (1-5 months)	10-200	10-200	
		6-11 months	10-276	10-276	
		1-3 years	7-202	7-247	
		4-6 years	8-235	8-291	
		7-9 years	Less than or equal to 120	Less than or equal to 94	
		10-12 years	Less than or equal to 92	Less than or equal to 123	
		13-15 years	Less than or equal to 95	Less than or equal to 107	
	16-17 years	Less than or equal to 106	Less than or equal to 47		
		18 years and older	Less than 50	Less than 33	
		Tanner Stage I	Less than or equal to 105	Less than or equal to 94	
		Tanner Stage II	Less than or equal to 108	Less than or equal to 136	
		Tanner Stage III	Less than or equal to 111	Less than or equal to 99	
		Tanner Stage IV & V	Less than or equal to 83	Less than or equal to 50	
		After metyrapone stimulation	Greater than 8000	Greater than 8000	
	17-Hydroxyprogesterone, HPLC-MS/MS				



	Age	Male (ng/dL)	Female (ng/dL)
	Premature (26-28 weeks)	124-841	124-841
	Premature (29-35 weeks)	26-568	26-568
	Full term Day 3	7-77	7-77
	4 days-30 days	Less than 200	7-106
	1 month-2 months	Less than 200	13-106
	3 months-5 months	3-90	13-106
	6 months-1 year	Less than or equal to 148	Less than or equal to 148
	2-3 years	Less than or equal to 228	Less than or equal to 256
	4-6 years	Less than or equal to 208	Less than or equal to 299
	7-9 years	Less than or equal to 63	Less than or equal to 71
	10-12 years	Less than or equal to 79	Less than or equal to 129
	13-15 years	9-140	9-208
	16-17 years	24-192	Less than or equal to 178
	18 years and older	Less than 139	Less than 207
	Follicular	Not Applicable	15-70
	Luteal	Not Applicable	35-290
	Tanner Stage I	Less than or equal to 62	Less than or equal to 74
	Tanner Stage II	Less than or equal to 104	Less than or equal to 164
	Tanner Stage III	Less than or equal to 151	13-209
	Tanner Stage IV- V	20-173	7-170
Androstenedione by TMS			



	Age	Male (ng/mL)	Female (ng/mL)
	Premature Infants (26-28 weeks Day 4)	0.92-2.82	0.92-2.82
	Premature Infants (31-35 weeks Day 4)	0.80-4.46	0.80-4.46
	Full Term Infants (1-7 days)	0.20-2.90	0.20-2.90
	8-30 days	0.18-0.80	0.18-0.80
	1-5 months	0.06-0.68	0.06-0.68
	6-24 months	0.03-0.15	Less than 0.15
	2-3 years	Less than 0.11	Less than 0.16
	4-5 years	0.02-0.17	0.02-0.21
	6-7 years	0.01-0.29	0.02-0.28
	8-9 years	0.03-0.30	0.04-0.42
	10-11 years	0.07-0.39	0.09-1.23
	12-13 years	0.10-0.64	0.24-1.73
	14-15 years	0.18-0.94	0.39-2.00
	16-17 years	0.30-1.13	0.35-2.12
	18-39 years	0.33-1.34	0.26-2.14
	40 years and older	0.23-0.89	0.13-0.82
	Premenopausal	Not Applicable	0.26-2.14
	Postmenopausal	Not Applicable	0.13-0.82
	Tanner Stage I	0.04-0.32	0.05-0.51
	Tanner Stage II	0.08-0.48	0.15-1.37
	Tanner Stage III	0.14-0.87	0.37-2.24
	Tanner Stage IV-V	0.27-1.07	0.35-2.05
Dehydroepiandrosterone by TMS			



A	I		
Age	Male (ng/mL)	Female (ng/mL)	
Premature	Less than 40	Less than 40	
0-1 days	Less than 11	Less than 11	
2-6 days	Less than 8.7	Less than 8.7	
7 days-1 month	Less than 5.8	Less than 5.8	
1-5 months	Less than 2.9	Less than 2.9	
6-24 months	Less than 2.5	Less than 1.99	
2-3 years	Less than 0.63	Less than 0.85	
4-5 years	Less than 0.95	Less than 1.03	
6-7 years	0.06-1.93	Less than 1.79	
8-9 years	0.10-2.08	0.14-2.35	
10-11 years	0.32-3.08	0.43-3.78	
12-13 years	0.57-4.10	0.89-6.21	
14-15 years	0.93-6.04	1.22-7.01	
16-17 years	1.17-6.52	1.42-9.00	
18-39 years	1.33-7.78	1.33-7.78	
40 years and older	0.63-4.70	0.63-4.70	
Postmenopausal	Not Applicable	0.60-5.73	
Tanner Stage I	0.11-2.37	0.14-2.76	
Tanner Stage II	0.37-3.66	0.83-4.87	
Tanner Stage III	0.75-5.24	1.08-7.56	
Tanner Stage IV-V	1.22-6.73	1.24-7.88	





TEST CHANGE

Dexamethasone, Serum or Plasma by LC-MS/MS 2003248, DEXA TMS			
Specimen Requirements:			
Patient Preparation:	Specimen should be collected between 8-10 a.m.		
Collect:	Serum separator tube, <u>serum from plain red,</u> lavender (EDTA), pink (K2EDTA), or green (sodium or lithium heparin).		
Specimen Preparation:	Separate serum or plasma from cells ASAP or within 2 hours of collection. Transfer 1 mL serum or plasma to an ARUP Standard Transport Tube. (Min: 0.5 mL)		
Transport Temperature:	Refrigerated.		
Unacceptable Conditions:			
Remarks:			
Stability:	After separation from cells: Ambient: <u>24 hoursUnacceptable;</u> Refrigerated: 1 week; Frozen: 6 months		
Methodology:	<u>Quantitative</u> Liquid Chromatography-Tandem Mass Spectrometry		
Performed:	Wed, Sat		
Reported:	2-5 days		
Note:			
CPT Codes:	80299		
New York DOH Approval Status:	This test is New York DOH approved.		
Interpretive Data:			
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.			

Reference Interval:

Adults baseline: Less than 50 ng/dL 8:00 AM draw following 1 mg dexamethasone between 11:00 pm and 12:00 am the previous evening: 140 - 295 ng/dL 8:00 AM draw following 8 mg dexamethasone (4 x 2 mg doses) between 11:00 pm and 12:00 am the previous evening: 1600 - 2850 ng/dL





N-methyl-D-Aspartate Recepto to Titer	r (NMDAR) Antibody, IgG by CBA-IFA, Serum With Reflex
2004221, NMDA IGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube <u>or red tube</u> .
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 1 mL serum to an ARUP standard transport tube. (Min: 0.15 mL)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	CSF or plasma. Contaminated, hemolyzed, or severely lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)
Methodology:	Semi-Quantitative Cell-Based Indirect Fluorescent Antibody
Performed:	Sun-Sat
Reported:	1-3 days
Note:	If NMDA antibody IgG is positive, then an NMDA antibody IgG titer is reported. Additional charges apply.
CPT Codes:	86255; if reflexed, add 86256
New York DOH Approval Status:	This test is New York DOH approved.

Interpretive Data:

NMDA receptor antibody is found in a subset of patients with autoimmune limbic encephalitis and may occur with or without associated tumor. Decreasing antibody levels may be associated with therapeutic response. In addition, positive results have been reported in patients with nonautoimmune phenotypes. A negative test result does not rule out a diagnosis of autoimmune limbic encephalitis. Results should be interpreted in correlation with the patient's clinical history and other laboratory findings. Serum testing should be paired with CSF testing for improved diagnostic sensitivity.

This indirect fluorescent antibody assay utilizes full-length GluN1 transfected cell lines for the detection and semiquantification of NMDA receptor IgG antibody.



Reference Interval:

Less than 1:10

Bilirubin, CSF	
2005248, BILCSF	
Specimen Requirements:	
Patient Preparation:	
Collect:	CSF. Collect at least 12 hours after suspected hemorrhage. Use the last (ideally the fourth) specimen post lumbar puncture. AND serum separator tube.
Specimen Preparation:	CSF: Transport to local lab quickly and avoid pneumatic tube transport if possible. Protect from light during collection, storage, and shipment. Centrifuge for 5 minutes ASAP or within 1 hour of collection. Transfer 2 mL CSF to an ARUP amber transport tube. (Min: 1 mL) Do not freeze. Serum: Protect from light during collection, storage and shipment. Allow specimen to clot completely at room temperature. Separate from cells ASAP or within 2 hours of collection. Transfer 1 mL serum to an ARUP amber transport tube. (Min: 0.5 mL)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Specimens not protected from light. Frozen CSF.
Remarks:	
Stability:	CSF: Ambient: 4 hours; Refrigerated: 1 week; Frozen: Unacceptable Serum: Ambient: 24 hours; Refrigerated: 1 week; Frozen: 6 months
Methodology:	Quantitative Spectrophotometry
Performed:	Sun-Sat
Reported:	Within 24 hours
Note:	
CPT Codes:	84311 x2; 82247
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	
Refer to report.	
Reference Interval:	



Test Number	Components	Reference Interval
	Net Absorbance, Bilirubin, CSF	0.007 AU or less
	Net Absorbance, Oxyhemoglobin, CSF	0.020 AU or less
	Serum Bilirubin	Refer to Report

HOTLINE NOTE: There is a numeric map change associated with this test. Refer to the Hotline Test Mix for interface build information.



Chromatin Antibody, IgG		
2005287, CHROMATIN		
Specimen Requirements:		
Patient Preparation:		
Collect:	Serum separator tube.	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.3 mL)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Urine or plasma. Contaminated, heat-inactivated, severely hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Semi-Quantitative Enzyme-Linked Immunosorbent Assay	
Performed:	<u>Sun, TueMon</u> , Wed, <u>Thu,</u> Fri	
Reported:	1-4 days	
Note:		
CPT Codes:	83516	
New York DOH Approval Status:	This test is New York DOH approved.	
Interpretive Data:		
The presence of anti-chromatin antibodies may be useful in the diagnosis of systemic lunus		

The presence of anti-chromatin antibodies may be useful in the diagnosis of systemic lupus erythematosus (SLE) or drug-induced lupus (DIL) and have been reported to be predictive of lupus nephritis, especially when antibody levels are high.

Component	Interpretation
Chromatin	19 Units or less
Antibody, IgG	Negative 20-60
	Units Moderate
	Positive 61 Units
	Strong Positive

Test	Components	Reference Interval
Number		



Chromatin Antibody, IgG

19 Units or less



Streptococcus pneumoniae Antibodies, IgG (23 Serotypes)

2005779, PNEUMO 23	
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube <u>or red tube</u> . Postimmunization specimen should be drawn 30 days after immunization and, if shipped separately, must be received within 60 days of preimmunization specimen.
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 1.5 mL serum to an ARUP standard transport tube. (Min: 0.25 mL) MARK SPECIMENS CLEARLY AS "PRE" OR "POST" SO SPECIMENS WILL BE SAVED AND TESTED SIMULTANEOUSLY
Transport Temperature:	Refrigerated. "Pre" and "post" pneumococcal vaccine specimens can be submitted separately or together for testing.
Unacceptable Conditions:	Plasma or other body fluids. Contaminated, hemolyzed, or severely lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 60 days (avoid repeated freeze/thaw cycles).
Methodology:	Quantitative Multiplex Chemiluminescent Immunoassay
Performed:	Tue, Fri
Reported:	1-5 days
Note:	
CPT Codes:	86581
New York DOH Approval Status:	This test is New York DOH approved.

Interpretive Data:

A pre- and postvaccination comparison is required to adequately assess the humoral immune response to the pure polysaccharide Pneumovax 23 (PNX) and/or the protein conjugated Prevnar 7 (P7), Prevnar 13 (P13), Prevnar 20 (P20), and Vaxneuvance (V15) Streptococcus pneumoniae vaccines. Prevaccination samples should be collected prior to vaccine administration. Postvaccination samples should be obtained at least 4 weeks after immunization. Testing of postvaccination samples alone will provide only general immune status of the individual to various pneumococcal serotypes.



In the case of pure polysaccharide vaccine, indication of immune system competence is further delineated as an adequate response to at least 50 percent of the serotypes in the vaccine challenge for those 2-5 years of age and to at least 70 percent of the serotypes in the vaccine challenge for those 6-65 years of age. Individual immune response may vary based on age, past exposure, immunocompetence, and pneumococcal serotype.

Responder Status Antibody Ratio

Good responder At least a twofold increase and/or a postvaccination concentration greater than or equal to 1.3 ug/mL

A response to 50-70 percent or more of the serotypes in the vaccine challenge is considered a normal humoral response. (Daly, 2014) Antibody concentration greater than 1.0-1.3 ug/mL is generally considered long-term protection. (Daly, 2015)

References:

1. Daly TM, Pickering JW, Zhang X, et al. Multilaboratory assessment of threshold versus foldchange algorithms for minimizing analytical variability in multiplexed pneumococcal IgG measurements. Clin Vaccine Immunol. 2014;21(7):982-988.

2. Daly TM, Hill HR. Use and clinical interpretation of pneumococcal antibody measurements in the evaluation of humoral immune function. Clin Vaccine Immunol. 2015;22(2):148-152.



Adenovirus, Quantitative PCR	
2007192, ADENO QNT	
Specimen Requirements:	
Patient Preparation:	
Collect:	Lavender (EDTA), pink (K2EDTA), or serum separator tube
Specimen Preparation:	Do not freeze whole blood specimens. Transport 1 mL whole blood, serum or plasma in a sterile container. (Min: 0.5 mL).
Transport Temperature:	Refrigerated
Unacceptable Conditions:	Frozen whole blood. Heparinized specimens.
Remarks:	Specimen source required.
Stability:	Ambient: 24 hours; Refrigerated: 5 days; Frozen: 1 year
Methodology:	Quantitative Real-Time Polymerase Chain Reaction
Performed:	Sun-Sat
Reported:	1-4 days
Note:	The limit of quantification for this DNA assay is 2.3.0 log copies/mL ($2001,000$ copies/mL). If the assay DID NOT DETECT the virus, the test result will be reported as "< 2.3.0 log copies/mL (< $200-1,000$ copies/mL)." If the assay DETECTED the presence of the virus but was not able to accurately quantify the number of copies, the test result will be reported as "Not Quantified."
CPT Codes:	87799
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	
The quantitative range of this assa copies/mL).	y is <u>2.</u> 3 <u>-6.3</u> .0- 7.0 log copies/mL (<u>200-2</u> 1,000-10,000,000
A negative result (less than $2.3-9$ log conjes/mL or less than $2001-000$ conjes/mL) does not rule	

A negative result (less than 2.3.0 log copies/mL or less than 2001,000 copies/mL) does not rule out the presence of PCR inhibitors in the patient specimen or aAdenovirus DNA concentrations below the level of detection of the assay. Inhibition may also lead to underestimation of viral quantitation.



No international standard is currently available for calibration of this assay. Caution should be taken when interpreting results generated by different assay methodologies.

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Not detected



Allergen, Food, Peanut Components IgE		
2007211, PEANUT COM		
Specimen Requirements:		
Patient Preparation:	Multiple patient encounters should be avoided	
Collect:	Serum separator tube (SST). Multiple specimen tubes should be avoided. Include all available specimen.	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer <u>1.0.6</u> mL serum plus 0.1 mL for each additional allergen ordered to an ARUP <u>standard transport tube.</u> Standard <u>Transport Tube.</u> (Min: 0. <u>5</u> 4 mL plus 0.04 mL for each <u>additional</u> allergen ordered). <u>Refer to "Allergen Specimen Collection</u> <u>Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)	
Transport Temperature:	Refrigerated	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun-Sat	
Reported:	1-3 days	
Note:	Test methodology uses solid-phase immunoassays against the whole peanut allergen (f13) and 6 antigenic epitopes (Ara h1, Ara h2, Ara h3, Ara h6, Ara h8, and Ara h9) and measures IgE antibody concentrations in patient serum or plasma. The binding of a specific IgE to an immobilized allergen component is detected by the addition of a secondary fluorescence-labeled anti-human IgE antibody.	
CPT Codes:	86003; 86008 x6	
New York DOH Approval Status:	This test is New York DOH approved.	
Interpretive Data:		



Allergen results of 0.10-0.34 kU/L for whole peanut are intended for specialist use as the clinical relevance is undetermined. Even though increasing ranges are reflective of increasing concentrations of allergen-specific IgE, these concentrations may not correlate with the degree of clinical response or skin testing results when challenged with a specific allergen. The correlation of allergy laboratory results with clinical history and in vivo reactivity to specific allergens is essential. A negative test may not rule out clinical allergy or even anaphylaxis.

Reporting Range (reported in kU/L)	Probability of IgE Mediated Clinical Reaction	Class Scoring
Less than 0.10	No significant level detected	0
0.10 - 0.34	Clinical relevance undetermined	0/1
0.35 - 0.70	Low	1
0.71 - 3.50	Moderate	2
3.51 - 17.50	High	3
17.51 - 50.00	Very high	4
50.01 - 100.00	Very high	5
Greater than 100.00	Very high	6

Test Number	Components	Reference Interval
	Allergen, Food, Peanut IgE	Less than or equal to 0.34 kU/L
	Allergen, Food, Severe Peanut Ara h 1	Less than or equal to 0.09 kU/L
	Allergen, Food, Severe Peanut Ara h 2	Less than or equal to 0.09 kU/L
	Allergen, Food, Severe Peanut Ara h 3	Less than or equal to 0.09 kU/L
	Allergen, Food, Severe Peanut Ara h 9	Less than or equal to 0.09 kU/L
	Allergen, Food, Mild Peanut Ara h 8	Less than or equal to 0.09 kU/L
	Allergen, Food, Severe Peanut Ara h 6	Less than or equal to 0.09 kU/L



Allergens, Food, Extended Panel IgG 2007213, G FOOD PAN		
Specimen Requirements:		
Patient Preparation:		
Collect:	Plain red or serumSerum separator tube (SST).	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>252 mL). Refer to</u> "Allergen Specimen Collection Instructions" at <u>www.aruplab.com/testing/resources/specimen./allergen</u>)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL	
CPT Codes:	86001 x19	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its performance characteristics determined by ARUP Laboratories. It		

has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.



Test Number	Component	Reference Interval
0097636	Allergen, Food, Wheat IgG	Less than 60.20 mcg/mL
0097641	Allergen, Food, Potato White IgG	Less than 6.09 mcg/mL
0097642	Allergen, Food, Rye IgG	Less than 26.71 mcg/mL
0097643	Allergen, Food, Soybean IgG	Less than 5.30 mcg/mL
0097644	Allergen, Food, Tomato IgG	Less than 7.20 mcg/mL
0097647	Allergen, Food, Orange IgG	Less than 8.65 mcg/mL
0097648	Allergen, Food, Peanut IgG	Less than 6.80 mcg/mL
0097649	Allergen, Food, Pork IgG	Less than 7.92 mcg/mL
0097651	Allergen, Food, Lettuce IgG	Less than 11.31 mcg/mL
0097652	Allergen, Food, Malt IgG	Less than 22.31 mcg/mL
0097653	Allergen, Food, Casein (Cow's Milk) IgG	Less than 38.70 mcg/mL.
0097654	Allergen, Food, Oat IgG	Less than 13.30 mcg/mL
0097656	Allergen, Food, Chicken IgG	Less than 6.25 mcg/mL
0097657	Allergen, Food, Chocolate IgG	Less than 20.41 mcg/mL
0097658	Allergen, Food, Corn IgG	Less than 10.50 mcg/mL
0097659	Allergen, Food, Egg White IgG	Less than 15.70 mcg/mL
0097706	Allergen, Food, Baker's Yeast IgG	Less than 11.41 mcg/mL
0097707	Allergen, Food, Barley IgG	Less than 20.31 mcg/mL
0097708	Allergen, Food, Beef IgG	Less than 22.01 mcg/mL



Allergens, Food, Meat Panel IgG			
Specimen Requirements:			
Patient Preparation:			
Collect:	Plain red or serum Serum separator tube (SST)).		
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to</u> <u>"Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)		
Transport Temperature:	Refrigerated.		
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.		
Remarks:			
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year		
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay		
Performed:	Sun		
Reported:	1-8 days		
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL		
CPT Codes:	86001 x3		
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.		
Interpretive Data:			
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.			
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was			

Reference Interval:

performed in a CLIA certified laboratory and is intended for clinical purposes.



Test Number	Component	Reference Interval
0097649	Allergen, Food, Pork IgG	Less than 7.92 mcg/mL
0097656	Allergen, Food, Chicken IgG	Less than 6.25 mcg/mL
0097708	Allergen, Food, Beef IgG	Less than 22.01 mcg/mL



Allergens, Food, Common Panel IgG			
2007215, G FOOD COM			
Specimen Requirements:			
Patient Preparation:			
Collect:	Plain red or serumSerum separator tube (SST).		
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>25</u> 2 mL). <u>Refer to</u> <u>"Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)		
Transport Temperature:	Refrigerated.		
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.		
Remarks:			
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year		
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay		
Performed:	Sun		
Reported:	1-8 days		
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL		
CPT Codes:	86001 x18		
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.		
Interpretive Data:			
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.			
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was			

Reference Interval:

performed in a CLIA certified laboratory and is intended for clinical purposes.



Test Number	Component	Reference Interval
0090284	Allergen, Food, Almond IgG	Less than 15.21 mcg/mL
0090286	Allergen, Food, Banana IgG	Less than 46.11 mcg/mL
0097653	Allergen, Food, Casein (Cow's Milk) IgG	Less than 38.70 mcg/mL
0097656	Allergen, Food, Chicken IgG	Less than 6.25 mcg/mL
0097658	Allergen, Food, Corn IgG	Less than 10.50 mcg/mL
0097659	Allergen, Food, Egg White IgG	Less than 15.70 mcg/mL
0090287	Allergen, Food, Garlic IgG	Less than 12.71 mcg/mL
0090289	Allergen, Food, Gluten IgG	Less than 47.41 mcg/mL
0097654	Allergen, Food, Oat IgG	Less than 13.30 mcg/mL
0097647	Allergen, Food, Orange IgG	Less than 8.65 mcg/mL
0097648	Allergen, Food, Peanut IgG	Less than 6.80 mcg/mL
0097641	Allergen, Food, Potato White IgG	Less than 6.09 mcg/mL
0097323	Allergen, Food, Rice IgG	Less than 16.71 mcg/mL
0097643	Allergen, Food, Soybean IgG	Less than 5.30 mcg/mL
0097644	Allergen, Food, Tomato IgG	Less than 7.20 mcg/mL
0097636	Allergen, Food, Wheat IgG	Less than 60.20 mcg/mL
0097291	Allergen, Food, Whey IgG	Less than 88.61 mcg/mL
0097706	Allergen, Food, Baker's Yeast IgG	Less than 11.41 mcg/mL



Allergens, Food, IgG Panel 2007216. IGG FOOD		
Specimen Requirements:		
Patient Preparation:		
Collect:	Plain red or serum Serum separator tube (SST)).	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.252 mL). Refer to "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:	The units of measure mcg/mL and mgA/L are interchangeable. 1 mg/L = 1000 mcg/1000 mL	
CPT Codes:	86001 x11	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its performance characteristics determined by ARUP Laboratories. It		

has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.



Test Number	Components	Reference Interval
0090289	Allergen, Food, Gluten IgG	Less than 47.41 mcg/mL
0097636	Allergen, Food, Wheat IgG	Less than 60.20 mcg/mL
0097658	Allergen, Food, Corn IgG	Less than 10.50 mcg/mL
0097659	Allergen, Food, Egg White IgG	Less than 15.70 mcg/mL
0097315	Allergen, Food, Egg Yolk IgG	Less than 19.61 mcg/mL
0097706	Allergen, Food, Baker's Yeast IgG	Less than 11.41 mcg/mL
0097657	Allergen, Food, Chocolate IgG	Less than 20.41 mcg/mL
0097648	Allergen, Food, Peanut IgG	Less than 6.80 mcg/mL
0097654	Allergen, Food, Oat IgG	Less than 13.30 mcg/mL
0097323	Allergen, Food, Rice IgG	Less than 16.71 mcg/mL
0097643	Allergen, Food, Soybean IgG	Less than 5.30 mcg/mL



Echinococcus Antibody, IgG 2007220, ECHINO IGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube (SST) or plain red.
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 1 mL serum to an ARUP standard transport tube. (Min: 0.15 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of acute specimens. Mark specimens plainly as acute or convalescent.
Transport Temperature:	Preferred transport temp: Refrigerated. Also acceptable: Frozen
Unacceptable Conditions:	Contaminated, heat-inactivated, grossly hemolyzed, or severely lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 month (avoid repeated freeze/thaw cycles)
Methodology:	Semi-Quantitative Enzyme-Linked Immunosorbent Assay (ELISA)
Performed:	TueMon, Thu
Reported:	1- <u>8</u> 5 days
Note:	
CPT Codes:	86682
New York DOH Approval Status:	This test is New York DOH approved.

Interpretive Data:

Patients with collagen vascular diseases, hepatic cirrhosis, schistosomiasis, and other parasitic infections can produce false-positive results. There is a strong cross-reaction between echinococcosis- and cysticercosis-positive sera.

Seroconversion between acute and convalescent sera is considered strong evidence of recent infection. The best evidence for infection is a significant change on two appropriately timed specimens where both tests are done in the same laboratory at the same time.


Component	Interpretation
Echinococcus	0-8
Antibody IgG	UNegative:
, ,	No significant level of
	Echinococcus IgG
	antibodies detected. 9-11
	UEquivocal:
	Recommend repeat
	testing in 2-4 weeks with
	fresh sample. 12 U or
	greaterPositive: IgG
	antibodies to
	Echinococcus detected,
	indicating current or past
	infection.

Reference Interval:

Test Number	Components	Reference Interval
	Echinococcus Antibody IgG	8 U or less

Iodine, Serum 2007463, IODINESEB	
Specimen Requirements:	
Patient Preparation:	Diet, medication, and nutritional supplements may introduce interfering substances. Patients should be encouraged to discontinue nutritional supplements, vitamins, minerals, nonessential over-the-counter medications for 48 hours (upon the advice of their physician). In addition, the administration of iodine-based contrast media and drugs containing iodine may yield elevated results. During venipuncture, do not use disinfectants (such as Betadine) that contain iodine.
Collect:	Royal blue (no additive).
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfeport 2 mL serum toin an ARUP Trace Element-Free Transport Tube (ARUP supply #43116) available online through eSupply using ARUP Connect(TM) or contact ARUP Client Services at (800-)-522-2787- (Min: 0.5 mL). Do not use utensils (i.e., syringes, needles, or pipettes) in the collection or transfer of the sample, pour directly into transport tube.)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Specimens not received in <u>trace element-freeTrace Element</u> Free transport tubes. Separator tubes and specimens that are not separated from the clot within 2 hours. Serum collected within 48 hours after administration of a gadolinium (Gd) or iodine (I) containing contrast media (may occur with MRI studies). Plasma.
Remarks:	
Stability:	Ambient: 30 days; Refrigerated: 30 days; Frozen: 30 days
Methodology:	Quantitative Inductively Coupled Plasma-Mass Spectrometry
Performed:	Tue, Thu, Sat
Reported:	1-5 days
Note:	
CPT Codes:	83018
New York DOH Approval Status:	This test is New York DOH approved.



Interpretive Data:

Values greater than 250 μ g/L may indicate iodine overload.

Elevated results may be due to skin or collection-related contamination, including the use of a noncertified metal-free collection/transport tube. If contamination concerns exist due to elevated levels of serum iodine, confirmation with a second specimen collected in a certified metal-free tube is recommended.

Reference Interval:

40-92 μg/L

Anti-C1q Antibody, IgG	
2007601, ANTI-C1Q	
Specimen Requirements:	
Patient Preparation:	
Collect:	Serum separator tube.
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.15 mL)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Urine or plasma. Contaminated, heat-inactivated, severely hemolyzed or severely lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)
Methodology:	Semi-Quantitative Enzyme-Linked Immunosorbent Assay
Performed:	<u>Wed</u> Mon
Reported:	1-15 days
Note:	
CPT Codes:	83516

New York DOH Approval Status: This test is New York DOH approved.

Interpretive Data:

The presence of the anti-C1q IgG antibody may be associated with increased risk of lupus nephritis or with systemic lupus erythematosus (SLE) global activity. Anti-C1q antibodies are not specific for SLE; strong clinical correlation with disease is recommended.

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Component	Interpretation
Anti-C1q Antibody, IgG	19 Units or less Negative 20-39 Equivocal 40 Units or greater Positive



Reference Interval:

Test Number	Components	Reference Interval
	Anti-C1q Antibody, IgG	19 Units or less



Phosphatidylserine and Prothrombin Antibody, IgG

Serum separator tube (SST) OR It. blue (sodium citrate)		
Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum (Min: 0.3 mL) OR 0.5 mL citrate plasma (Min: 0.3 mL) to an ARUP standard transport tube.		
Refrigerated. Also acceptable: Frozen.		
Contaminated, heat-inactivated, clots, fibrin, gross red blood cells, severely lipemic, severely hemolyzed, or severely icteric specimens.		
After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)		
Semi-Quantitative Enzyme-Linked Immunosorbent Assay (ELISA)		
<u>Wed</u> Thu		
1-8 days		
83516		
This test is New York DOH approved.		
Elevated and persistent aPS/PT IgG antibody (with or without lupus anticoagulant activity) may serve as a risk marker of thrombotic events in patients with certain autoimmune diseases, including antiphospholipid syndrome (APS) and systemic lupus erythematosus (SLE).		

0-30 Units



Phosphatidylserine and Prothrombin Antibodies, IgG and IgM

TEST CHANGE

2009451, APS/PT PAN Specimen Requirements: Patient Preparation: Serum separator tube (SST) OR lt. blue (sodium citrate) Collect: Specimen Preparation: Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum (Min: 0.3 mL) OR 0.5 mL citrate plasma (Min: 0.3 mL) to an ARUP standard transport tube. Transport Temperature: Refrigerated. Also acceptable: Frozen. Unacceptable Conditions: Contaminated, heat-inactivated, clots, fibrin, gross red blood cells, severely lipemic, severely hemolyzed, or severely icteric specimens. Remarks: After separation from cells: Ambient: 48 hours; Refrigerated: 2 Stability: weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles) Methodology: Semi-Quantitative Enzyme-Linked Immunosorbent Assay (ELISA) Performed: WedThu Reported: 1-8 days Note: CPT Codes: 83516 x2 New York DOH Approval Status: This test is New York DOH approved.

Interpretive Data:

The presence of elevated and persistent aPS/PT IgG and IgM antibodies (with or without lupus anticoagulant activity) may serve as a risk marker of thrombotic events in patients with certain autoimmune diseases, including antiphospholipid syndrome (APS) and systemic lupus erythematosus (SLE). Strong clinical correlation is recommended for isolated IgM aPS/PT antibody.

Reference Interval:

Test Number	Components	Reference Interval
	Phosphatidylserine and Prothrombin IgG	0-30 Units
	Phosphatidylserine and Prothrombin IgM	0-30 Units

ARUP Laboratories | 500 Chipeta Way | Salt Lake City, UT 84108 | 800-522-2787 | aruplab.com





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A nonprofit enterprise of the University of Utah and its Department of Pathology

Effective Date: July 21, 2025

TEST CHANGE

Carbapenem-Resistant Organism Culture

2011418, MC CRO	
Specimen Requirements:	
Patient Preparation:	
Collect:	Rectal swab, respiratory, skin or wound. Before submitting specimen(s), call (800-)-242-2787, ext. 2248, to notify the Bacteriology Laboratory of the number of specimens being shipped and the date of shipment.
Specimen Preparation:	Transport swab in Eswab transport media (ARUP Supply #45877). Available online through eSupply using ARUP Connect(TM) or contact ARUP Client Services at <u>800-522-2787</u> . <u>Respiratory specimens are acceptable in sterile transport</u> <u>containers or Eswabs</u> . (800) 522-2787.
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	
Remarks:	
Stability:	Eswab: Ambient: 48 hours; Refrigerated: 48 hours; Frozen: Unacceptable <u>Respiratory specimen in a sterile container:</u> Ambient: 2 hours; Refrigerated: 24 hours; Frozen: Unacceptable
Methodology:	Culture
Performed:	Sun-Sat
Reported:	1-7 days
Note:	Identification and susceptibility tests are billed separately from culture.
CPT Codes:	87081; Identification CPT codes may vary based on method
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
This test was developed and its p nas not been cleared or approved performed in a CLIA certified labo	erformance characteristics determined by ARUP Laboratories. It by the US Food and Drug Administration. This test was ratory and is intended for clinical purposes.



A nonprofit enterprise of the University of Utah and its Department of Pathology

Effective Date: July 21, 2025

Culture negative for carbapenem-resistant organisms.

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Allergen, Food, Avocado IgG		
2011723, AVOCADOIGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Plain red or serumSerum separator tube (SST)).	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0. <u>25</u> 2 mL). <u>Refer to</u> <u>"Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:	N/A	
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.		
Reference Interval:		

Less than 9.14 mcg/mL





Allergen, Food, Broccoli IgG		
2011725, BROCC IGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Plain red or serum Serum separator tube (SST)).	
Specimen Preparation:	Separate <u>serum</u> from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum or <u>plasma</u> to an ARUP <u>standard</u> <u>transport tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to "Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:		
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL repr antibody.	resent absent or undetectable levels of allergen-specific IgG	
This test was developed and its pe has not been cleared or approved to performed in a CLIA certified labor	rformance characteristics determined by ARUP Laboratories. It by the US Food and Drug Administration. This test was atory and is intended for clinical purposes.	
neierence interval:		
Less than 9.33 mcg/mL		





Allergen, Food, Cashew IgG	
2011727, CASHEW IGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Plain red or serum Serum separator tube (SST)).
Specimen Preparation:	Separate <u>serum</u> from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum or <u>plasma</u> to an ARUP <u>standard</u> <u>transport tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to "Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL repr antibody.	resent absent or undetectable levels of allergen-specific IgG
This test was developed and its pe has not been cleared or approved to performed in a CLIA certified labor	rformance characteristics determined by ARUP Laboratories. It by the US Food and Drug Administration. This test was atory and is intended for clinical purposes.
Reference Interval:	
Less than 7.19 mcg/mL	





Allergen, Food, Cheese Mold Ig	G
2011729, CHSMLD IGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Green (<u>sodium heparin), lavender</u> Sodium Heparin), Lavender (K2EDTA), <u>I</u> Lavender (K3EDTA), <u>p</u> Pink (K2EDTA) <u>plain red</u> or <u>s</u> Serum separator tube (SST <u>)</u> .
Specimen Preparation:	Separate <u>serum</u> from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum or plasma to an ARUP <u>standard</u> <u>transport tube.Standard Transport Tube.</u> (Min: 0. <u>252</u> mL <u>).</u> <u>Refer to "Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL repr antibody.	resent absent or undetectable levels of allergen-specific IgG
This test was developed and its pe has not been cleared or approved b performed in a CLIA certified labora	rformance characteristics determined by ARUP Laboratories. It by the US Food and Drug Administration. This test was atory and is intended for clinical purposes.

Reference Interval:

Less than 85.57 mcg/mL





Allergen, Food, Clam IgG	
2011731, CLAM IGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Plain red or serum Serum separator tube (SST)).
Specimen Preparation:	Separate <u>serum</u> from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum or <u>plasma</u> to an ARUP <u>standard</u> <u>transport tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to "Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.	
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.	
Reference Interval:	
Less than 12.81 mcg/mL	





Allergen, Food, Coconut IgG	
2011733, COCONUTIGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Plain red or serum Serum separator tube (SST)).
Specimen Preparation:	Separate <u>serum</u> from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum or <u>plasma</u> to an ARUP <u>standard</u> <u>transport tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to "Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.	
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.	
Less than 6.30 mcg/mL	





Allergen, Food, Crab IgG	
2011735, CRAB IGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Plain red or serum Serum separator tube (SST)).
Specimen Preparation:	Separate <u>serum</u> from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum or <u>plasma</u> to an ARUP <u>standard</u> <u>transport tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to "Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.	
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.	
Reference Interval:	
Less than 9.13 mcg/mL	





Allergen, Food, Lobster IgG	
2011737, LOBSTERIGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Plain red or serum Serum separator tube (SST)).
Specimen Preparation:	Separate <u>serum</u> from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum or <u>plasma</u> to an ARUP <u>standard</u> <u>transport tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to "Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.	
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.	
Reterence Interval:	
Less than 7.29 mcg/mL	





Allergen, Food, Oyster IgG	
2011739, OYSTER IGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Plain red or serum Serum separator tube (SST).
Specimen Preparation:	Separate <u>serum</u> from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum or <u>plasma</u> to an ARUP <u>standard</u> <u>transport tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to "Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.	
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.	
Reference Interval:	
Less than 13.42 mcg/mL	





Allergen, Food, Pineapple IgG	
2011741, PNAPPL IGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Plain red or serum Serum separator tube (SST)).
Specimen Preparation:	Separate <u>serum</u> from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum or <u>plasma</u> to an ARUP <u>standard</u> <u>transport tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to "Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL repr antibody.	resent absent or undetectable levels of allergen-specific IgG
This test was developed and its pe has not been cleared or approved to performed in a CLIA certified labore	rformance characteristics determined by ARUP Laboratories. It by the US Food and Drug Administration. This test was atory and is intended for clinical purposes.
Reterence Interval:	
Less than 12.48 mcg/mL	





Allergen, Food, Scallop IgG	
2011743, SCALLOPIGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Plain red or serum Serum separator tube (SST)).
Specimen Preparation:	Separate <u>serum</u> from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum or <u>plasma</u> to an ARUP <u>standard</u> <u>transport tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to "Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.	
This test was developed and its pe has not been cleared or approved b performed in a CLIA certified labora	rformance characteristics determined by ARUP Laboratories. It by the US Food and Drug Administration. This test was atory and is intended for clinical purposes.
Reference Interval:	
Less than 10.99 mcg/mL	





Allergen, Food, Shrimp IgG 2011745, SHRIMP IGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Plain red or serum Serum separator tube (SST)).
Specimen Preparation:	Separate <u>serum</u> from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum or <u>plasma</u> to an ARUP <u>standard</u> <u>transport tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to "Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.	
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.	
Reference Interval:	
Less than 10.41 mcg/mL	





Allergen, Food, Strawberry IgG	
2011747, STRWBRYIGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	Plain red or serum Serum separator tube (SST)).
Specimen Preparation:	Separate <u>serum</u> from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum <u>or plasma</u> to an ARUP <u>standard</u> <u>transport tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to "Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Values less than 2.00 mcg/mL repr antibody.	resent absent or undetectable levels of allergen-specific IgG
This test was developed and its per has not been cleared or approved b performed in a CLIA certified labora	rformance characteristics determined by ARUP Laboratories. It by the US Food and Drug Administration. This test was atory and is intended for clinical purposes.
Reference Interval:	
Less than 12.11 mcg/mL	




Allergen, Food, Tuna IgG		
2011749, TUNA IGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Plain red or serum Serum separator tube (SST)).	
Specimen Preparation:	Separate <u>serum</u> from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum or <u>plasma</u> to an ARUP <u>standard</u> <u>transport tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to "Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:		
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.		
Less than 3.76 mcg/mL		





Allergen, Food, Turkey IgG		
2011751, TURKEY IGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Plain red or serum Serum separator tube (SST).	
Specimen Preparation:	Separate <u>serum</u> from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum or <u>plasma</u> to an ARUP <u>standard</u> <u>transport tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to "Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:		
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.		
Loss than 6 15 mag/ml		
Less than 6.15 mcg/mL		





Allergen, Food, Walnut IgG		
2011753, WALNUT IGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Plain red or serumSerum separator tube (SST)).	
Specimen Preparation:	Separate <u>serum</u> from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum or <u>plasma</u> to an ARUP <u>standard</u> <u>transport tube.Standard Transport Tube.</u> (Min: 0.252 mL). <u>Refer to "Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:		
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.		
Reference Interval:		
Less than 6.84 mcg/mL		





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TEST CHANGE

Chikungunya	Antibody, IgG	
2011808, CHIK	G	
Specimen Req	uirements:	
Patient Prep	paration:	
Collect:		<u>Serumseparator tube (SST). Also acceptable: Plain red</u> Serum or plasma (heparin, citrate, or EDTA <u>).</u> }
Specimen P	reparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 1 mL serum to an ARUP <u>standard transport</u> <u>tubeStandard Transport Tube</u> . (Min: 0.15 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of the acute specimens. Mark specimens plainly as "acute or convalescent."
Transport T	emperature:	Refrigerated.
Unacceptab	le Conditions:	Contaminated, heat-inactivated, hemolyzed, or severely lipemic specimens.
Remarks:		
Stability:		After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)
Methodology:		Semi-Quantitative Enzyme-Linked Immunosorbent Assay
Performed:		Wed
Reported:		1-8 days
Note:		
CPT Codes:		86790
New York DOH Approval Status:		This test is New York DOH approved.
Interpretive Data:		
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.		
Reference Inte	rval:	
0.79 Index or less	Negative: No significant level of Chikungunya IgG antibody detected.	

Deleted Cells

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0.80-1.09 Index	Equivocal:	
	Questionable	
	presence of	
	Chikungunya IgG	
	antibody	
	detected. Repeat	
	testing in 10-14	
	days may be	
	helpful.	
1 10 Index or	Positive [.]	
greater	Chikungunya lgG	
5	antibody	
	detected:	
	suggests current	
	or past infection.	
	or past infection.	

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TEST CHANGE

Chikungunya Antibody, IgM		
Specimen Requirements:		
Patient Preparation:		
Collect:	<u>Serumseparator tube (SST). Also acceptable: Plain red</u> Serum or plasma (heparin, citrate, or EDTA).	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 1 mL serum to an ARUP <u>standard transport</u> <u>tubeStandard Transport Tube</u> . (Min: 0.15 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of the acute specimens. Mark specimens plainly as "acute or convalescent."	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Contaminated, heat-inactivated, hemolyzed, or severely lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)	
Methodology:	Semi-Quantitative Enzyme-Linked Immunosorbent Assay	
Performed:	Wed	
Reported:	1-8 days	
Note:		
CPT Codes:	86790	
New York DOH Approval Status:	This test is New York DOH approved.	
Interpretive Data:		
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.		
Reference Interval:		
0.79 Index or less Negative: No significant level of Chikungunya IgM antibody detected.		

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Deleted Cells



Effective Date: July 21, 2025

0.80-1.09 Index	Equivocal: Questionable presence of Chikungunya IgM antibody detected. Repeat testing in 10-14 days may be helpful.
1.10 Index or greater	Positive: Chikungunya IgM antibody detected.

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Effective Date: July 21, 2025

TEST CHANGE

Chikungunya Antibodies, IgG and IgM 2011812, CHIKPAN

Specimen Requirements:	
Patient Preparation:	
Collect:	Serumseparator tube (SST). Also acceptable: Plain redSerum or plasma (heparin, citrate, or EDTDA).
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 1 mL serum to an ARUP <u>standard transport</u> <u>tubeStandard Transport Tube</u> . (Min: 0.15 mL) Parallel testing is preferred and convalescent specimens must be received within 30 days from receipt of the acute specimens. Mark specimens plainly as "acute or convalescent."
Transport Temperature:	Refrigerated
Unacceptable Conditions:	Contaminated, heat-inactivated, hemolyzed, or severely lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)
Methodology:	Semi-Quantitative Enzyme-Linked Immunosorbent Assay
Performed:	Wed
Reported:	1-8 days
Note:	
CPT Codes:	86790 x2
New York DOH Approval Status:	This test is New York DOH approved.
Internetive Deter	

Interpretive Data:

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Deleted Cells

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Effective Date: July 21, 2025

Test Code	Component	Reference Interval	
2011808 Chikungunya Antibody, IgG	0.79 Index or less	Negative: No significant level of Chikungunya IgG antibody detected.	
		0.80-1.09 Index	Equivocal: Questionable presence of Chikungunya IgG antibody detected. Repeat testing in 10-14 days may be helpful.
	1.10 Index or greater	Positive: Chikungunya IgG antibody detected; suggests current or past infection.	
2011810 Chikungunya Antibody, IgM	0.79 Index or less	Negative: No significant level of Chikungunya IgM antibody detected.	
		0.80-1.09 Index	Equivocal: Questionable presence of Chikungunya IgM antibody detected. Repeat testing in 10-14 days may be helpful.
		1.10 Index or greater	Positive: Chikungunya IgM antibody detected.

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Allergen, Food, Olives IgG	
2011815, OLIVES IGG	
Specimen Requirements:	
Patient Preparation:	N/A
Collect:	Plain red or serumSerum separator tube (SST).
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum or plasma to an ARUP standard transport tube.Standard Transport Tube. (Min: 0.252 mL). Refer to "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	N/A
CPT Codes:	86001
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	

Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

Less than 21.61 mcg/mL





Allergen, Food, Cheddar Cheese IgG 2011817, CHEDCHEESE		
Specimen Requirements:		
Patient Preparation:	N/A	
Collect:	Plain red or serum Serum separator tube (SST).	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.252 mL). Refer to <u>"Allergen Specimen Collection Instructions" at</u> <u>www.aruplab.com/testing/resources/specimen.</u>)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:	N/A	
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.		
Reference Interval:		

Less than 46.23 mcg/mL





Allergen, Food, Whole Egg, IgG		
2011819, WHOLE EGG		
Specimen Requirements:		
Patient Preparation:		
Collect:	Plain red or serum Serum separator tube (SST)).	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum or plasma to an ARUP standard transport tube.Standard Transport Tube. (Min: 0.252 mL). Refer to "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun	
Reported:	1-8 days	
Note:		
CPT Codes:	86001	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Values less than 2.00 mcg/mL represent absent or undetectable levels of allergen-specific IgG antibody.		
This test was developed and its per has not been cleared or approved b performed in a CLIA certified labora Reference Interval:	rformance characteristics determined by ARUP Laboratories. It by the US Food and Drug Administration. This test was atory and is intended for clinical purposes.	
Less than 30.21 mcg/mL		





TEST CHANGE

Dihydropyrimidine Dehydrogena	ase (DPYD)
2012166, DPYD	
Specimen Requirements:	
Patient Preparation:	
Collect:	Lavender (EDTA), pink (K2EDTA), or yellow (ACD solution A or B).
Specimen Preparation:	Transport 3 mL whole blood. (Min: 1 mL)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Plasma or serum. Heparinized specimens. Frozen specimens in glass collection tubes.
Remarks:	
Stability:	Ambient: 72 hours; Refrigerated: 1 week; Frozen: 1 month.
Methodology:	Polymerase Chain Reaction (PCR)/Fluorescence Monitoring
Performed:	Varies
Reported:	5-10 days
Note:	
CPT Codes:	81232
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	
Refer to report.	
Reference Interval:	



Maternal Serum Screen, Alpha Fetoprotein, hCG, Estriol, and Inhibin A (Quad)		
3000143, MS QUAD		
Specimen Requirements:		
Patient Preparation:	Specimen must be drawn between 14 weeks, 0 days and 24 weeks, 6 days gestation. The recommended time for maternal serum screening is 16 to 18 weeks gestation.	
Collect:	Serum Separator Tube (SST) or Plain Red.	
Specimen Preparation:	Separate from cells ASAP or within 2 hours of collection. Transfer 3 mL serum to an ARUP Standard Transport Tube. (Min: 1 mL)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Plasma. Hemolyzed specimens.	
Remarks:	Submit with Order: Patient's date of birth, current weight, due date, dating method (US, LMP), number of fetuses present, patient's race, if the patient was diabetic at the time of conception, if there is a known family history of neural tube defects, if the patient has had a previous pregnancy with a trisomy, if the patient is currently smoking, if the patient is taking valproic acid or carbamazepine (Tegretol), if this is a repeat sample, and the age of the egg donor if in vitro fertilization.	
Stability:	After separation from cells: Ambient: 72 hours; Refrigerated: 2 weeks; Frozen: 1 year (Avoid repeated freeze/thaw cycles.)	
Methodology:	Quantitative Chemiluminescent Immunoassay	
Performed:	Sun-Sat	
Reported:	2-3 days	
Note:	This test is used to screen for fetal risk of Down syndrome (trisomy 21), trisomy 18, and Open Neural Tube Defect (ONTD, spina bifida).	
CPT Codes:	81511	
New York DOH Approval Status:	This test is New York DOH approved.	
Interpretive Data:		



Refer to report.

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

By report



Maternal Serum Screen, Alpha Fetoprotein 3000144, MS AFP		
Specimen Requirements:		
Patient Preparation:	Specimen must be drawn between 14 weeks, 0 days and 24 weeks, 6 days gestation.	
Collect:	Serum Separator Tube (SST) or Plain Red.	
Specimen Preparation:	Separate from cells ASAP or within 2 hours of collection. Transfer 1 mL serum to an ARUP Standard Transport Tube. (Min: 0.5 mL)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Plasma. Hemolyzed specimens.	
Remarks:	Submit with Order: Patient's date of birth, current weight, due date, dating method (US, LMP), number of fetuses present, patient's race, if the patient was diabetic at the time of conception, if there is a known family history of neural tube defects, if the patient is currently smoking, if the patient is taking valproic acid or carbamazepine (Tegretol), if this is a repeat sample, and the age of the egg donor if an in vitro fertilization.	
Stability:	After separation from cells: Ambient: 72 hours; Refrigerated: 2 weeks; Frozen: 1 year (Avoid repeated freeze/thaw cycles.)	
Methodology:	Quantitative Chemiluminescent Immunoassay (CLIA)	
Performed:	Sun-Sat	
Reported:	2-3 days	
Note:	This test is used to screen for fetal risk of Open Neural Tube Defect (i.e., spina bifida).	
CPT Codes:	82105	
New York DOH Approval Status:	This test is New York DOH approved.	
Interpretive Data:		
Refer to report.		



This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

By report



Maternal Serum Screen, First Trimester, hCG, PAPP-A, NT		
3000145, MS FTS		
Specimen Requirements:		
Patient Preparation:	Specimen must be drawn between 11 weeks, 0 days and 13 weeks, 6 days gestation. (Crown-rump length (CRL) must be between 43-83.9 mm at time of specimen collection.)	
Collect:	Serum separator tube (SST) or plain red.	
Specimen Preparation:	Separate from cells ASAP or within 2 hours of collection. Transfer 3 mL serum to an ARUP standard transport tube. (Min: 1 mL)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Plasma. Hemolyzed specimens.	
Remarks:	Submit with Order: Patient's date of birth, current weight, number of fetuses present, patient's race, if the patient has had a previous pregnancy with a trisomy, if the patient is currently smoking, if this is a repeat sample, and the age of the egg donor if in vitro fertilization. In addition to the above: the date of ultrasound, the CRL measurement, the nuchal translucency (NT) measurement ,and the name and certification number of the sonographer is required. NT must be measured when the CRL is between 38-83.9 mm. The NT measurement must also be performed by an ultrasonographer that is certified by the Fetal Medicine Foundation (FMF). To avoid possible test delays for an ultrasonographer that is new to our database, please contact a genetic counselor at 800-242-2787 extension 2141 prior to sending specimen. If an NT is unobtainable, order Maternal Serum Screening, Integrated (ARUP test codes 3000147 (collect in first trimester) and 3000149 (collect in second trimester), which can be interpreted without an NT value.	
Stability:	After separation from cells: Ambient: 72 hours; Refrigerated: 2 weeks; Frozen: 1 year (Avoid repeated freeze/thaw cycles.)	
Methodology:	Quantitative Chemiluminescent Immunoassay (CLIA)	
Performed:	Sun-Sat	
Reported:	2-4 days	



Note:	This test does not screen for open neural tube defect (ONTD). This test is used to screen for fetal risk of Down syndrome (trisomy 21) and trisomy 18.
CPT Codes:	81508
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	
Refer to report.	
Reference Interval:	
By report	



Maternal Screening, Sequential, Specimen #1, hCG, PAPP-A, NT		
3000146, MS SEQ1		
Specimen Requirements:		
Patient Preparation:	Specimen must be drawn between 11 weeks, 0 days and 13 weeks, 6 days gestation. (Crown-rump length (CRL) must be between 43-83.9 mm at time of specimen collection.)	
Collect:	Serum separator tube (SST) or plain red.	
Specimen Preparation:	Separate from cells ASAP or within 2 hours of collection. Transfer 3 mL serum to an ARUP standard transport tube. (Min: 1 mL)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Plasma. Hemolyzed specimens.	
Remarks:	Submit with Order: Patient's date of birth, current weight, number of fetuses present, patient's race, if the patient was diabetic at the time of conception, if there is a known family history of neural tube defects, if the patient has had a previous pregnancy with a trisomy, if the patient is currently smoking, if the patient is taking valproic acid or carbamazepine (Tegretol), if this is a repeat sample, and the age of the egg donor if in vitro fertilization. In addition to the above: the date of ultrasound, the CRL measurement, the nuchal translucency (NT) measurement, and the name and certification number of the sonographer is required. NT must be measured when the CRL is between 38-83.9 mm. The NT measurement must also be performed by an ultrasonographer that is certified by the Fetal Medicine Foundation (FMF). To avoid possible test delays for an ultrasonographer that is new to our database, please contact a genetic counselor at 800-242-2787 <u>ext.extension</u> 2141 prior to sending specimen. If an NT is unobtainable, order Maternal Serum Screening, Integrated (ARUP test codes 3000147 (collect in first trimester) and 3000149 (collect in second trimester) <u>)</u> , which can be interpreted without an NT value.	
Stability:	After separation from cells: Ambient: 72 hours; Refrigerated: 2 weeks; Frozen: 1 year (Avoid repeated freeze/thaw cycles.)	
Methodology:	Quantitative Chemiluminescent Immunoassay (CLIA)	
Performed:	Sun-Sat	



Reported:	2-4 days
Note:	The first specimen of a sequential maternal serum screening is used to measure PAPP-A and hCG. This test is used to screen for fetal risk of Down syndrome (trisomy 21) and trisomy 18. Final interpretative report, which also includes fetal risk for open neural tube defect (ONTD), will be available when the second specimen test results are complete.
CPT Codes:	81508
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	
Refer to report.	
Reference Interval:	
By report	



Maternal Serum Screening, Integrated, Specimen #1, PAPP-A, NT		
3000147, MS INT1		
Specimen Requirements:		
Patient Preparation:	Specimen must be drawn between 10 weeks, 0 days and 13 weeks, 6 days gestation. (If gestational age is based on crown-rump length (CRL), the specimen must be collected when the CRL is between 32.4-83.9 mm.)	
Collect:	Serum separator tube (SST) or plain ped.	
Specimen Preparation:	Separate from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP standard transport tube. (Min: 0.3 mL)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Plasma. Hemolyzed specimens.	
Remarks:	Submit with Order: Patient's date of birth, current weight, number of fetuses present, patient's race, if the patient was diabetic at the time of conception, if there is a known family history of neural tube defects, if the patient has had a previous pregnancy with a trisomy, if the patient is currently smoking, if the patient is taking valproic acid or carbamazepine (Tegretol), if this is a repeat sample, and the age of the egg donor if in vitro fertilization. In addition to the above, if a NT measurement is performed: the date of ultrasound, the CRL measurement, the nuchal translucency (NT) measurement, and the name and certification number of the sonographer is required. NT must be measured when the CRL is between 38-83.9 mm. or If no NT measurement is performed a due date or CRL measurement with the date of ultrasound is required. The NT measurement must also be performed by an ultrasonographer that is certified by the Fetal Medicine Foundation (FMF). To avoid possible test delays for an ultrasonographer that is new to our database, please contact a genetic counselor at 800-242-2787 <u>ext.extension-</u> 2141 prior to sending specimen.	
Stability:	After separation from cells: Ambient: 72 hours; Refrigerated: 2 weeks; Frozen: 1 year (Avoid repeated freeze/thaw cycles.)	
Methodology:	Quantitative Chemiluminescent Immunoassay (CLIA)	
Performed:	Sun-Sat	



Reported:	2-4 days
Note:	The first specimen of an integrated maternal serum screening is used to measure PAPP-A. Final interpretative report will be available when the second specimen test results are complete.
CPT Codes:	84163
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	
Refer to report.	
Reference Interval:	
By report	



Aspergillus fumigatus Antibody IgG	
3000876, ASPERF IGG	
Specimen Requirements:	
Patient Preparation:	
Collect:	<u>Plain red or serum</u> Serum separator tube (SST <u>)</u> .
Specimen Preparation:	Separate from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP standard transport tube. (Min: 0.25 mL). Refer to "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen. 2-mL)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Postmortem samples
Remarks:	
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay
Performed:	Sun
Reported:	1-8 days
Note:	
CPT Codes:	86317
New York DOH Approval Status:	This test is New York DOH approved.

Interpretive Data:

A positivity cutoff of 40 μ g/mL was established based on a comparison study to precipitin assay results. Note that elevated specific IgG concentrations to Aspergillus fumigatus are not disease specific and could be found in healthy individuals. Results must be interpreted in the context of patient's clinical, laboratory, and radiologic findings, and in concordance with current practice guidelines.

Reference Interval:

Test Number	Components	Reference Interval
	Aspergillus fumigatus Ab IgG	Less than or equal to 40.0 $\mu\text{g/mL}$





Hereditary Hemolytic Anemia Cascade 3000894. HHACASCADE		
Specimen Requirements:		
Patient Preparation:		
Collect:	3 whole blood Lavender (K2EDTA) or Pink (K2EDTA) specimens and 3-5 peripheral blood smears.	
Specimen Preparation:	Transfer specimens using ARUP kit (ARUP supply # 54388) available online through eSupply using ARUP Connect or contact ARUP Client Services at (800) 522-2787.	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:		
Remarks:	Submit with Order: Patient history form, including information from a recent CBC, is required for interpretation.	
Stability:	Ambient: Unacceptable; Refrigerated: 1 week; Frozen: Unacceptable	
Methodology:	High Performance Liquid Chromatography (HPLC)/Electrophoresis/RBC Solubility/Polymerase Chain Reaction/Fluorescence Resonance Energy Transfer/Sequencing/Spectrophotometry/Visual Identification/Quantitative Enzymatic Assay/Quantitative Flow Cytometry/Cytochemical Stain/Multiplex Ligation-Dependent Probe Amplification/Massively Parallel Sequencing	
Performed:	Sun-Sat	
Reported:	Varies	
Note:	The Hereditary Hemolytic Anemia Cascade begins with initial standard tests to detect possible causes of hemolytic anemia. If the results of the initial tests are suggestive of an abnormal or unstable hemoglobin, RBC membrane instability, or an enzyme or protein deficiency; or if the CBC data is suggestive of a hemoglobinopathy, appropriate testing will be performed at an additional charge. Depending on findings, one or more reflex tests may be required in order to provide a clinical interpretation. Tests added may include electrophoresis, solubility testing, mutational analysis, and/or sequencing. Quantitation of hemoglobin by HPLC or electrophoresis is most definitive in individuals one year and older. If quantitation of	



		hemoglobin was recommended. <i>A</i> additional testin	performed before age one, repeat testing is bnormal hemoglobin variants may require g, which increases TAT up to 21 days.	
CPT Coo	les:	84220; 88184; 8 separately. Addir 85007; 83068; 8 81405; 85660; 8	2955; 83021. Reflex components billed tional CPT codes may apply: 85555; 85060; 1269; 81259; 81363; 81364; 81249; 81404; 3020; 81479.	
New York DOH Approval Status:		Specimens from York DOH approv	New York clients will be sent out to a New /ed laboratory, if possible.	
Interpretive Data:				
Reference Interval:				
Test Number	Components		Reference Interval	

CYP3A4 and CYP3A5				
3001518, 3A4/3A5				
Specimen Requirements:				
Patient Preparation:				
Collect:	Lavender (EDTA), pink (K2EDTA), or yellow (ACD solution A or B).			
Specimen Preparation:	Transport 3 mL whole blood. (Min: 1 mL)			
Transport Temperature:	Refrigerated.			
Unacceptable Conditions:	Plasma or serum. Specimens collected in sodium heparin or lithium heparin. Frozen specimens in glass collection tubes.			
Remarks:				
Stability:	Ambient: 72 hours; Refrigerated: 1 week; Frozen: 1 month			
Methodology:	Polymerase Chain Reaction (PCR)/Fluorescence Monitoring			
Performed:	Varies			
Reported:	5-10 days			
Note:				
CPT Codes:	81230; 81231			
New York DOH Approval Status:	This test is New York DOH approved.			
Interpretive Data:				
Refer to report				
Counseling and informed consent are recommended for genetic testing. Consent forms are available online.				
Reference Interval:				
By report				



Cytochrome P450 Genotyping Panel					
3001524, CYP PANEL					
Specimen Requirements:					
Patient Preparation:					
Collect:	Lavender (K2EDTA), pink (K2EDTA), or yellow (ACD solution A or B).				
Specimen Preparation:	Transport 3 mL whole blood. (Min: 1 mL)				
Transport Temperature:	Refrigerated.				
Unacceptable Conditions:	Plasma or serum. Specimens collected in sodium heparin or lithium heparin. Frozen specimens in glass collection tubes.				
Remarks:					
Stability:	Ambient: 72 hours; Refrigerated: 1 week; Frozen: 1 month				
Methodology:	Polymerase Chain Reaction (PCR)/Fluorescence Monitoring/Sequencing				
Performed:	Varies				
Reported:	5-10 days				
Note:	If long-range PCR/duplication testing is performed, additional charges will apply. Approximately less than 5% of samples require 2D6 copy number determination.				
CPT Codes:	81225; 81226; 81227; 81230; 81231; 81479; if reflexed, add 81479				
New York DOH Approval Status:	This test is New York DOH approved.				
Interpretive Data:					
Refer to report.					
Couseling and informed consent are recommended for genetic testing. Consent forms are available online.					
Reference Interval:					

By report

HOTLINE NOTE: There is a component change associated with this test. One or more components



have been added or removed. Refer to the Hotline Test Mix for interface build information.


Troponin T (cTnT) 5th Generation			
3001831, CTNT			
Specimen Requirements:			
Patient Preparation:			
Collect:		Plasma separator tube (PST) or green (lithium heparin)	
Specimen Preparation:		Separate from cells ASAP or within 2 hours of collection. Transfer 1 mL plasma to an ARUP standard transport tube. (Min: 0.5 mL)	
Transport Tem	perature:	CRITICAL FROZEN. Separate specimens must be submitted when multiple tests are ordered.	
Unacceptable Conditions:		Specimens collected in lavender or pink (K2EDTA), lavender (K3EDTA), potassium oxalate, sodium fluoride, or sodium citrate. Grossly hemolyzed specimens.	
Remarks:			
Stability:		Ambient: Unacceptable; Refrigerated: 24 hours; Frozen: 1 year	
Methodology:		Electrochemiluminescent Immunoassay (ECLIA)	
Performed:		<u>Sun-Sat</u> Mon, Wed, Fri	
Reported:		Within 24 hours 1-4 days	
Note:			
CPT Codes:		84484	
New York DOH Approval Status:		This test is New York DOH approved.	
Interpretive Data:			
Reference Interval:			
Female Les equ Male Les	ss than or ual to 10 ng/L ss than or		

equal to 15 ng/L





TEST CHANGE

Anti-gp210 Antibody, IgG		
3002477, GP210 AB		
Specimen Requirements:		
Patient Preparation:		
Collect:	Serum <u>separator tube</u> Separator Tube (SST).	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to ARUP <u>standard transport</u> <u>tube.</u> Standard Transport Tube. (Min: 0.1 mL)	
Transport Temperature:	Refrigerated	
Unacceptable Conditions:	NonserumNon-serum, heat-inactivated, contaminated, grossly icteric, severely lipemic, grossly hemolyzed specimens, or inclusion of fibrin clot.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 month (avoid repeated freeze/thaw cycles)	
Methodology:	Semi-Quantitative Enzyme-Linked Immunosorbent Assay	
Performed:	<u>Thu</u> Wed	
Reported:	1-8 days	
Note:		
CPT Codes:	83516	
New York DOH Approval Status:	This test is New York DOH approved.	

Interpretive Data:

GP210 IgG antibodies can be detected in patients with primary biliary cholangitis (PBC) and may be of diagnostic relevance in a subset of patients with PBC who are negative for anti-mitochondrial antibodies (AMA). These antibodies have a relatively low sensitivity with excellent specificity for PBC. A negative result does not rule out PBC.

Component Interpretation Anti-gp210 20.1 Units or less Negative 20.1-Antibody, IgG 24.9 Units Equivocal 25.0 Units or greater Positive

Reference Interval:



Test Number	Components	Reference Interval
	Anti-gp210 Antibody, IgG	24.9 Units or less



TEST CHANGE

Anti-sp100 Antibody, IgG		
3002478, SP100 AB		
Specimen Requirements:		
Patient Preparation:		
Collect:	Serum <u>separator tube</u> Separator Tube (SST).	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to ARUP <u>standard transport</u> <u>tube.</u> Standard Transport Tube. (Min: 0.1 mL)	
Transport Temperature:	Refrigerated	
Unacceptable Conditions:	Non-Serum, heat-inactivated, contaminated, grossly icteric, severely lipemic, grossly hemolyzed specimens, or inclusion of fibrin clot.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 month (avoid repeated freeze/thaw cycles)	
Methodology:	Semi-Quantitative Enzyme-Linked Immunosorbent Assay	
Performed:	<u>Thu</u> Wed	
Reported:	1-8 days	
Note:		
CPT Codes:	83516	
New York DOH Approval Status:	This test is New York DOH approved.	

Interpretive Data:

SP100 IgG antibodies can be detected in patients with primary biliary cholangitis (PBC) and may be of diagnostic relevance in a subset of patients with PBC who are negative for anti-mitochondrial antibodies (AMA). These antibodies have a relatively low sensitivity with excellent specificity for PBC. A negative result does not rule out PBC.

Interpretation Component Anti-sp100 20.1 Units or less Negative 20.1-Antibody, IgG 24.9 Units Equivocal 25.0 Units or greater Positive

Reference Interval:



Test Number	Components	Reference Interval
	Anti-sp100 Antibody, IgG	24.9 Units or less



TEST CHANGE

Anti-sp100 and 3002482, SP100	anti-gp210 Ant GP210	tibodies, IgG	
Specimen Requir	ements:		
Patient Prepar	ration:		
Collect:		Serum <u>separator tube</u> Separator Tube (SST).	
Specimen Preparation:		Separate from cells ASAP or within 2 hours of collection. Transfer 1.0 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.3 mL)	
Transport Tem	nperature:	Refrigerated.	
Unacceptable Conditions:		NonserumNon-serum, heat-inactivated, contaminated, grossly icteric, severely lipemic, grossly hemolyzed specimens, or inclusion of fibrin clot.	
Remarks:			
Stability:		After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 month (avoid repeated freeze/thaw cycles)	
Methodology:		Semi-Quantitative Enzyme-Linked Immunosorbent Assay	
Performed:		<u>Thu</u> Wed	
Reported:		1-8 days	
Note:			
CPT Codes:		83516 x2	
New York DOH Approval Status:		This test is New York DOH approved.	
Interpretive Data:	:		
Refer to report			
Component Int	terpretation		
Anti-sp100 20 Antibody, IgG Ne 24 Eq Un Po	0.0 Units or less egative 20.1- 1.9 Units quivocal 25.0 nits or greater ositive		
Anti-gp210 20 Antibody, IgG Ne 24 Eq Un	0.0 Units or less egative 20.1- 1.9 Units quivocal 25.0 hits or greater		



Positive

Reference Interval:

Test Number	Components	Reference Interval
	Anti-gp210 Antibody, IgG	24.9 Units or less
	Anti-sp100 Antibody, IgG	24.9 Units or less



Phosphatidylethanol (PEth), Whole Blood, Quantitative

3002598, PETH	
Specimen Requirements:	
Patient Preparation:	
Collect:	Lavender (K2 or K3EDTA), pink (K2EDTA), dark green (lithium heparin), or gray (potassium oxalate).
Specimen Preparation:	Transport 1 mL whole blood. (Min: 0.5 mL)
Transport Temperature:	Refrigerated. Also acceptable: Frozen.
Unacceptable Conditions:	Gel separator tubes, plain red, light blue (citrate), or yellow (SPS or ACD solution).
Remarks:	
Stability:	Ambient: $\underline{32}$ hours; Refrigerated: 2 weeks; Frozen: 1 month (-20 Degrees C)
Methodology:	Quantitative Liquid Chromatography-Tandem Mass Spectrometry
Performed:	Sun-Sat
Reported:	1-4 days
Note:	
CPT Codes:	80321 (Alt code: G0480)

New York DOH Approval Status: This test is New York DOH approved.

Interpretive Data:

Phosphatidylethanol (PEth) is a group of phospholipids formed in the presence of ethanol, phospholipase D, and phosphatidylcholine. PEth is known to be a direct alcohol biomarker. The predominant PEth homologues are PEth 16:0/18:1 (POPEth) and PEth 16:0/18:2 (PLPEth), which account for 37-46% and 26-28% of the total PEth homologues, respectively. PEth is incorporated into the phospholipid membrane of red blood cells and has a general half-life of 4-10 days and a window of detection of 2-4 weeks. However, the window of detection is longer in individuals who chronically or excessively consume alcohol. Serial monitoring of PEth may be helpful in monitoring alcohol abstinence over time. PEth results should be interpreted in the context of the patient's clinical and behavioral history. Patients with advanced liver disease may have falsely elevated PEth concentrations (Nguyen VL, et al, Alcoholism: Clinical and Experimental Research, 2018).

Reference Interval:

Effective September 8, 2020



By Report



Non-Invasive Prenatal Aneuploidy Screen by cell-free DNA Sequencing

3003043, NIPT NGSAN		
Specimen Requirements:		
Patient Preparation:	Specimen must be collected at 10 weeks gestation or greater. Testing will be canceled for specimens collected at less than 10 weeks of gestation. Number of fetuses must be provided. Testing will be canceled if number of fetuses is not provided.	
Collect:	Black-and-tan top <u>or tan top</u> cell-free DNA BCT (Streck) tube (ARUP Supply #56435).) Available online through eSupply using ARUP Connect(TM) or contact ARUP Client Services at 800-522-2787.	
Specimen Preparation:	Transport 10 mL maternal whole blood (Min: 7 mL)	
Transport Temperature:	Refrigerated	
Unacceptable Conditions:	Ambient and frozen specimens.	
Remarks:	Patient History and Consent forms for the <u>INNon-invasive</u> prenatal aneuploidy screening test (NIPT) are available on the ARUP Web site or by contacting Client Services at 800-522- 2787.	
Stability:	Ambient: Unacceptable; Refrigerated: 10 days; Frozen: Unacceptable.	
Methodology:	Massively Parallel Sequencing	
Performed:	Varies	
Reported:	5-7 days	
Note:	Results will not be reported without a gestational age greater than or equal to 10 weeks. Testing will not be performed without number of fetuses provided. ARUP only performs testing on singleton pregnancies. Multiple gestation samples will be sent to Sequenom Laboratories to perform the MaterniT21 PLUS Core (chr21,18,13) test.	
CPT Codes:	81420	
New York DOH Approval Status:	This test is New York DOH approved.	
Interpretive Data:		



Refer to report.

Reference Interval:

N/A



Inflammatory Bowel Disease Differentiation Panel

3003748, IBD-PAN	
Specimen Requirements:	
Patient Preparation:	N/A
Collect:	Serum <u>separator tube</u> Separator Tube (SST).
Specimen Preparation:	Separate from cells ASAP or within 2 hours of collection. Transfer 1.5 mL serum to an ARUP standard transport tube. (Min: 0.6 mL)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Contaminated, heat-inactivated, hemolyzed, or severely lipemic specimens.
Remarks:	N/A
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 30 days (avoid repeated freeze/thaw cycles)
Methodology:	Semi-Quantitative Indirect Fluorescent Antibody (IFA)/Semi- Quantitative Enzyme Immunoassay (EIA)
Performed:	<u>Mon, Wed, Fri</u> Sun-Sat
Reported:	1-4 days
Note:	This test may be a useful tool for distinguishing ulcerative colitis (UC) from Crohn disease (CD) in patients with suspected inflammatory bowel disease. ANCA IFA is simultaneously tested on ethanol- and formalin-fixed slides to allow differentiation of C- and P-ANCA patterns.
CPT Codes:	86036; 86671 x2
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	
Refer to report.	
Reference Interval:	



Test Number	Components	Reference Inte	rval
	S. cerevisiae Antibody, IgG		
		20.0 Units or less	Negative
		20.1-24.9 Units	Equivocal
		25.0 Units or greater	Positive
	S. cerevisiae Antibody, IgA		
		20.0 Units or less	Negative
		20.1-24.9 Units	Equivocal
		25.0 Units or greater	Positive
	ANCA IFA Titer	Less than 1:20	
	ANCA IFA Pattern	None Detected	



Allergen, Food, Alpha-Gal (galad 3003924, ALPHAGALPN	ctose-alpha-1,3-galatose) Panel	
Specimen Requirements:		
Patient Preparation:	Multiple patient encounters should be avoided	
Collect:	Plain <u>red serum</u> Serum separator tube <u>(SST).</u> - Multiple specimen tubes should be avoided.	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer <u>1.0.25</u> mL serum plus 0.1 mL for each additional allergen ordered to an ARUP <u>standard transport tube</u> . Standard <u>Transport Tube</u> . (Min: 0. <u>425</u> mL plus 0.04 mL for each <u>additional</u> allergen ordered). For multiple allergen orders refer to "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Hemolyzed, icteric, or lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun-Sat	
Reported:	1-3 days	
Note:		
CPT Codes:	86008; 86003 x3	
New York DOH Approval Status:	This test is New York DOH approved.	
Interpretive Data:		
Allergen results of 0.10-0.34 kU/L a undetermined. Even though increase	are intended for specialist use as the clinical relevance is sing ranges are reflective of increasing concentrations of	

Allergen results of 0.10-0.34 kU/L are intended for specialist use as the clinical relevance is undetermined. Even though increasing ranges are reflective of increasing concentrations of allergen-specific IgE, these concentrations may not correlate with the degree of clinical response or skin testing results when challenged with a specific allergen. The correlation of allergy laboratory results with clinical history and in vivo reactivity to specific allergens is essential. A negative test may not rule out clinical allergy or even anaphylaxis.

Reference Interval:



Reporting Range (reported in kU/L)	Probability of IgE Mediated Clinical Reaction	Class Scoring
Less than 0.10	No significant level detected	0
0.10-0.34	Clinical relevance undetermined	0/1
0.35-0.70	Low	1
0.71-3.50	Moderate	2
3.51-17.50	High	3
17.51-50.00	Very high	4
50.01-100.00	Very high	5
Greater than 100.00	Very high	6



3003992, CARP IGG			
Specimen Requirements:			
Patient Preparation:			
Collect:	Serum separator tube.		
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum to an ARUP <u>standard transport</u> <u>tube.Standard Transport Tube.</u> (Min: 0.3 mL)		
Transport Temperature:	Refrigerated.		
Unacceptable Conditions:	Contaminated, heat-inactivated, hemolyzed, icteric, or lipemic specimens.		
Remarks:			
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 month (avoid repeated freeze/thaw cycles)		
Methodology:	Semi-Quantitative Enzyme-Linked Immunosorbent Assay		
Performed:	<u>Mon, Wed, Fri</u> Sun, Tue, Thu		
Reported:	1-4 days		
Note:			
CPT Codes:	83516		
New York DOH Approval Status:	This test is New York DOH approved.		

Interpretive Data:

Anti-carbamylated protein (anti-CarP) IgG antibodies are present in about 34-53 percent of patients with RA, have specificities of greater than 90 percent and can occur in RA patients seronegative for both rheumatoid factor and anti-CCP. These autoantibodies may be present in the preclinical phase of disease, are associated with future RA development, and may predict radiographic joint destruction. Patients with weak positive results should be monitored and testing repeated.

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

0-19 Units





Cytochrome P450 Genotyping Panel, with GeneDose Access			
3004255, CYP GD			
Specimen Requirements:			
Patient Preparation:			
Collect:	Lavender (K2EDTA), pink (K2EDTA), or yellow (ACD solution A or B).		
Specimen Preparation:	Transport 3 mL whole blood. (Min: 1 mL)		
Transport Temperature:	Refrigerated.		
Unacceptable Conditions:	Plasma or serum. Specimens collected in sodium heparin or lithium heparin. Frozen specimens in glass collection tubes.		
Remarks:			
Stability:	Ambient: 72 hours; Refrigerated: 1 week; Frozen: 1 month		
Methodology:	Polymerase Chain Reaction/Fluorescence Monitoring/Sequencing		
Performed:	Varies		
Reported:	5-10 days		
Note:	If long-range PCR/duplication testing is performed, additional charges will apply. Approximately less than 5% of samples require 2D6 copy number determination.		
CPT Codes:	81225; 81226; 81227; 81230; 81231; 81479; if reflexed, add 81479		
New York DOH Approval Status:	This test is New York DOH approved.		
Interpretive Data:			
Refer to report.			
Couseling and informed consent are recommended for genetic testing. Consent forms are available online.			
Reference Interval:			
By report			

HOTLINE NOTE: There is a component change associated with this test. One or more components



have been added or removed. Refer to the Hotline Test Mix for interface build information.



Cytogenomic Molecular Inversion Probe Array FFPE Tissue - Products of Conception

3004273, CMAPFFPE		
Specimen Requirements:		
Patient Preparation:		
Collect:	Fetal autopsy or products of conception.	
Specimen Preparation:	FFPE Fetal tissue: Transport ten slides, each with 5 microm unstained sections or four 20 microm scrolls or tissue block. OR FFPE villi: Transport one H&E stained slide and ten slides, each with 5 microm unstained sections or tissue block. New York State Clients: Transport 1 FFPE block or 10 slides (10 microm each).	
Transport Temperature:	Room temperature. Also acceptable: Refrigerated. Ship in cooled container during summer months.	
Unacceptable Conditions:	Specimens fixed or processed in alternative fixatives or heavy metal fixatives (B-4 or B-5).	
Remarks:	If multiple specimens (blocks or slides) are sent to ARUP, they must be accompanied by one of the following: an order comment indicating that the ARUP pathologist should choose the specimen most appropriate for testing (e.g., "Choose best block"), or individual orders for each sample submitted. A Pathologist Block Selection Fee (ARUP test code 3002076) will be added to orders that utilize the first option. If multiple specimens are sent to ARUP without a request for pathologist block/slide selection or individual orders, they will be held until clarification is provided.	
Stability:	Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable	
Methodology:	Molecular Inversion Probe Array	
Performed:	Sun-Sat	
Reported:	14-21 days	
Note:	If sending placenta instead of fetal tissue, at least 80% villi for products of conception specimens. This test must be ordered using Cytogenetic test request form #43098 or through your ARUP interface. Please submit the Patient History for Prenatal Cytogenetics form with the electronic packing list	



(http://ltd.aruplab.com/Tests/Pdf/65).

New York DOH Approval Status: Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	CPT Codes:	81229
	New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.

Interpretive Data:

For detection of copy number alterations and loss of heterozygosity in FFPE specimens. Refer to report.

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Counseling and informed consent are recommended for genetic testing. Consent forms are available online.

Reference Interval:

HOTLINE NOTE: There is a component change associated with this test. One or more components have been added or removed. Refer to the Hotline Test Mix for interface build information.



Cytogenomic Molecular Inversion Probe Array FFPE Tissue - Oncology

3004275, FFPEARRAY			
Specimen Requirements:			
Patient Preparation:			
Collect:	Tumor tissue		
Specimen Preparation:	Formalin fix (10 percent neutral buffered formalin) and paraffin embed tissue. Protect from excessive heat. Transport 10 slides, each with 5-micron unstained sections or four 20- micron scrolls or tissue block. Tissue block will be returned after testing. Transport tissue in a tissue transport kit (ARUP supply #47808) available online through eSupply using ARUP Connect(TM) or contact ARUP Client Services at (800) 522- 2787.		
Transport Temperature:	Room temperature. Also acceptable: Refrigerated. Ship in cooled container during summer months.		
Unacceptable Conditions:	Specimens fixed or processed in alternative fixatives or heavy metal fixatives (B-4 or B-5).		
Remarks:	If multiple specimens (blocks or slides) are sent to ARUP, they must be accompanied by one of the following: an order comment indicating that the ARUP pathologist should choose the specimen most appropriate for testing (e.g., "Choose best block"), or individual orders for each sample submitted. A Pathologist Block Selection Fee (ARUP test code 3002076) will be added to orders that utilize the first option. If multiple specimens are sent to ARUP without a request for pathologist block/slide selection or individual orders, they will be held until clarification is provided.		
Stability:	Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable		
Methodology:	Molecular Inversion Probe Array		
Performed:	Sun-Sat		
Reported:	14-21 days		
Note:	Samples must contain a region with at least 50 percent tumor.		
CPT Codes:	81277		



New York DOH Approval Status:

Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.

Interpretive Data:

For detection of copy number alterations and loss of heterozygosity in FFPE specimens. Refer to report.

This test was developed and its performance characteristics determined by ARUP Laboratories. It has not been cleared or approved by the US Food and Drug Administration. This test was performed in a CLIA certified laboratory and is intended for clinical purposes.

Reference Interval:

HOTLINE NOTE: There is a component change associated with this test. One or more components have been added or removed. Refer to the Hotline Test Mix for interface build information.



Pharmacogenetics Panel: Psychotropics				
3004471, PGX PSYCH	3004471, PGX PSYCH			
Specimen Requirements:				
Patient Preparation:				
Collect:	Whole Blood: Lavender (EDTA), pink (K2EDTA), or yellow (ACD solution A or B).			
Specimen Preparation:	Transport 3 mL whole blood. (Min: 1 mL)			
Transport Temperature:	Refrigerated.			
Unacceptable Conditions:	Plasma or serum. Specimens collected in sodium heparin or lithium heparin. Frozen specimens in glass collection tubes.			
Remarks:				
Stability:	Ambient: 72 hours; Refrigerated: 1 week; Frozen: 1 month			
Methodology:	Polymerase Chain Reaction (PCR)/Fluorescence Monitoring/Sequencing			
Performed:	Varies			
Reported:	5-10 days			
Note:	If long-range PCR/duplication testing is performed, additional charges apply. Approximately less than 5% of samples require 2D6 copy number determination.			
CPT Codes:	81225; 81226; 81227; 81230; 81231; 81291; 81479; if reflexed, add 81479			
New York DOH Approval Status:	This test is New York DOH approved.			
Interpretive Data:				
Refer to report				
Counseling and informed consent are recommended for genetic testing. Consent forms are available online.				
Reference Interval:				
By report				

HOTLINE NOTE: There is a component change associated with this test. One or more components



have been added or removed. Refer to the Hotline Test Mix for interface build information.



Drug Profile, Expanded Targeted Panel by LC-MS/MS, Serum/Plasma 3004833, COMPDRUGSP			
Specimen Requirements:			
Patient Preparation:			
Collect:	Plain red (no additives, serum gel or SST tubes are not acceptable), gray top (NaF/oxalate), lavender (K2 or K3EDTA) or pink (K2EDTA).		
Specimen Preparation:	Separate serum or plasma from cells ASAP or within 2 hours of collection. Transfer 4 mL of serum or plasma into an ARUP standard transport tube. (Min: 2 mL)		
Transport Temperature:	Refrigerated.		
Unacceptable Conditions:	Gel separator tubes or light blue (sodium citrate). Specimens exposed to repeated freeze/thaw cycles. Postmortem specimens.		
Remarks:			
Stability:	After separation from cells: Ambient: 24 hours; Refrigerated: 2 weeks; Frozen: 2 months.		
Methodology:	Qualitative Liquid Chromatography-Tandem Mass Spectrometry		
Performed:	Mon, Fri		
Reported:	1- <mark>98</mark> days		
Note:	If a drug class is reported as present, the test will report the specific drug present from the drug class. (Classes include ARUP test codes 3005045, 3005046, 3005047, 3005048, 3005049, 3005050, 3005051, 3005052, 3005053, 3005054, 3005055, 3005056, 3005057, 3005058, or 3005059.)		
CPT Codes:	80323; 80326; 80329; 80334; 80337; 80338; 80341; 80344; 80346; 80348; 80353; 80354; 80356; 80357; 80358; 80359; 80360; 80361; 80363; 80365; 80366; 80368; 80370; 80371; 80372; 80373; 80377; 83992 (Alt code: G0483)		
New York DOH Approval Status:	This test is New York DOH approved.		
Interpretive Data:			



The qualitative drug panel can detect 127 drugs and drug metabolites by LC-MS/MS. The absence of expected drug(s) and/or drug metabolite(s) may indicate noncompliance, inappropriate timing of specimen collection relative to drug administration, poor drug absorption, or limitations of testing. The concentration at which the test can detect a drug or metabolite varies within a drug class. The concentration value must be greater than or equal to the cutoff to be reported as positive. Interpretive questions should be directed to the laboratory.

This assay is for medical purposes only; not valid for forensic use.

For a complete list of drugs and drug metabolites detected, refer to the Additional Technical Information.

Reference Interval:

By report



Drug Profile, Expanded Targeted Panel by LC-MS/MS, Urine

3005060, COMPDRUGUR		
Specimen Requirements:		
Patient Preparation:		
Collect:	Random urine with no additives.	
Specimen Preparation:	Transport 4 mL urine. (Min: 2 mL)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Specimens exposed to repeated freeze/thaw cycles.	
Remarks:		
Stability:	Ambient: 1 day; Refrigerated: 2 weeks; Frozen: 2 months	
Methodology:	Qualitative Liquid Chromatography-Tandem Mass Spectrometry	
Performed:	Mon, Fri	
Reported:	1- <u>9</u> 8 days	
Note:	If a drug class is reported as present, the test will report the specific drug present from the drug class at no additional charge. (Classes include ARUP test codes 3005225, 3005226, 3005227, 3005228, 3005229, 3005230, 3005231, 3005232, 3005233, 3005234, 3005235, 3005236, 3005237, 3005238, or 3005239.)	
CPT Codes:	80323; 80325; 80329; 80334; 80337; 80338; 80341; 80344; 80347; 80348; 80353; 80354; 80355; 80356; 80357; 80359; 80360; 80361; 80363; 80365; 80366; 80368; 80370; 80371; 80372; 80373; 80377; 83992 (Alt code: G0483)	
New York DOH Approval Status:	This test is New York DOH approved.	

Interpretive Data:

The qualitative drug panel can detect 127 drugs and drug metabolites by LC-MS/MS. The absence of expected drug(s) and/or drug metabolite(s) may indicate noncompliance, inappropriate timing of specimen collection relative to drug administration, poor drug absorption, diluted/adulterated urine, or limitations of testing. The concentration at which the test can detect a drug or metabolite varies within a drug class. The concentration value must be greater than or equal to the cutoff to be reported as positive. Interpretive questions should be directed to the laboratory.



This assay is for medical purposes only; not valid for forensic use.

For a complete list of drugs and drug metabolites detected, refer to Additional Technical Information.

Reference Interval:

By Report



TEST CHANGE

Toxocara Antibodies, IgG by ELISA 3006066, TOXOCARA G			
Specimen Requirements:			
Patient Preparation:			
Collect:	Serum separator tube (SST) or plain red.		
Specimen Preparation:	Transfer 1 mL serum to an ARUP standard transport tube. (Min: 0.5 mL)		
Transport Temperature:	Preferred transport temp: Refrigerated. Also acceptable: Frozen		
Unacceptable Conditions:	Contaminated, heat-inactivated, grossly hemolyzed, or lipemic specimens.		
Remarks:			
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 month		
Methodology:	Semi-QuantitativeSemiquantitativeEnzyme-Linked ImmunosorbentAssay <u>(ELISA)</u>		
Performed:	ThuWed, Sat		
Reported:	1- <u>8</u> 7 days		
Note:	N/A		
CPT Codes:	86682		
New York DOH Approval Status:	This test is New York DOH approved.		
Interpretive Data:			
Component Interpretation			
Toxocara< 9 U Negative:Antibodies, IgG byNo significantELISAlevel of ToxocaraIgG antibodiesdetected. 911U Equivocal:Recommendrepeat testing in2-4 weeks. >11 UPositive: IgGantibodies toToxocaradetected,indicating current			



are possible.

Reference Interval:

Test Number	Components	Reference Interval
	Toxocara Antibodies, IgG by ELISA	8 U or less



Pharmacogenetics Panel: Psychotropics, with GeneDose Access	
Specimen Requirements:	
Patient Prenaration:	
Collect:	Lavender (K2EDTA), pink (K2EDTA), or yellow (ACD solution A or B).
Specimen Preparation:	Transport 3 mL whole blood. (Min: 1 mL)
Transport Temperature:	Refrigerated
Unacceptable Conditions:	Plasma or serum. Specimens collected in sodium heparin or lithium heparin. Frozen specimens in glass collection tubes.
Remarks:	
Stability:	Ambient: 72 hours; Refrigerated: 1 week; Frozen: 1 month
Methodology:	Polymerase Chain Reaction (PCR)/Fluorescence Monitoring/Sequencing
Performed:	Varies
Reported:	5-10 days
Note:	If long-range PCR/duplication testing is performed, additional charges apply. Approximately less than 5% of samples require 2D6 copy number determination.
CPT Codes:	81225; 81226; 81227; 81230; 81231; 81291; 81479; if reflexed, add 81479
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	
Refer to report.	
Counseling and informed consent are recommended for genetic testing. Consent forms are available online.	
Reference Interval:	

HOTLINE NOTE: There is a component change associated with this test. One or more components have been added or removed. Refer to the Hotline Test Mix for interface build information.





HPV Primary Screen by PCR With Reflex to Cytology

Specimen Requirements:		
Patient Preparation:		
Collect:	Cervical or endocervical specimen with brush or spatula from ThinPrep kit collection kit. (ARUP supply #41785 ThinPrep (Vial and Broom) or #51369 ThinPrep (Vial, Brush, and Spatula)) available online through eSupply using ARUP <u>Connect(TM) or Connector</u> contact ARUP Client Services at 800-522-2787.	
Specimen Preparation:	Place collection device in corresponding ThinPrep media vial.	
Transport Temperature:	Refrigerated	
Unacceptable Conditions:	Bloody or dark brown specimens. Specimens in any media other than indicated above.	
Remarks:		
Stability:	Ambient: 6 months; Refrigerated: 6 months; Frozen: Unacceptable	
Methodology:	Qualitative Polymerase Chain Reaction (PCR)	
Performed:	Tue-Sat	
Reported:	<u>1-5 days</u> Within 24 hours	
Note:	If HPV assay is positive, then ThinPrep PAP Test (Standalone) (3018968) will be added. Additional charges apply. For cervical sources, a negative high-risk HPV result does not exclude the possibility of future cytologic abnormalities, underlying CIN2-3, or cancer.	
CPT Codes:	8762 <u>4</u> 6; if reflexed, add <u>88175 (</u> 88142 <u>if manual);</u> ; if reviewed by pathologist, add 88141	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
This test amplifies DNA of HPV16, HPV18, and 12 other high-risk HPV types (31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, and 68) associated with cervical cancer and its precursor lesions. Sensitivity may		



be affected by specimen collection methods, stage of infection, and the presence of interfering substances. <u>Specimens positive for high-risk HPV types are reflexed to cytology</u>. <u>Patients positive</u> for high-risk HPV 16 or HPV 18 should be managed according to current ASCCP guidelines (2019). <u>Results should be interpreted in conjunction with other available laboratory and clinical data</u>. A <u>negative high-risk HPV result does not exclude the presence of other high-risk HPV types, the</u> <u>possibility of future cytologic abnormalities, underlying CIN2-3, or cancer</u>.

HPV testing should not be used for screening or management of atypical squamous cells of undetermined significance (ASCUS) in women under age 21.

Reference Interval:

Negative


Anti-Phospholipase A2 Receptor (PLA2R) Antibody, IgG by ELISA		
3016767, ANTI-PLA2R		
Specimen Requirements:		
Patient Preparation:	Separate serum from cells ASAP or within 2 hours of collection.	
Collect:	Serum separator tube.	
Specimen Preparation:	Transfer 1 mL serum to an ARUP standard transport tube. (Min: 0.5 mL)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Contaminated, heat-inactivated, grossly hemolyzed, grossly icteric, or grossly lipemic specimens.	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 2 weeks	
Methodology:	Semi-Quantitative Enzyme-Linked Immunosorbent Assay (ELISA)	
Performed:	<u>Wed, FriMon, Thu</u>	
Reported:	1-7 days	
Note:		
CPT Codes:	83516	
New York DOH Approval Status:	This test is New York DOH approved.	

Interpretive Data:

A positive anti-phospholipase A2 receptor (PLA2R) antibody result by ELISA or IFA in conjunction with clinical symptoms and other laboratory findings is suggestive of primary membranous nephropathy (pMN). Absence of circulating anti-PLA2R receptor autoantibodies does not rule out a diagnosis of pMN. Anti-PLA2R antibody titers, due to its high predictive value, can be useful for assessing disease severity and monitoring clinical remission. In patients with pMN undergoing treatment, low antibody titers are associated with disease remission and high titers indicate loss of kidney function and need for an aggressive therapeutic approach.

Component	Interpretive Data
Anti-	Negative: Less
Phospholipase A2	than 14 RU/mL
Receptor, IgG	Borderline: 14-19
	RU/mL Positive:
	Greater than or



equal to 20 RU/mL

Test Number	Components	Reference Interval
	Anti-Phospholipase A2 Receptor, IgG	< 14 RU/mL



Thrombotic Risk Reflex Panel	
3017156, THROMRISK	
Specimen Requirements:	
Patient Preparation:	Fasting preferred. Refer to Specimen Handling at aruplab.com for hemostasis/thrombosis specimen handling guidelines.
Collect:	Four light blue (sodium citrate) AND two lavender (EDTA) AND two serum separator tubes (SSTs). Also acceptable in place of one of the serum separator tubes (SSTs): green (sodium or lithium heparin) or EDTA (K2 or K3).
Specimen Preparation:	One serum separator tube (SST), green (sodium or lithium heparin),) or EDTA (K2 or K3) must be centrifuged and serum or plasma separated within 1 hour of collection. Transfer 1 mL centrifuged serum or plasma to ARUP standard transport tube and label centrifuged tube for homocysteine testing. (Min: 0.5 mL) AND Transfer 2 mL serum into 2 ARUP standard transport tubes, label as serum. (Min: 0.5 mL/tube) AND Transfer 7.5 mL platelet poor plasma prepared from the sodium citrate tubes to 5 ARUP standard transport tubes, label as sodium citrate. (Min: 1 mL/tube) AND Transfer 3 mL lavender whole blood to 2 ARUP standard transport tubes. (Min: 1 mL/tube)
Transport Temperature:	Light blue (sodium citrate): CRITICAL FROZEN. Separate specimens must be submitted when multiple tests are ordered. Lavender whole blood and serum, green (sodium or lithium heparin).) or EDTA (K2 or K3): Frozen.
Unacceptable Conditions:	Specimens collected in any tube type not listed above.
Remarks:	
Stability:	Light blue (sodium citrate): Ambient: Unacceptable; Refrigerated: Unacceptable; Frozen: 2 weeks Lavender whole blood: Ambient: 7 days; Refrigerated: 1 week; Frozen: 1 month Serum: Ambient: 2 hours; Refrigerated: 1 week; Frozen: 2 weeks Green (sodium or lithium heparin) or EDTA (K2 or K3): Ambient: 4 days; Refrigerated: 1 month; Frozen: 10 months
Methodology:	Electromagnetic Mechanical Clot Detection/Chromogenic Assay/Quantitative Enzymatic Assay/Semi-Quantitative Enzyme-Linked Immunosorbent Assay (ELISA)/Polymerase Chain Reaction (PCR)/Fluorescence Monitoring/Microlatex Particle-Mediated Immunoassay
Performed:	Varies



Reported: 2-7 days Note: Testing will include Antithrombin, Enzymatic (Activity) (0030010); Protein S Free, Antigen (0098894); Protein C, Functional (0030113); Beta-2 Glycoprotein 1 Antibodies, IgG and IgM (0050321); Cardiolipin Antibodies, IgG and IgM (0099344); Lupus Anticoagulant Reflex Panel (3017009); Prothrombin (F2) c.*97G>A (G20210A) Pathogenic Variant (0056060); APC Resistance Profile with Reflex to Factor V Leiden (0030192); and Homocysteine, Total (0099869). If APC resistance is low, or if a valid result cannot be obtained for the APC portion of the profile, then Factor V Leiden by PCR will be added. Additional charges apply. For the Lupus Anticoagulant Reflex Panel (3017009) portion of the panel, if PTT-LA Ratio and dRVVT Screen Ratio are normal, then no further clot-based testing is performed. If either the PTT-LA Ratio or dRVVT Screen Ratio are elevated, then Anti-Xa Qualitative Interpretation is added. If PTT-LA Ratio is elevated, then Thrombin Time is also added. If Anti-Xa Qualitative Interpretation is Present and Thrombin Time is greater than 30 seconds, then Hepzyme treatment is added. If PTT-LA Ratio is normal and Anti-Xa Qualitative Interpretation is Present, or Thrombin Time is greater than 30 seconds, and Anti-Xa Qualitative Interpretation is Not Present, or Thrombin Time is less than 30 seconds, and Anti-Xa Qualitative Interpretation is Present, then DOAC-Stop treatment is added. If either Hepzyme or DOAC-Stop treatment is added, then Neutralized PTT-LA Ratio and/or Neutralized dRVVT Screen Ratio are added. If dRVVT Screen Ratio is elevated in the absence of Hepzyme or DOAC-Stop, or if Neutralized dRVVT Screen Ratio is elevated, then dRVVT 1:1 Mix Ratio and dRVVT Confirmation Ratio are added. If PTT-LA Ratio is elevated in the absence of Hepzyme or DOAC-Stop treatment, or if Neutralized PTT-LA Ratio is elevated, then Hexagonal Phospholipid Confirmation is added. Additional charges apply. False elevations of plasma or serum homocysteine may occur if the plasma or serum is not promptly separated from the cells at the time of collection. CPT Codes: 81240; 83090; 85300; 85303; 85306; 85307; 85610; 85613; 85730; 86147x2; 86146x2; if reflexed, additional CPT codes may apply: 81241; 85520; 85525; 85598; 85613; 85670; 85730. New York DOH Approval Status: This test is New York DOH approved. Interpretive Data: Refer to individual components.



Effective Date: July 21, 2025

Reference Interval:

Refer to individual components.



Herpes Simplex Virus Type 1 and/or 2 Antibodies, IgG (CSF) With Reflex to Type 1 and 2 Glycoprotein G-Specific Ab, IgG

3017747, HERPR CSF

Specimen Requirements:

Patient Preparation:	
Collect:	CSF
Specimen Preparation:	Transfer 1 mL CSF to an ARUP standard transport tube. (Min: 0.5 mL)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Specimen types other than CSF. Contaminated, heat- inactivated <u>, icteric</u> , or hemolyzed specimens.
Remarks:	
Stability:	Ambient: 8 hours; Refrigerated: 2 weeks; Frozen: 1 year
Methodology:	<u>Semi-Quantitative Chemiluminescent Immunoassay (CLIA) /</u> <u>Semi-Quantitative Enzyme-Linked Immunosorbent Assay</u> (ELISA)
Performed:	Sun-Sat
Reported:	1-2 days
Note:	If HSV 1/2 IgG, CSF is greater than or equal to 1.10 IV , then HSV 1 gG-specific IgG, CSF and HSV 2 gG-specific IgG, CSF will be added. Additional charges apply.
CPT Codes:	86694; if reflexed, add 86695; 86696
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	

False positive results are possible. Consider additional testing for HSV-2, particularly if the result for HSV-2 is less than or equal to </= 3.0 IV.

Herpes Simplex0.89 IV or less:Virus Type 1Negative. Noand/or 2significant levelAntibodies, IgG,of detectable HSVCSFIgG antibody.0.90-1.09 IV:Equivocal.



Questionable presence of IgG antibodies. Repeat testing in 10-14 days may be helpful. 1.10 IV or greater: Positive. IgG antibody to HSV detected which	
Positive. IgG antibody to HSV detected which	
may indicate a current or past HSV infection.	

Reference Interval:

0.89 IV or less



SC5b-9	
3017902, SC5B-9	
Specimen Requirements:	
Patient Preparation:	
Collect:	Pink (K2EDTA), <u>t</u> an (K2EDTA), <u>r</u> Royal blue (K2EDTA), or <u>l</u> Lavender (EDTA).
Specimen Preparation:	Separate plasma within 2 hours (1 hour is preferable) by centrifugation <u>.~2700 rpm (1300 100 g) for 10 minutes</u> . Transfer plasma (minimum 0.5 mL) to an ARUP standard transport tube and freeze immediately.
Transport Temperature:	CRITICAL FROZEN. Separate specimens must be submitted when multiple tests are ordered.
Unacceptable Conditions:	Nonfrozen specimens. Specimens exposed to repeated freeze/thaw cycles. Grossly hemolyzed, lipemic, and icteric specimens. Serum samples. Heparinized and lithium samples.
Remarks:	
Stability:	Ambient: Unacceptable; Refrigerated: Unacceptable; Frozen: 30 days
Methodology:	Quantitative Enzyme-Linked Immunosorbent Assay (ELISA)
Performed:	Sun, Wed
Reported:	2-12 days
Note:	
CPT Codes:	86160

New York DOH Approval Status: This test is New York DOH approved.

Interpretive Data:

Elevated soluble C5b-9 (SC5b-9) levels indicate recent or ongoing activation of the complement system, while normal or reduced levels suggest no excessive activation. High SC5b-9 concentrations are associated with transplant-associated thrombotic microangiopathy (TA-TMA), a complication of hematopoietic stem cell transplants. Increased SC5b-9 may also occur in various conditions involving primary or secondary complement activation, such as immune-complex disease, infection, atypical hemolytic uremic syndrome, and C3 glomerulopathies. Due to a low specificity for SC5b-9 testing, results should be interpreted in combination with other clinical and laboratory evidence of disease activity. Plasma SC5b-9 levels may be used to monitor the efficacy of complement inhibitor drugs, as elevated levels suggest insufficient complement blockage to



effectively prevent the formation of the terminal attack complex.

Test Number	Components	Reference Interval
	C5b9 Soluble Terminal Complement Complex	Less than or equal to 260 ng/mL



Allergen, Food, Tree Nuts With Reflex to Components, IgE 3018799, TNUT PAN R		
Specimen Requirements:		
Patient Preparation:	Multiple patient encounters should be avoided.	
Collect:	Plain red or serum Serum separator tube (SST)-	
Specimen Preparation:	Separate serum from cells ASAP or within 2 hours of collection. Transfer <u>2.01.5</u> mL serum to an ARUP standard transport tube. (Min: 1.5 mL plus 0.04 mL for each additional allergen ordered mL). For multiple allergen orders refer to "Allergen Specimen Collection Instructions" at www.aruplab.com/testing/resources/specimen.	
Transport Temperature:	Refrigerated	
Unacceptable Conditions:	Postmortem samples	
Remarks:		
Stability:	After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 month	
Methodology:	Quantitative ImmunoCAP Fluorescent Enzyme Immunoassay	
Performed:	Sun-Sat	
Reported:	1-3 days	
Note:	This assay will initially test brazil nut, cashew, pistachio, hazelnut, walnut, pecan, almond. If the brazil nut whole allergen result is greater than or equal to 0.1 kU/L, the brazil nut component Ber e 1 will be ordered. If the cashew whole allergen result is greater than or equal to 0.1 kU/L, the cashew component Ana o 3 will be ordered. If the hazelnut whole allergen result is greater than or equal to 0.1 kU/L, the hazelnut components Cor a 1, Cor a 8, Cor a 9, Cor a 14 will be ordered. If the walnut whole allergen result is greater than or equal to 0.1 kU/L, the walnut components Jug r 1 and Jug r 3 will be ordered. Additional charges apply.	
CPT Codes:	86003 x7; if reflexed, add 86008 per component	
New York DOH Approval Status:	This test is New York DOH approved.	
Interpretive Data:		



Allergen results of 0.10-0.34 kU/L are intended for specialist use as the clinical relevance is undetermined. Even though increasing ranges are reflective of increasing concentrations of allergen-specific IgE, these concentrations may not correlate with the degree of clinical response or skin testing results when challenged with a specific allergen. The correlation of allergy laboratory results with clinical history and in vivo reactivity to specific allergens is essential. A negative test may not rule out clinical allergy or even anaphylaxis.

Reporting Range (reported in Ku/L)	Probability of IgE Mediated Clinical Reaction	Class Scoring
Less than 0.10	No significant level detected	0
0.10-0.34	Clinical relevance undetermined	0/1
0.35-0.70	Low	1
0.71-3.50	Moderate	2
3.51-17.50	High	3
17.51-50.00	Very high	4
50.01-100.00	Very high	5
Greater than 100.00	Very high	6

Test Number	Components	Reference Interval
	Allergen, Food, Almond IgE	Less than or equal to 0.34 kU/L
	Allergen, Food, Brazil Nut IgE	Less than or equal to 0.34 kU/L
	Allergen, Food, Cashew IgE	Less than or equal to 0.34 kU/L
	Allergen, Food, Pecan IgE	Less than or equal to 0.34 kU/L
	Allergen, Food, Hazelnut IgE	Less than or equal to 0.34 kU/L
	Allergen, Food, Pistachio IgE	Less than or equal to 0.34 kU/L
	Allergen, Food, Walnut (Juglans spp) IgE	Less than or equal to 0.34 kU/L



Dermatomyositis and Polymyositis Panel 3018866, COMBI PAN2			
Specimen Requirements:			
Patient Preparation:			
Collect:	Serum separator tube (SST).		
Specimen Preparation:	Separate from cells ASAP or within 2 hours of collection. Transfer $\frac{3 \text{ to}}{4}$ mL to ARUP standard transport tube. (Min: $\frac{31.5}{2}$ mL)		
Transport Temperature:	Refrigerated.		
Unacceptable Conditior	s: Hemolyzed, hyperlipemic, icteric, heat-treated, or contaminated specimens		
Remarks:			
Stability:	Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 month		
Methodology:	Qualitative Immunoprecipitation / Semi-Quantitative Multiplex Bead Assay / Qualitative Immunoblot / Semi-Quantitative Indirect Fluorescent Antibody (IFA) / Qualitative Particle-Based Multianalyte Technology (PMAT)		
Performed:	Sun-Sat		
Reported:	7-18 days		
Note:	Antibodies: PL-7, PL12, EJ, OJ, SRP, Jo-1, Mi-2, P155/140, SAE1, MDA5, NXP2, TIF1-gamma, ANA, Ha, Ks, Zo		
CPT Codes:	83516 x11; 84182 x3; 86235; 86039		
New York DOH Approval S	atus: This test is New York DOH approved.		
Interpretive Data:			
Refer to report.			
Component Interpretation Jo-1 Antibody, 29 AU/mL or la IgG Negative 30-4 AU/mL: Equive 41 AU/mL or greater: Positi	ess:) ical /e		



Test Number	Components	Reference Interval
	Zo (phenylalanyl-tRNA synthetase) Ab	Negative
	Ks (asparaginyl-tRNA synthetase) Ab	Negative
	Ha (tyrosyl-tRNA synthetase) Ab	Negative
	TIF-1 gamma (155 kDa) Ab	Negative
	NXP2 (Nuclear matrix protein-2) Ab	Negative
	MDA5 (CADM-140) Ab	Negative
	SAE1 (SUMO activating enzyme) Ab	Negative
	Antinuclear Antibody (ANA), HEp-2, IgG	Less than 1:80
	OJ (isoleucyl-tRNA synthetase) Antibody	Negative
	SRP (Signal Recognition Particle) Ab	Negative
	EJ (glycyl-tRNA synthetase) Antibody	Negative
	P155/140 Antibody	Negative
	PL-12 (alanyl-tRNA synthetase) Antibody	Negative
	PL-7 (threonyl-tRNA synthetase) Antibody	Negative
	Mi-2 (nuclear helicase protein) Antibody	Negative
	Jo-1 (Histidyl-tRNA Synthetase) Ab, IgG	40 AU/mL or less



Extended Myositis Panel			
3018867, MYOS EXT2			
Specimen Requirements:			
Patient Preparation:			
Collect:	Serum separator tube (SST), red top tube		
Specimen Preparation:	Separate from cells ASAP or within 2 hours of collection. Transfer $\frac{3 \text{ to } 4}{3 \text{ to } 4}$ mL to ARUP standard transport tube. (Min: $\frac{31.5}{mL}$ mL)		
Transport Temperature:	Refrigerated		
Unacceptable Conditions:	Hemolyzed, hyperlipemic, icteric, heat-treated, or contaminated specimens.		
Remarks:			
Stability:	Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 month		
Methodology:	Semi-Quantitative Enzyme-Linked Immunosorbent Assay (ELISA) / Qualitative Immunoprecipitation / Semi-Quantitative Multiplex Bead Assay / Qualitative Immunoblot / Semi- Quantitative Indirect Fluorescent Antibody (IFA) / Qualitative Particle-Based Multianalyte Technology (PMAT)		
Performed:	Sun-Sat		
Reported:	7-18 days		
Note:	Antibodies: Mi-2, PL-7, PL12, P155/140, EJ, Ku, OJ, PM/Scl, SRP, Smith/RNP, Ro52, Ro60, Jo-1, U3 Fib, SAE1, NXP2, MDA5, TIF1-gamma, ANA, Ha, Ks, Zo If HMGCR antibody, IgG is positive, additional testing to follow.		
CPT Codes:	83516 x13; 84182 x3; 86039; 86235 x6; if reflexed, add 83516		
New York DOH Approval Status:	This test is New York DOH approved.		
Interpretive Data:			



Refer to report.

Component	Interpretation
SSA-52 (Ro52) (ENA) Antibody, IgG	29 AU/mL or less: Negative 30-40 AU/mL: Equivocal 41 AU/mL or greater: Positive
SSA-60 (Ro60) (ENA) Antibody, IgG	29 AU/mL or less: Negative 30-40 AU/mL: Equivocal 41 AU/mL or greater: Positive
Smith/RNP (ENA) Antibody, IgG	19 Units or less: Negative 20-39 Units: Weak Positive 40-80 Units: Moderate Positive 81 Units or greater: Strong Positive
Jo-1 Antibody, IgG	29 AU/mL or less: Negative 30-40 AU/mL: Equivocal 41 AU/mL or greater: Positive



Test Number	Components	Reference Interval
	HMGCR Antibody Screen	Negative
	Zo (phenylalanyl-tRNA synthetase) Ab	Negative
	Ks (asparaginyl-tRNA synthetase) Ab	Negative
	Ha (tyrosyl-tRNA synthetase) Ab	Negative
	TIF-1 gamma (155 kDa) Ab	Negative
	NXP2 (Nuclear matrix protein-2) Ab	Negative
	MDA5 (CADM-140) Ab	Negative
	SAE1 (SUMO activating enzyme) Ab	Negative
	Antinuclear Antibody (ANA), HEp-2, IgG	Less than 1:80
	Fibrillarin (U3 RNP) Ab, IgG	Negative
	SSA-60 (Ro60) (ENA) Antibody, IgG	40 AU/mL or less
	OJ (isoleucyl-tRNA synthetase) Antibody	Negative
	SRP (Signal Recognition Particle) Ab	Negative
	Ku Antibody	Negative
	EJ (glycyl-tRNA synthetase) Antibody	Negative
	P155/140 Antibody	Negative
	PL-12 (alanyl-tRNA synthetase) Antibody	Negative
	PL-7 (threonyl-tRNA synthetase) Antibody	Negative
	Mi-2 (nuclear helicase protein) Antibody	Negative
	PM/Scl 100 Antibody, IgG	Negative
	Jo-1 (Histidyl-tRNA Synthetase) Ab, IgG	40 AU/mL or less
	SSA-52 (Ro52) (ENA) Antibody, IgG	40 AU/mL or less
	Smith/RNP (ENA) Ab, IgG	19 Units or less



Polymyositis Panel		
3018868, PO	LY MYO2	
Specimen Requirements:		
Patient Pre	eparation:	
Collect:		Serum separator tube (SST)
Specimen	Preparation:	Separate from cells ASAP or within 2 hours of collection. Transfer $4two - 1$ mL serum aliquots to an ARUP standard transport tubes. (Min: 30.5 mL/aliquot)
Transport [*]	Temperature:	Refrigerated.
Unaccepta	ble Conditions:	Hemolyzed, hyperlipemic, icteric, heat-treated, or contaminated specimens
Remarks:		
Stability:		Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 month
Methodology:		Qualitative Immunoprecipitation / Semi-Quantitative Multiplex Bead Assay / Semi-Quantitative Indirect Fluorescent Antibody (IFA) / Qualitative Immunoblot
Performed:		Mon, Tue, Thu, Fri
Reported:		7-18 days
Note:		Antibodies: PL-7, PL12, EJ, OJ, SRP, Jo-1, ANA, Ha, Ks, Zo
CPT Codes:		83516 x5; 86235; 84182 x3; 86039
New York DOH Approval Status:		This test is New York DOH approved.
Interpretive Data:		
Refer to report.		
Component Jo-1 Antibody, IgG	Interpretation 29 AU/mL or less: Negative 30-40 AU/mL: Equivocal 41 AU/mL or greater: Positive	



Test Number	Components	Reference Interval
	Zo (phenylalanyl-tRNA synthetase) Ab	Negative
	Ks (asparaginyl-tRNA synthetase) Ab	Negative
	Ha (tyrosyl-tRNA synthetase) Ab	Negative
	Antinuclear Antibody (ANA), HEp-2, IgG	Less than 1:80
	OJ (isoleucyl-tRNA synthetase) Antibody	Negative
	SRP (Signal Recognition Particle) Ab	Negative
	EJ (glycyl-tRNA synthetase) Antibody	Negative
	PL-12 (alanyl-tRNA synthetase) Antibody	Negative
	PL-7 (threonyl-tRNA synthetase) Antibody	Negative
	Jo-1 (Histidyl-tRNA Synthetase) Ab, IgG	40 AU/mL or less



Interstitial Lung Disease Autoantibody Panel
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3018869, ILD PANEL2				
Specimen Requirements:				
Patient Preparation:				
Collect:	Serum separator tube (SST).			
Specimen Preparation:	Separate from cells ASAP or within 2 hours of collection. Transfer $\frac{3 + 6 - 4.5}{5}$ mL to ARUP standard transport tube. (Min: $\frac{3}{1}$.5 mL)			
Transport Temperature:	Refrigerated.			
Unacceptable Conditions:	Hemolyzed, hyperlipemic, icteric, heat-treated, or contaminated specimens.			
Remarks:				
Stability:	Ambient: 24 hours; Refrigerated: 1 weeks; Frozen: 1 month			
Methodology:	Qualitative Immunoprecipitation / Semi-Quantitative Multiplex Bead Assay / Qualitative Immunoblot / Semi-Quantitative Enzyme-Linked Immunosorbent Assay (ELISA) / Quantitative Immunoturbidimetry / Semi-Quantitative Indirect Fluorescent Antibody (IFA) / Qualitative Particle-Based Multianalyte Technology (PMAT)			
Performed:	Sun-Sat			
Reported:	7-18 days			
Note:	Antibodies: Ro52, Ro60, Jo-1, PL-7, PL12, EJ, Ku, SRP, OJ, PM/Scl-100, MDA5, CCP, Scl-70, RA, ANA, NXP-2, RNA Polymerase III, Ha, Ks, Zo			
CPT Codes:	86235 x5; 83516 x9; 84182 x3; 86431; 86200; 86039			
New York DOH Approval Status:	This test is New York DOH approved.			
Interpretive Data:				



SSA-52 (Ro52) (ENA) Antibody, IgG	29 AU/mL or Less: Negative 30-40 AU/mL: Equivocal 41 AU/mL or greater: Positive
SSA-60 (Ro60) (ENA) Antibody, IgG	29 AU/mL or Less: Negative 30-40 AU/mL: Equivocal 41 AU/mL or greater: Positive
Scleroderma (Scl- 70) (ENA) Antibody, IgG	29 AU/mL or Less: Negative 30-40 AU/mL: Equivocal 41 AU/mL or greater: Positive
Jo-1 Antibody, IgG	29 AU/mL or Less: Negative 30-40 AU/mL: Equivocal 41 AU/mL or greater: Positive
Cyclic Citrullinated Peptide (CCP) Ab, IgG/A	19 Units or less: Negative 20-39 Units: Weak positive 40-59 Units: Moderate positive 60 Units or Greater: Strong positive
RNA Polymerase III Antibody, IgG	19 Units or less: Negative 20-39 Units: Weak positive 40-80 Units: Moderate positive 81 Units or Greater: Strong positive



Test Number	Components	Reference Interval
	Zo (phenylalanyl-tRNA synthetase) Ab	Negative
	Ks (asparaginyl-tRNA synthetase) Ab	Negative
	Ha (tyrosyl-tRNA synthetase) Ab	Negative
	Cyclic Citrullinated Peptide Ab, IgG/A	19 Units or less
	NXP2 (Nuclear matrix protein-2) Ab	Negative
	MDA5 (CADM-140) Ab	Negative
	Antinuclear Antibody (ANA), HEp-2, IgG	Less than 1:80
	SSA-60 (Ro60) (ENA) Antibody, IgG	40 AU/mL or less
	OJ (isoleucyl-tRNA synthetase) Antibody	Negative
	SRP (Signal Recognition Particle) Ab	Negative
	Ku Antibody	Negative
	EJ (glycyl-tRNA synthetase) Antibody	Negative
	PL-12 (alanyl-tRNA synthetase) Antibody	Negative
	PL-7 (threonyl-tRNA synthetase) Antibody	Negative
	PM/Scl 100 Antibody, IgG	Negative
	RNA Polymerase III Antibody, IgG	19 Units or less
	Jo-1 (Histidyl-tRNA Synthetase) Ab, IgG	40 AU/mL or less
	SSA-52 (Ro52) (ENA) Antibody, IgG	40 AU/mL or less
	Scleroderma (Scl-70) (ENA) Antibody, IgG	40 AU/mL or less
	Rheumatoid Factor	0-14 IU/mL



Dermatomyositis Autoantibody Panel			
3018870, DERM PAN2			
Specimen Requirements:			
Patient Preparation:			
Collect:	Serum separator tube (SST)		
Specimen Preparation:	Separate from cells ASAP or within 2 hours of collection. Transfer $\frac{3 \text{ to } 4}{3 \text{ to } 4}$ mL to ARUP standard transport tube. (Min: $\frac{31.5}{21.5}$ mL)		
Transport Temperature:	Refrigerated		
Unacceptable Conditions:	Hemolyzed, hyperlipemic, icteric, heat-treated, or contaminated specimens		
Remarks:			
Stability:	Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year		
Methodology:	Qualitative Immunoprecipitation / Qualitative Immunoblot / Semi-Quantitative Indirect Fluorescent Antibody (IFA) / Qualitative Particle-Based Multianalyte Technology (PMAT)		
Performed:	Sun-Sat		
Reported:	7-18 days		
Note:	Antibodies: Mi-2, P155/140, SAE1, MDA5, NXP2, TIF1-gamma, ANA		
CPT Codes:	83516 x6; 86039		
New York DOH Approval Status:	This test is New York DOH approved.		
Interpretive Data:			
Refer to report.			
Reference Interval:			



Test Number	Components	Reference Interval
	TIF-1 gamma (155 kDa) Ab	Negative
	NXP2 (Nuclear matrix protein-2) Ab	Negative
	MDA5 (CADM-140) Ab	Negative
	SAE1 (SUMO activating enzyme) Ab	Negative
	Antinuclear Antibody (ANA), HEp-2, IgG	Less than 1:80
	P155/140 Antibody	Negative
	Mi-2 (nuclear helicase protein) Antibody	Negative



NEW TEST

Click for Pricing

ThinPrep PAP Test (Standalone 3018968, TP REQUEST)
Specimen Requirements:	
Patient Preparation:	
Collect:	Cervical specimen in a ThinPrep Pap Test Collection Vial, PK/25 (ARUP Supply #51325). Cytology collection devices available: Rover Combi Brush - PK/25 (ARUP Supply #64001), Broom - PK/100 (ARUP Supply #22218), Combi Brush/Spatula - PK/25 (ARUP Supply #51326), Endocervical Brush - Each (ARUP Supply #11440). Available online through eSupply using ARUP Connect(TM) or contact ARUP Client Services at 800-522-2787. For specific instructions refer to Specimen Collection and Handling.
Specimen Preparation:	Transport cervical specimen in the original collection kit.
Transport Temperature:	Room temperature.
Unacceptable Conditions:	Specimens not collected in a ThinPrep Pap Test collection vial. Specimens submitted in an expired collection vial.
Remarks:	
Stability:	Ambient: 3 weeks; Refrigerated: 3 weeks; Frozen: Unacceptable
Methodology:	Qualitative Microscopy / Qualitative Computer Assisted Analysis
Performed:	Mon-Fri
Reported:	3-7 days
Note:	The ThinPrep 2000/5000 Systems are for use in screening for the presence of atypical cells, cervical cancer, or precursor lesions (LSIL, HSIL) as well as other cytologic categories as defined by the Bethesda System for Reporting Cervical Cytology, and is intended as a replacement for the conventional method of Pap smears. The Pap test is a screening test for cervical cancer and its precursors with an inherent false- negative rate. Store collection vials without cytologic samples at room temperature (15to 30). Do not use solution beyond expiration date marked on the vial.
CPT Codes:	88175 (88142 if manual); If reviewed by pathologist add 88141



New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Refer to report.	
Reference Interval:	



NEW TEST

Click for Pricing

ThinPrep PAP Test With Reflex to HPV if ASCUS

3018971, TA REQUEST

nen Requirements:
nen Requirements:

Patient Preparation.	Patient	Preparation:	
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Collect:	Cervical specimen in a ThinPrep Pap Test Collection Vial, PK/25 (ARUP Supply #51325). Cytology collection devices available: Rover Combi Brush - PK/25 (ARUP Supply #64001), Broom - PK/100 (ARUP Supply #22218), Combi Brush/Spatula - PK/25 (ARUP Supply #51326), Endocervical Brush - Each (ARUP Supply #11440). Available online through eSupply using ARUP Connect(TM) or contact ARUP Client Services at 800-522-2787. For specific instructions refer to Specimen Collection and Handling.
Specimen Preparation:	Transport cervical specimen in the original collection kit.
Transport Temperature:	Ambient
Unacceptable Conditions:	Specimens not collected in a ThinPrep Pap Test collection vial. Specimens submitted in an expired collection vial.
Remarks:	
Stability:	Ambient: 3 weeks; Refrigerated: 3 weeks; Frozen: Unacceptable
Methodology:	Qualitative Microscopy / Qualitative Computer Assisted Analysis
Performed:	Mon-Fri
Reported:	3-7 days
Note:	If the ThinPrep Pap test is interpreted as atypical squamous of undetermined significance (ASCUS), then Human Papillomavirus (HPV), High Risk Screen by Transcription- Mediated Amplification (TMA), with Reflex to Genotypes 16 and 18/45 (ARUP test code 3016945) will be performed and reported under a separate accession. Additional charges apply. The Pap test is a screening test for cervical cancer and its precursors with an inherent false-negative rate. Store collection vials without cytologic samples at room temperature (15to 30). Do not use solution beyond expiration date marked on the vial.



CPT Codes:	88175 (88142 if manual); If reviewed by pathologist add 88141; if reflexed, add 87624 HPV
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Refer to report.	
Reference Interval:	



NEW TEST

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ThinPrep PAP Test With Co-Test HPV

3018973, TM REQUEST

Specimen Requirements:

Patient Preparation:

Collect:	Cervical specimen in a ThinPrep Pap Test Collection Vial, PK/25 (ARUP Supply #51325). Cytology collection devices available: Rover Combi Brush - PK/25 (ARUP Supply #64001), Broom - PK/100 (ARUP Supply #22218), Combi Brush/Spatula - PK/25 (ARUP Supply #51326), Endocervical Brush - Each (ARUP Supply #11440). Available online through eSupply using ARUP Connect(TM) or contact ARUP Client Services at 800-522-2787. For specific instructions refer to Specimen Collection and Handling.
Specimen Preparation:	Transport cervical specimen in the original collection kit.
Transport Temperature:	Ambient
Unacceptable Conditions:	Specimens not collected in a ThinPrep Pap Test collection vial or specimens submitted in an expired collection vial.
Remarks:	
Stability:	Ambient: 3 weeks; Refrigerated: 3 weeks; Frozen: Unacceptable
Methodology:	Qualitative Microscopy / Qualitative Computer Assisted Analysis
Performed:	Mon-Fri
Reported:	3-7 days
Note:	In addition to the ThinPrep Pap test, Human Papillomavirus (HPV), High Risk Screen by Transcription-Mediated Amplification (TMA), With Reflex to Genotypes 16 and 18/45 (ARUP test code 3016945) will be performed and reported under a separate accession. Additional charges apply. The Pap test is a screening test for cervical cancer and its precursors with an inherent false-negative rate. Store collection vials without cytologic samples at room temperature (15to 30). Do not use solution beyond expiration date marked on the vial.
CPT Codes:	88175 (88142 if manual); If reviewed by pathologist add 88141;



	87624 HPV
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Data:	
Refer to report.	
Reference Interval:	



NEW TEST

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Hemochromatosis (HFE) 3 Vari	ants
3019007, HFEPCR	
Specimen Requirements:	
Patient Preparation:	
Collect:	Lavender (EDTA), pink (K2EDTA), or yellow (ACD solution A or B).
Specimen Preparation:	Transport 3 mL whole blood. (Min: 1 mL)
Transport Temperature:	Refrigerated
Unacceptable Conditions:	Frozen specimens in glass collection tubes.
Remarks:	
Stability:	Ambient: 72 hours; Refrigerated: 1 week; Frozen: 1 month
Methodology:	Polymerase Chain Reaction (PCR) / Fluorescence Monitoring
Performed:	Sun-Sat
Reported:	2-7 days
Note:	This test is not recommended for asymptomatic patients under 18 years of age.
CPT Codes:	81256
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	
Refer to report.	
Reference Interval:	



NEW TEST - Available Now

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Measles Virus by Qualitative NAAT

3019269, MEASLESPCR	
Specimen Requirements:	
Patient Preparation:	
Collect:	Preferred: Respiratory swab (nasal, nasopharyngeal, oropharyngeal, and throat) Also Acceptable: Urine in Aptima Urine Collection Kit.
Specimen Preparation:	Swab: Place swab in viral transport media (ARUP Supply #12884). Available online through eSupply using ARUP Connect(TM) or contact ARUP Client Services at 800-522-2787. Urine: Transfer 2 mL urine within 24 hours to an Aptima Urine Specimen Transport Tube (ARUP supply #28908 PK/50 or 54556 PK/10). Liquid level must be between fill lines on tube.
Transport Temperature:	Frozen
Unacceptable Conditions:	Swabs not in viral transport media.
Remarks:	
Stability:	Ambient: 3 days; Refrigerated: 7 days; Frozen: 30 days
Methodology:	Qualitative Nucleic Acid Amplification Test (NAAT)
Performed:	Sun-Sat
Reported:	1-3 days
Note:	
CPT Codes:	87798 x2
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.
Interpretive Deter	

Interpretive Data:

A negative result does not rule out the presence of PCR inhibitors in the patient specimen or assayspecific nucleic acid in

concentrations below the level of detection by this assay.



Test Number	Components	Reference Interval
	Measles Virus, NAAT	Not Detected
	Measles Virus Vaccine Strain, NAAT	Not Detected



NEW TEST

Click for Pricing

Medium Chain Acyl-CoA Dehydrogenase (ACADM) 2 Variants

3019336, MCAD_PCR

Specimen Requirements:	
Patient Preparation:	
Collect:	Lavender (EDTA), pink (K2EDTA), or yellow (ACD solution A or B).
Specimen Preparation:	Transport 3 mL whole blood. (Min: 1 mL)
Transport Temperature:	Refrigerated.
Unacceptable Conditions:	Frozen specimens in glass collection tubes.
Remarks:	
Stability:	Ambient: 72 hours; Refrigerated: 1 week; Frozen: 1 month
Methodology:	Polymerase Chain Reaction (PCR) / Fluorescence Monitoring
Performed:	Sun-Sat
Reported:	3-5 days
Note:	
CPT Codes:	81401
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	
Reference Interval:	

Test Components Reference Interval Number



NEW TEST

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RhD Gene (RHD) Copy Number by PCR		
3019342, RHD PCR		
Specimen Requirements:		
Patient Preparation:		
Collect:	Lavender (EDTA), pink (K2EDTA), or yellow (ACD solution A or B).	
Specimen Preparation:	Transport 3 mL whole blood. (Min: 1 mL)	
Transport Temperature:	Refrigerated.	
Unacceptable Conditions:	Frozen specimens in glass collection tubes.	
Remarks:	Patient History Form is available on the ARUP website or by contacting ARUP Client Services.	
Stability:	Ambient: 72 hours; Refrigerated: 1 week; Frozen: 1 month	
Methodology:	Polymerase Chain Reaction (PCR) / Fluorescence Monitoring	
Performed:	Varies	
Reported:	2-7 days	
Note:	Whole blood is acceptable for paternal specimens.	
CPT Codes:	81403	
New York DOH Approval Status:	This test is New York DOH approved.	
Interpretive Data:		
Refer to report.		
Reference Interval:		
By report		



NEW TEST - Available Now

Click for Pricing			
ICOS by Immunohistochemistry			
3019353, ICOS_IHC			
Specimen Requirements:			
Patient Preparation:			
Collect:	Tissue or cells.		
Specimen Preparation:	Formalin fix (10 percent neutral buffered formalin) and paraffin embed specimen (cells must be prepared into a cellblock). Protect paraffin block and/or slides from excessive heat. Transport tissue block or 5 unstained (3- to 5-micron thick sections), positively charged slides in a tissue transport kit (ARUP supply #47808). Available online through eSupply using ARUP Connect(TM) or contact ARUP Client Services at 800- 522-2787. (Min: 2 slides). If sending precut slides, do not oven bake.		
Transport Temperature:	Room temperature or refrigerated. Ship in cooled container during summer months.		
Unacceptable Conditions:	Specimens submitted with nonrepresentative tissue type. Depleted specimens.		
Remarks:	IMMUNOHISTOCHEMISTRY ORDERING AND SUBMISSION DETAILS : Submit electronic request. If you do not have electronic ordering capability, use an ARUP Immunohistochemistry Stain Form (#32978) with an ARUP client number. For additional technical details, contact ARUP Client Services at 800-522-2787.		
Stability:	Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable		
Methodology:	Immunohistochemistry (IHC)		
Performed:	Mon-Fri		
Reported:	1-3 days		
Note:	This test is performed as a stain and return (technical) service only.		
CPT Codes:	88342		



New York DOH Approval Status: This test is New York DOH approved.

Interpretive Data:

Reference Interval:

Test	Components	Reference Interval
Number		


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Ankylosing Spondylitis (HLA-B27) Genotyping 3019466, HLA-B27PCR Specimen Requirements: Patient Preparation: Collect: Lavender (EDTA), pink (K2EDTA), or yellow (ACD solution A or B). Specimen Preparation: Transport 3 mL whole blood. (Min: 1 mL) Refrigerated. Transport Temperature: Unacceptable Conditions: Plasma or serum; collection of specimen in sodium heparin tubes. Frozen specimens in glass collection tubes. Remarks: Ambient: 72 hours; Refrigerated: 1 week; Frozen: 1 month Stability: Methodology: Polymerase Chain Reaction (PCR)/Fluorescence Monitoring Performed: Sun-Sat Reported: 3-7 days Note: CPT Codes: 81374 New York DOH Approval Status: This test is New York DOH approved. Interpretive Data: Refer to report. Counseling and informed consent are recommended for genetic testing. Consent forms are available online. **Reference Interval:** By report



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THC Metabolite, with Ratio, Urine		
3019471, THC CRT		
Specimen Requirements:		
Patient Preparation:		
Collect:	Random urine.	
Specimen Preparation:	Transfer 1 mL urine with no additives or preservatives to an ARUP standard transport tube. (Min: 0.5 mL)	
Transport Temperature:	Room temperature.	
Unacceptable Conditions:	Unknown fluids, pharmaceutical preparation, and breast milk.	
Remarks:		
Stability:	Ambient: 1 week; Refrigerated: 1 month; Frozen: 1 Month	
Methodology:	Quantitative High Performance Liquid Chromatography- Tandem Mass Spectrometry	
Performed:	Sun-Sat	
Reported:	1-5 days	
Note:		
CPT Codes:	82570; 80349 (Alt code: G0480)	
New York DOH Approval Status:	Specimens from New York clients will be sent out to a New York DOH approved laboratory, if possible.	
Interpretive Data:		
Reference Interval:		
Test Components Number	Reference Interval	



NEW TEST - Available Now

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H3K36M by Immunohistochemistry

3019538, H3K36MIHC	
Specimen Requirements:	
Patient Preparation:	
Collect:	Tissue or cells.
Specimen Preparation:	Formalin fix (10 percent neutral buffered formalin) and paraffin embed specimen (cells must be prepared into a cellblock). Protect paraffin block and/or slides from excessive heat. Transport tissue block or 5 unstained (3- to 5-micron thick sections), positively charged slides in a tissue transport kit (ARUP supply #47808). Available online through eSupply using ARUP Connect(TM) or contact ARUP Client Services at 800- 522-2787. (Min: 2 slides). If sending precut slides, do not oven bake.
Transport Temperature:	Room temperature or refrigerated. Ship in cooled container during summer months.
Unacceptable Conditions:	Specimens submitted with nonrepresentative tissue type. Depleted specimens.
Remarks:	IMMUNOHISTOCHEMISTRY ORDERING AND SUBMISSION DETAILS : Submit electronic request. If you do not have electronic ordering capability, use an ARUP Immunohistochemistry Stain Form (#32978) with an ARUP client number. For additional technical details, contact ARUP Client Services at 800-522-2787.
Stability:	Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable
Methodology:	
Performed:	Mon-Fri
Reported:	1-3 days
Note:	This test is performed as a stain and return (technical) service only.
CPT Codes:	88342



New York DOH Approval Status: This test is New York DOH approved.

Interpretive Data:

Reference Interval:

Test	Components	Reference Interval
Number		



NEW TEST – Available Now

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Alpha Globin (HBA1 and HBA2) Sequencing and Deletion/Duplication, Fetal

3019566, HBA FGA FE	
Specimen Requirements:	
Patient Preparation:	
Collect:	Fetal Specimen: Two T-25 flasks at 90% confluent of cultured amniocytes or cultured chorionic villus sampling (CVS). AND Maternal Whole Blood Specimen: Lavender (EDTA), pink (K2EDTA), or yellow (ACD solution A or B).
Specimen Preparation:	Cultured Amniocytes or Cultured CVS: Fill flasks with culture media. Transport two T-25 flasks at 90 percent confluent of cultured amniocytes or cultured CVS filled with culture media. Backup cultures must be retained at the client's insitution until testing is complete. If ARUP receives a sample below the minimum confluence, Cytogenetics Grow and Send (0040182) will be added on by ARUP, and additional charges will apply. If clients are unable to culture specimens, Cytogenetics Grow and Send should be added to initial order. Maternal Whole Blood Specimen: Transport 3 mL whole blood. (Min: 1 mL).
Transport Temperature:	Cultured Amniocytes or Culture CVS: CRITICAL ROOM TEMPERATURE. Separate specimens must be submitted when mulitple tests are ordered. Must be received within 48 hours of shipment due to viability of cells. Maternal Specimen: Refrigerated
Unacceptable Conditions:	Frozen specimens
Remarks:	
Stability:	Cultured Amniocytes or Cultured CVS: Room temperature: 48 hours; Refrigerated: Unacceptable; Frozen: Unacceptable Maternal Whole Blood Specimen: Room temperature: 7 days; Refrigerated: 1 month; Frozen: Unacceptable
Methodology:	Qualitative Multiplex Ligation-Dependent Probe Amplification (MLPA)/Qualitative Sequencing
Performed:	Varies
Reported:	14-21 days
Note:	Reported times are based on receiving the two T-25 flasks at 90 percent confluent. Cell culture time is independent of



testing turnaround time. Maternal specimen is recommended for proper test interpretation. Order Maternal Cell Contamination.

By report	
Reference Interval:	
Refer to report	
Interpretive Data:	
New York DOH Approval Status:	This test is New York DOH approved.
CPT Codes:	81259; 81269; 81265



Click for Pricing	
Bicarbonate, Urine	
3019581, BICARBON	
Specimen Requirements:	
Patient Preparation:	
Collect:	Urine
Specimen Preparation:	Transfer 1 mL urine to an ARUP standard transport tube. (Min: 0.3 mL) Test is not performed at ARUP; separate specimens must be submitted when multiple tests are ordered.
Transport Temperature:	
Unacceptable Conditions:	
Remarks:	
Stability:	Ambient: 48 hours; Refrigerated: 30 days; Frozen: 1 month
Methodology:	Quantitative Enzymatic Assay
Performed:	Varies
Reported:	5-9 days
Note:	
CPT Codes:	82374
New York DOH Approval Status:	This test is New York DOH approved.
Interpretive Data:	
Reference Interval:	
Test Components	Reference Interval

Number	



<u>Click for</u>	Pricing		
Bicarbo	onate, Body Fluid		
301958	3, BICARB BF		
Specime	en Requirements:		
Patie	nt Preparation:		
Colle	ct:	Body fluid	
Spec	imen Preparation:	Transfer 1 mL bo (Min: 0.3 mL) Te specimens must	ody fluid to an ARUP standard transport tube. est is not performed at ARUP; separate be submitted when multiple tests are ordered.
Trans	sport Temperature:	Refrigerated. Als	o acceptable: Ambient or frozen
Unac	ceptable Conditions:		
Rema	arks:		
Stabi	lity:	Ambient: Undete Undetermined	rmined; Refrigerated: Undetermined; Frozen:
Method	ology:	Quantitative Enz	ymatic Assay
Perform	ed:	Varies	
Reporte	d:	8-11 days	
Note:			
CPT Coo	les:	82374	
New York DOH Approval Status:		This test is New York DOH approved.	
Interpre	tive Data:		
Referen	ce Interval:		
Test Number	Components		Reference Interval



NEW TEST – Available Now

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Antimicrobial Susceptibility - Carbapenemase Detection by CARBA5

3019585, MA CARBA5		
Specimen Requirements:		
Patient Preparation:		
Collect:	Actively growing Enterobacterales or Pseudomonas aeruginosa in pure culture.	
Specimen Preparation:	Transport sealed container with pure culture on agar slant/bacterial transport media. Place each specimen in an individually sealed bag.	
Transport Temperature:		
Unacceptable Conditions:	Specimen: FFPE and specimens in formalin. Rectal eSwab.	
Remarks:	Isolate identification (for cultures) and specimen source required.	
Stability:	Ambient: 1 week; Refrigerated: 1 week; Frozen: Unacceptable	
Methodology:	Qualitative Immunochromatography	
Performed:	Sun-Sat	
Reported:	2-3 days	
Note:	An additional processing fee will be billed for all isolates not submitted in pure culture, as indicated in the specimen requirements. If species identification is not provided, identification will be performed at ARUP. Additional charges apply.	
CPT Codes:	87185 x5	
New York DOH Approval Status:	This test is New York DOH approved.	
Interpretive Data:		
Reference Interval:		
Test Components Number	Reference Interval	





Click for Pricing

Number

Methot	rexate, Serum or Plasma	a	
301964	8, METHOTREX		
Specime	en Requirements:		
Patie	nt Preparation:		
Colle	ct:	Plain red, lavend	er (K2EDTA), or green (sodium heparin)
Spec	imen Preparation:	Transfer 1 mL se tube. (Min: 0.3 m specimens must	erum or plasma to an ARUP standard transport nL) Test is not performed at ARUP; separate be submitted when multiple tests are ordered.
Trans	sport Temperature:	Room temperatu	re. Also acceptable: Refrigerated or frozen.
Unac	ceptable Conditions:	Polymer gel sepa	aration tube (SST or PST)
Rema	arks:		
Stabi	lity:	Ambient: 2 week	s; Refrigerated: 2 weeks; Frozen: 2 weeks
Method	ology:	Liquid Chromato	graphy-Tandem Mass Spectrometry
Perform	ed:	Varies	
Reporte	d:	6-8 days	
Note:			
CPT Coo	les:	80204	
New Yo	k DOH Approval Status:	This test is New	York DOH approved.
Interpre	tive Data:		
Referen	ce Interval:		
Test	Components		Reference Interval



Click for Pricing

Lactate, Plasma			
3019650, LACTATE P			
Specimen Requirements:			
Patient Preparation:	Patient should b exercise of the a hand clenching a tourniquet if pos unclenching han buildup from the	e fasting and at complete rest and avoid any rm or hand before or during collection. Avoid and draw the specimen without the use of a sible. A tourniquet with patient clenching and d will lead to high potassium and lactic acid hand muscles, and pH will decrease.	
Collect:	Gray (potassium bath. Collect blo	oxalate/sodium fluoride) prechilled in ice od without stasis.	
Specimen Preparation:	Mix well by gent from cells within ARUP standard t performed at AR when multiple te	le inversion then return to ice bath. Separate 15 minutes and transfer 1 mL plasma to an cransport tube. (Min: 0.5 mL) Test is not UP; separate specimens must be submitted ests are ordered.	
Transport Temperature:	Refrigerated		
Unacceptable Conditions:			
Remarks:			
Stability:	Ambient: 2 week	s; Refrigerated: 2 weeks; Frozen: 2 weeks	
Methodology:	Quantitative Spe	ctrophotometry	
Performed:	Varies		
Reported:	4-6 days		
Note:			
CPT Codes:	83605		
New York DOH Approval Status:	This test is New York DOH approved.		
Interpretive Data:			
Reference Interval:			
Test Components Number		Reference Interval	





NEW TEST - Available Now

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TRBC1 by Immunohistochemistry

3019652, TRBC1_IHC	
Specimen Requirements:	
Patient Preparation:	
Collect:	Tissue or cells.
Specimen Preparation:	Formalin fix (10 percent neutral buffered formalin) and paraffin embed specimen (cells must be prepared into a cellblock).Protect paraffin block and/or slides from excessive heat. Transport tissue block or 5 unstained (3- to 5-micron thicksections), positively charged slides in a tissue transport kit (ARUP supply #47808). Available online through eSupply usingARUP Connect(TM) or contact ARUP Client Services at 800-522-2787. (Min: 2 slides). If sending precut slides, do not ovenbake.
Transport Temperature:	Room temperature or refrigerated. Ship in cooled container during summer months.
Unacceptable Conditions:	Specimens submitted with nonrepresentative tissue type. Depleted specimens.
Remarks:	IMMUNOHISTOCHEMISTRY ORDERING AND SUBMISSION DETAILS : Submit electronic request. If you do not have electronicordering capability, use an ARUP Immunohistochemistry Stain Form (#32978) with an ARUP client number. For additionaltechnical details, contact ARUP Client Services at 800-522-2787.
Stability:	Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable
Methodology:	Immunohistochemistry (IHC)
Performed:	Mon-Fri
Reported:	1-3 days
Note:	This test is performed as a stain and return (technical) service only.
CPT Codes:	88342



New York DOH Approval Status: This test is New York DOH approved.

Interpretive Data:

Reference Interval:

Test	Components	Reference Interval
Number		



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Lactate	Lactate, CSF			
3019672, LACT CSF				
Specimen Requirements:				
Patient Preparation:				
Colle	ct:	CSF		
Spec	imen Preparation:	Centrifuge to ren ARUP standard t performed at AR when multiple te	nove cellular material. Transfer 1 mL CSF to an ransport tube. (Min: 0.5 mL) Test is not UP; separate specimens must be submitted sts are ordered.	
Trans	sport Temperature:	CRITICAL FROZE	EN	
Unacceptable Conditions:				
Remarks:				
Stability:		Ambient: 3 hours; Refrigerated: 24 hours; Frozen: 2 months		
Methodology:		Quantitative Colorimetry		
Performed:		Varies		
Reported:		7-11 days		
Note:				
CPT Codes:		83605		
New York DOH Approval Status:		This test is New York DOH approved.		
Interpretive Data:				
Reference Interval:				
Test Number	Components		Reference Interval	



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UPD Glucuronosyltransferase 1A1 (UGT1A1) and Dihydropyrimidine Dehydrogenase (DPYD) Genotyping

3019841, UGT1A1DPYD

Specimen Requirements:				
Patient Preparation:				
Collect:	Two tubes of lave	ender (EDTA), pink (K2EDTA).		
Specimen Preparation:	Transport 3 mL v tube)	whole blood in each tube. (Min: 1 mL in each		
Transport Temperature:	Refrigerated.			
Unacceptable Conditions:	Heparinized spec tubes.	cimens. Frozen specimens in glass collection		
Remarks:				
Stability:	Ambient: 72 hours; Refrigerated: 1 week			
Methodology:	Polymerase Chain Reaction (PCR) / Fluorescence Monitoring / Fragment Analysis			
Performed:	Varies			
Reported:	5-10 days			
Note:				
CPT Codes:	81232; 81350			
New York DOH Approval Status:	This test is New York DOH approved.			
Interpretive Data:				
Reference Interval:				
Test Components Number		Reference Interval		



Inactivations

The following will be discontinued from ARUP's test menu on July 21, 2025 Replacement test options are indicated when applicable.

Test Number	Test Name	Refer to Replacement Test
0020045	Lactic Acid, Plasma(Change effective as of 07/21/25: Refer to 3019650 in the July Hotline)	Lactate, Plasma (3019650)
0020245	Bicarbonate (HCO[3]), Urine (Change effective as of 07/21/25: Refer to 3019581 in the July Hotline)	Bicarbonate, Urine (3019581)
0020504	Lactic Acid, Body Fluid(Inactive as of 07/21/25)	
0020516	Lactic Acid, CSF(Change effective as of 07/21/25: Refer to 3019672 in the July Hotline)	Lactate CSF (3019672)
0050392	Ankylosing Spondylitis (HLA-B27) Genotyping (Change effective as of 7/21/2025: Refer to 3019466 in the July Hotline)	Ankylosing Spondylitis (HLA-B27) Genotyping (3019466)
0051205	Medium Chain Acyl-CoA Dehydrogenase (ACADM) 2 Mutations (Change effective as of 07/21/25: Refer to 3019336 in the July Hotline)	Medium Chain Acyl-CoA Dehydrogenase (ACADM) 2 Variants (3019336)
0051368	RhD Gene (RHD) Copy Number (Change effective as of 7/21/25: Refer to 3019342 in the July Hotline)	RhD Gene (RHD) Copy number (3019342)
0055656	Hemochromatosis (HFE) 3 Mutations (Change effective as of 07/21/25: Refer to 3019007 in the July Hotline)	Hemochromatosis (HFE) 3 Variants (3019007)
0080315	Phenylalanine Monitoring, Plasma (Change effective as of 07/21/25: Refer to 0080336)	Phenylalanine and Tyrosine (0080336)
0080355	Tyrosine, Plasma (Change effective as of 07/21/25: Refer to 0080336)	Phenylalanine and Tyrosine (0080336)



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Test Number	Test Name	Refer to Replacement Test
0081105	Cystinuria Panel (Change effective as of 07/21/25: Refer to 2009419)	Amino Acids Quantitative by LC-MS/MS, Urine (2009419)
2000136	Cytology, ThinPrep Pap Test and Human Papillomavirus (HPV) High Risk Screen by Transcription-Mediated Amplification (TMA), With Reflex to Genotypes 16 and 18/45 (Change effective as of 07/21/25: Refer to 3018973 in the July Hotline)	ThinPrep PAP Test With Co-Test HPV (3018973)
2000137	Cytology, ThinPrep Pap Test (Change effective as of 07/21/25: Refer to 3018968 in the July Hotline)	ThinPrep PAP Test (Standalone) (3018968)
2000138	Cytology, ThinPrep Pap Test With Reflex to Human Papillomavirus (HPV), High Risk Screen by Transcription-Mediated Amplification (TMA), With Reflex to Genotypes 16 and 18/45 (Change effective as of 07/21/25: Refer to 3018971 in the July Hotline)	ThinPrep PAP test with reflex to HPV if ASCUS (3018971)
2005405	Methotrexate, Sensitive(Change effective as of 07/21/25: Refer to 3019648 in the July Hotline)	Methotrexate, Serum or Plasma (3019648)
3000462	Immature PLT Fraction(Inactive as of 07/21/25)	
3005425	Chimerism, Posttransplant, Sorted Cells (Monocytes) (Inactive as of 7/21/25)	
3017866	Dihydropyrimidine Dehydrogenase (DPYD) and UPD Glucuronosyltransferase 1A1 (UGT1A1) Genotyping (Change effective as of 07/21/25: Refer to 3019841 in the July Hotline)	UPD Glucuronosyltransferase 1A1 (UGT1A1) and Dihydropyrimidine Dehydrogenase (DPYD) Genotyping (3019841)